Article

Synthesis and Photophysical Properties of Charge-Transfer-Based Pyrimidine-Derived α -Amino Acids

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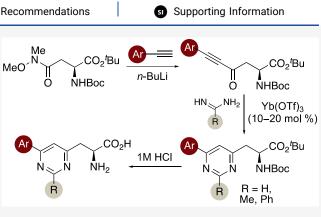


ABSTRACT: The four-step synthesis of fluorescent pyrimidinederived α -amino acids from an L-aspartic acid derivative is described. The key synthetic steps involved preparation of ynone intermediates via the reaction of alkynyl lithium salts with a Weinreb amide, followed by an ytterbium-catalyzed heterocyclization reaction with amidines. Variation of substituents at the C2and C4-position of the pyrimidine ring allowed tuning of the photoluminescent properties of the α -amino acids. This revealed that a combination of highly conjugated or electron-rich aryl substituents with the π -deficient pyrimidine motif resulted in fluorophores with the highest quantum yields and overall brightness. Further analysis of the most fluorogenic α -amino acid demonstrated solvatochromism and sensitivity to pH.

INTRODUCTION

The importance of α -amino acids as the building blocks of life along with their role in fundamental biological processes continues to drive new discoveries and applications of unnatural analogues.¹ In organic chemistry, nonproteinogenic α -amino acids are widely used in synthesis as the chiral component of ligands and auxiliaries for novel asymmetric methods, while readily available proteinogenic α -amino acids are used as chiral starting materials in total synthesis.² In medicinal chemistry and chemical biology, unnatural α -amino acids are used as enzyme inhibitors and as probes to study biological mechanisms, protein—protein interactions, and peptide conformations.³

In combination with the continued advances of fluorescence spectroscopy techniques, which allow the study of biological processes and the imaging of cellular processes,⁴ there has been significant recent interest in the development of unnatural α amino acids as fluorescent probes.⁵ This is partly due to the limitations of other approaches. The attachment of large extrinsic fluorescent labels to a protein such as green fluorescent protein⁶ or commercially available chromophores can alter structure and function. The naturally occurring proteinogenic α -amino acids, phenylalanine 1, tyrosine 2, and tryptophan 3, have poor photoluminescent properties and the presence of these at multiple sites and in different environments within a protein complicates analysis (Figure 1).^{4a} These limitations have led to the development of unnatural mimics of these α -amino acids, which are more similar in size and can be selectively embedded into peptides without altering structure.⁵ For example, 4-biphenyl-L-phenylalanine (4) was incorporated into dihydrofolate reductase to study conforma-



tional changes of inhibitor binding using Förster resonance energy-transfer (FRET) measurements.⁷ Tyrosine analogues with extended conjugation and improved photoluminescent properties such as bis-styrene **5** have been incorporated into cell-penetrating peptides and used for cell imaging.⁸ As tryptophan has the strongest fluorescent properties of the proteinogenic amino acids, many studies have focused on the modification of this α -amino acid.⁹ In particular, the reactivity of the C2-position of the indole ring has allowed the preparation of a wide range of analogues with extended conjugation (e.g., **6**).¹⁰ Several of these tryptophan mimics have been incorporated into peptides and used to image fungal infections and cancer cells.^{10c,e}

We have been interested in developing fluorescent α -amino acids with biaryl side chains as brighter, structural analogues of phenylalanine, tyrosine, and tryptophan.^{11,12} As well as the synthesis of pyrazole- and benzotriazole-derived α -amino acids, we recently reported the synthesis and photoluminescent properties of pyridine-derived α -amino acids 7 (Figure 1).¹³ These were prepared using a Lewis-acid-catalyzed hetero-Diels–Alder reaction of enone-derived α -amino acids with ethyl vinyl ether, followed by a Knoevenagel–Stobbe reaction to access the pyridine motif. Analysis of the photoluminescent

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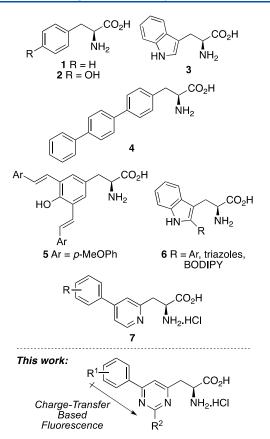
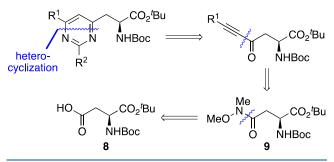


Figure 1. Fluorescent, proteinogenic α -amino acids and selected unnatural mimics.

properties of these α -amino acids revealed that a combination of electron-rich aryl substituents with the π -deficient pyridines resulted in charge-transfer-based fluorescence. Although several of the pyridine-derived α -amino acids displayed good quantum yields (0.18-0.46) and brightness, the main absorption bands were found at similar wavelengths to the fluorescent proteinogenic α -amino acids, thus restricting applications of these compounds. To overcome this limitation, we considered various structural changes that may result in fluorescent α -amino acids with red-shifted absorption and emission properties. To avoid increasing the size of the side chain resulting in α -amino acids significantly larger than proteinogenic analogues, we proposed that the incorporation of a more π -deficient heterocycle, such as a pyrimidine would enhance charge-transfer properties of the biaryl system, leading to a bathochromic shift of optical properties. Here, we report the synthesis of pyrimidine-derived α -amino acids (Figure 1) using an ytterbium-catalyzed heterocyclization reaction of ynone-derived α -amino acids with amidines as the key step. By tuning the fluorescent properties of these compounds through variation of the C2- and C4-substituents of the pyrimidine, we also demonstrate the most effective biaryl systems that generates charge-transfer-based fluorescent α -amino acids with red-shifted photoluminescent properties.

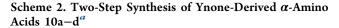
RESULTS AND DISCUSSION

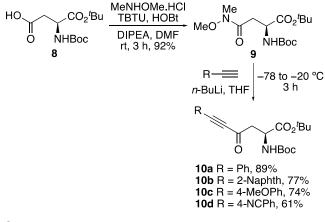
Our proposed synthesis of pyrimidine-derived α -amino acids involved three key disconnections (Scheme 1).¹⁴ Preparation of the pyrimidine ring and introduction of late-stage diversity would be achieved by heterocyclization of ynone-derived α amino acids with amidines. The ynone-derived α -amino acids Scheme 1. Proposed Synthesis of Pyrimidine-Derived α -Amino Acids



would be prepared by the chemoselective reaction of alkynyl lithium salts with Weinreb amide 9, which would also allow the incorporation of various side chains. Weinreb amide 9 would be prepared under standard conditions from commercially available *N*-Boc L-aspartic acid *t*-butyl ester 8.

The first stage of the synthetic program focused on the scalable synthesis of ynone-derived α -amino acids. Renault and co-workers previously reported an efficient route to these compounds and thus, with some modifications, this was used for the preparation of ynones **10a**-**10d** (Scheme 2).¹⁵ Initially,





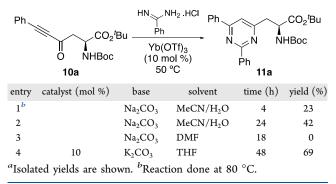
^{*a*}Isolated yields are shown.

commercially available N-Boc L-aspartic acid *t*-butyl ester **8** was treated with *N*,*O*-dimethyl hydroxylamine hydrochloride in the presence of TBTU and HOBt, which gave Weinreb amide **9** in 92% yield. The ynone-derived α -amino acids were then prepared by the reaction of Weinreb amide **9** with alkynyl lithium salts, generated by the deprotonation of aryl-substituted alkynes with *n*-butyl lithium. This gave ynones **10a–10d** cleanly, in 61–89% yield. Although the majority of ynones prepared possessed conjugated or electron-rich aryl side chains, which would lead to charge transfer with the π -deficient pyrimidine, an electron-deficient analogue bearing a 4-cyanophenyl group (**10d**) was also synthesized to examine the interaction of this with the pyrimidine ring.

In 1946, Bowden and Jones reported the synthesis of a pyrimidine by the reaction of an ynone with guanidine under basic conditions.¹⁶ More recently, similar procedures have been used for the synthesis of pyrimidines from ynones,¹⁷ including a study by Baldwin and co-workers that reported the synthesis of pyrimidine-derived α -amino acids from L-aspartic

acid and L-glutamic acid derivatives.¹⁸ The majority of these procedures involved reaction of the ynone with an amidine by heating under reflux in the presence of sodium carbonate. Our initial studies investigated the reaction of phenyl-substituted ynone **10a** with benzamidine hydrochloride under similar conditions (Table 1, entry 1). Although this gave pyrimidine

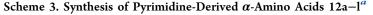
Table 1. Optimization Studies for Pyrimidine Formation from Ynone $10a^{a}$

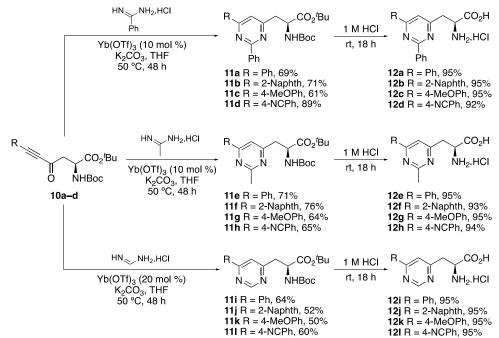


11a cleanly, the compound was isolated in only 23% yield. Attempts were then made to modify this approach. In addition, we wanted to avoid the combination of basic conditions and high temperatures (80 °C) and so a reaction at 50 °C and with a longer reaction time (24 h) was investigated (entry 2). This gave pyrimidine 11a in an improved 42% yield. To avoid the use of water as a cosolvent, DMF was then investigated under the same conditions, but this gave no reaction (entry 3). In their work, Baldwin and co-workers highlighted that reaction of ynones with formamidine under these conditions gave low yields of the corresponding pyrimidine. This was a concern as we believed that pyrimidine-derived α -amino acids with no C2-substituent were likely to produce the most effective

fluorophores. For this reason and the modest yields already observed for base-mediated heterocyclizations, we considered a different approach for the preparation of the pyrimidines. Bagley and co-workers showed that ynones could be activated with ytterbium salts for reaction with enamines and the subsequent synthesis of pyridine derivatives.¹⁹ Based on this, the use of ytterbium triflate as a Lewis acid catalyst for pyrimidine synthesis was investigated. An initial attempt involved the reaction of phenyl-substituted ynone **10a** with benzamidine hydrochloride in the presence of ytterbium triflate (10 mol %) (entry 4). Using THF as the solvent and potassium carbonate to neutralize the benzamidine salt, this generated pyrimidine **11a** cleanly and with an isolated yield of 69%.

The scope of the ytterbium triflate catalyzed heterocyclization reaction of ynone-derived α -amino acids 10a-10d with benzamidine, acetamidine, and formamidine was then explored (Scheme 3). The reaction of ynones 10a-10d with benzamidine and acetamidine under the Lewis-acid-catalyzed conditions was found to be highly effective and gave the pyrimidine products in 61-89% yields.²⁰ For the more challenging heterocyclization reaction with formamidine, the optimized conditions gave low yields. For example, reaction of p-methoxyphenyl-substituted ynone 10c with formamidine hydrochloride and using ytterbium triflate (10 mol %) gave the corresponding pyrimidine 11k in 19% yield. However, it was found that the use of higher catalyst loading (20 mol %) resulted in an improved 50% yield. This modification was also effective with the other ynones, allowing the synthesis of pyrimidines 11i-11l in 50-64% yields. Having successfully synthesized a small library of pyrimidine-derived α -amino acids, the protecting groups were removed in the presence of 1 M hydrochloric acid under mild conditions, which gave parent amino acids 12a-12l in excellent yields.





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amino acid	$\lambda_{\rm Abs} \ ({\rm nm})^a$	$\varepsilon ~({\rm cm^{-1}~M^{-1}})$	$\lambda_{\rm Em} \ ({\rm nm})^a$	Stokes shift (cm ⁻¹)	$\Phi_{\mathrm{F}}{}^{b}$	brightness (cm ⁻¹ M ⁻¹)
12b	311	24,000	497	12,033	0.12	2880
12c	305	12,700	314	940	0.003	38
12f	305	12,800	490	12,379	0.11	1408
12g	299	16,400	314, 381	1598, 7198	0.016	262
12j	310	10,400	421	8505	0.30	3120
12k	306	13,600	384	6638	0.27	3672
^{<i>a</i>} Spectra were recorded at 2 μ M in methanol. ^{<i>b</i>} Quantum yields (Φ_F) were determined in methanol using anthracene and L-tryptophan as standards.						

Table 2. Photophysical Data of Pyrimidine-Derived α -Amino Acids

On synthesis of the pyrimidine-derived α -amino acids, the photoluminescent properties were measured for each compound. The ultraviolet-visible (UV-vis) absorption and photoluminescence spectra of the α -amino acids were recorded in methanol at a concentration of 2 μ M. As expected, pyrimidines with weakly donating (Ph) or electron-withdrawing (4-NCPh) C4-substituents displayed weak fluorescence and low brightness (see the Supporting Information). The most interesting properties were found for pyrimidines with highly conjugated (naphthyl) and strongly electrondonating (4-MeOPh) C4-substituents (Table 2). These α amino acids exhibited red-shifted absorption bands in comparison to proteinogenic α -amino acids 1-3 and the previously reported pyridine analogues,¹³ with fluorescence in the visible region. For the C4-naphthyl compounds (12b, 12f, and 12j), all three compounds showed absorption bands between 305 and 311 nm, possessed megaStokes shifts, and good quantum yields, resulting in the brightest series of α amino acids (Table 2 and Figure 2). The main difference in this series was observed in the fluorescence spectra, in which α amino acids 12b and 12f showed emission maxima between 490 and 500 nm, while a hypsochromic shift in emission to 421 nm was observed for C2-unsubstituted analogue 12j (Figure 2b). For 12j, we believe that the lack of a C2-substituent allows emission from a more planar locally excited state, while α amino acids 12b and 12f, which have more distorted conformations due to C2-substituents, emit from twisted intramolecular charge-transfer excited states. For the pmethoxyphenyl series, the trend of emission maxima was found to be reversed. The C2-substituted compounds 12c and 12g displayed weak emission maxima at 314 nm, while 12k with no C2-substituent showed a bathochromic shift in emission to 384 nm (Figure 3b). In this series, the C2substituent obviously has a strong influence on the interaction between the C4-aryl group and the pyrimidine ring. While the C2-phenyl and methyl groups disrupt this interaction, no substituent at C2 allows strong interplay between the electronrich *p*-methoxyphenyl ring and the π -deficient pyrimidine heterocycle, resulting in strong intramolecular charge-transfer emission. As well as strong fluorescence, α -amino acid 12k displayed red-shifted absorption compared to the corresponding pyridine (283 nm),¹³ a large Stokes shift, as well as a good quantum yield (0.27) and brightness. Overall, these results provide insight into the relationship between the structure and photoluminescence properties of these α -amino acids and, in particular, the use of substituents to control biaryl conformation, leading to emission from either locally excited or twisted/planar intramolecular charge-transfer excited states.

Although the naphthyl series of α -amino acids gave strong, red-shifted emission and good quantum yields, the *p*methoxyphenyl pyrimidine-derived α -amino acid 12k was found to be the brightest. For this reason, the properties of 12k

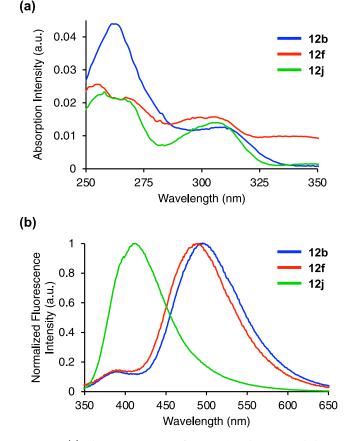


Figure 2. (a) Absorption spectra of **12b**, **12f**, and **12j**, recorded at 2 μ M in methanol. (b) Emission spectra of **12b**, **12f**, and **12j**, recorded at 2 μ M in methanol.

were further explored via solvatochromic and pH studies. Analysis of α -amino acid 12k in a range of solvents produced similar absorption spectra, indicating that in the ground state, the absorbance is independent of solvent polarity (see the Supporting Information). In contrast, a bathochromic shift in emission maxima was observed with increasing solvent polarity (Figure 4a). For example, the emission maximum was found at 352 nm in ethyl acetate, while this shifted to 384 nm in water. The solvatochromism displayed by α -amino acid 12k confirms the intramolecular charge-transfer character of the excited state, which is stabilized in more polar solvents. To determine the effect of pyrimidine ring protonation on the photophysical properties of α -amino acid 12k, pH studies were conducted. A change of pH from 7 to 1, resulted in stronger absorbance around 300 nm and the formation of a second minor band at longer wavelength (~360 nm) (see the Supporting Information). A significant change was also observed for the emission properties of α -amino acid 12k (Figure 4b). While the position

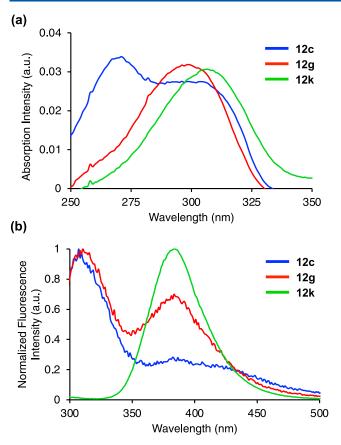


Figure 3. (a) Absorption spectra of **12c**, **12g**, and **12k**, recorded at 2 μ M in methanol. (b) Emission spectra of **12c**, **12g**, and **12k**, recorded at 2 μ M in methanol.

of the emission band is unchanged, the fluorescence is "turned off" following acidification. Protonation of substituted pyrimidines typically leads to a change in conformation, resulting in either a hypsochromic or bathochromic shift of emission bands.²¹ For α -amino acid **12k**, formation of a positively charged pyrimidine ring results in effective fluorescence quenching. These results suggest that α -amino acid **12k** may have potential as a fluorescent probe for biological applications, which involve a change of polarity or pH conditions.

CONCLUSIONS

In conclusion, a small library of α -amino acids with pyrimidine side chains were prepared using a novel ytterbium-catalyzed heterocyclization reaction of ynone-derived α -amino acids with amidines as the key step. The pyrimidine heterocycle was selected as a strong π -deficient motif, which with conjugated or electron-rich aryl substituents would result in strong fluorescence. Analysis of the optical properties of these compounds proved this to be the case. Pyrimidines with no C2-substituent and with highly conjugating (12j) or electrondonating C4-substituents (12k) displayed red-shifted absorption bands (compared to the corresponding pyridine analogue),¹³ strong fluorescence emission in the visible region, large Stokes shifts, and good quantum yields (0.27-0.30). The intermolecular charge-transfer properties of the brightest α amino acid 12k were confirmed with a solvatochromic study, which showed a bathochromic shift of emission in more polar solvents. α -Amino acid 12k was also found to be highly

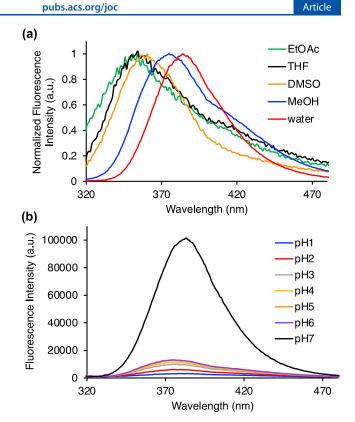


Figure 4. (a) Normalized fluorescence spectra of 12k in various solvents. (b) Emission spectra of 12k at various pH in methanol. All spectra were recorded using a concentration of 5 μ M.

sensitive to pH, with protonation of the pyrimidine ring resulting in fluorescence quenching. This study has generated further insight into the relationship between structure, conformation, and photoluminescent properties of biarylderived α -amino acids such as **12k** that have the potential to act as fluorescent probes. Current work is investigating this potential for biological applications.

EXPERIMENTAL SECTION

All reagents and starting materials were obtained from commercial sources and used as received. Reactions were performed open to air unless otherwise mentioned. All reactions performed at elevated temperatures were heated using an oil bath. Brine refers to a saturated aqueous solution of sodium chloride. Flash column chromatography was performed using silica gel 60 (40-63 μ m). Aluminum-backed plates precoated with silica gel 60F254 were used for thin-layer chromatography and were visualized with a UV lamp or by staining with potassium permanganate, vanillin, or ninhydrin. ¹H NMR spectra were recorded on a Bruker DPX 400 MHz spectrometer and data are reported as follows: chemical shift in ppm relative to the solvent as an internal standard (CHCl₃, δ 7.26 ppm; CH₃OH, δ 3.31 ppm), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet or overlap of nonequivalent resonances, integration). The abbreviation br s refers to broad singlet. ¹³C NMR spectra were recorded on an NMR spectrometer at 101 MHz and data are reported as follows: chemical shift in ppm relative to tetramethylsilane or the solvent as an internal standard (CDCl₃, δ 77.2 ppm; CD₃OD, δ 49.0 ppm). Infrared spectra were recorded using a Shimadzu IR Prestige-21 spectrometer; wavenumbers are indicated in cm⁻¹. Mass spectra were recorded using electrospray techniques. HRMS spectra were recorded using Bruker micrOTOF-Q or Agilent 6546 LC/Q-TOF mass spectrometers. Melting points are uncorrected. Optical rotations were determined as solutions irradiating with the sodium D line (λ = 589 nm) using an Autopol V polarimeter. $[\alpha]_D$ values are given in

units 10^{-1} deg cm⁻¹ g⁻¹. UV–vis spectra were recorded on a PerkinElmer Lambda 25 instrument. Fluorescence spectra were recorded on a Shimadzu RF-5301PC spectrofluorophotometer. Absorbance spectra were recorded with an integration time of 0.05 s and a band pass of 5 nm. Fluorescence spectra were recorded with excitation and emission band pass of 10 nm, an integration time of 0.1 or 2 s, and with detector accumulations set to 1. Quantum yield data were measured using anthracene and L-tryptophan as standard references.

tert-Butyl (2S)-(tert-Butoxycarbonylamino)-4-[methoxy-(methyl)amino]-4-oxobutanoate (9).¹⁵ A mixture of 1-tert-butyl (2S)-(tert-butoxycarbonylamino)-4-butan-1,4-dioic acid (8) (0.50 g, 1.76 mmol), N,O-dimethylhydroxyamine hydrochloride (0.24 g, 2.44 mmol), and O-(1H-benzotriazol-1-yl)-N,N,N',N'-tetramethyluronium tetrafluoroborate (0.73 g, 1.93 mmol) was stirred in N,N'dimethylformamide (3 mL) at 0 °C. Diisopropylethylamine (0.75 mL, 4.40 mmol) was then added dropwise. The mixture was stirred at room temperature for 3 h. An aqueous solution of 1 M sodium hydrogen sulfate (25 mL) was then added, and the mixture was extracted with ethyl acetate $(3 \times 20 \text{ mL})$. The combined organic layers were then washed with brine $(3 \times 20 \text{ mL})$, dried (MgSO₄), and concentrated under vacuum. Purification by flash column chromatography eluting with 20% ethyl acetate in dichloromethane gave tertbutyl (2S)-(tert-butoxycarbonylamino)-4-[methoxy(methyl)amino]-4-oxobutanoate (9) as a colorless oil (0.59 g, 92%). $[\alpha]_{D}^{20}$ -14.5 (c 0.6, MeOH) [lit.¹⁵ $[\alpha]_D^{25}$ -12.3 (c 1.0, MeOH)]; ¹H NMR (CDCl₃, 400 MHz): δ 5.67 (d, J = 8.3 Hz, 1H), 4.47–4.43 (m, 1H), 3.18–3.12 (m, 4H), 3.69 (s, 3H), 2.87 (dd, J = 16.8, 2.7 Hz, 1H), 1.46 (s, 9H),1.44 (s, 9H); ${}^{13}C{}^{1}H$ NMR (CDCl₃, 101 MHz): δ 171.8, 170.5, 155.7, 81.7, 79.5, 61.2, 50.4, 34.7, 32.0, 28.3, 27.9; MS (ESI) m/z 355 $(M + Na^{+}, 100).$

tert-Butyl (25)-(*tert*-Butoxycarbonylamino)-4-oxo-6-phenyl-hex-5-ynoate (10a).¹⁵ To a solution of phenylacetylene (0.20 mL, 1.9 mmol) in tetrahydrofuran (10 mL) at -78 °C was slowly added a solution of n-butyl lithium (2.5 M in hexane, 0.43 mL, 1.1 mmol). The mixture was stirred for 0.75 h and then added dropwise to a solution of tert-butyl (2S)-(tert-butoxycarbonylamino)-4-[methoxy-(methyl)amino]-4-oxobutanoate (9) (0.12 g, 0.36 mmol) in tetrahydrofuran (25 mL) at -78 °C. The reaction mixture was stirred at -78 °C for 1 h and then at -20 °C for 2 h. An aqueous 1 M solution of dipotassium hydrogen phosphate (10 mL) was added, and the mixture was extracted with diethyl ether $(3 \times 30 \text{ mL})$. The combined organic layers were washed with brine $(4 \times 30 \text{ mL})$ and dried (MgSO₄). After filtration, the solution was concentrated under vacuum. Purification by column chromatography eluting with 20% diethyl ether in petroleum gave tert-butyl (2S)-(tert-butoxycarbonylamino)-4-oxo-6-phenylhex-5-ynoate (10a) as a yellow oil (0.13 g, 89%). $[\alpha]_{D}^{20}$ -7.5 (c 0.5, MeOH) [lit.¹⁵ $[\alpha]_{D}^{25}$ -3.8 (c 1.0, MeOH)]; ¹H NMR (CDCl₃, 400 MHz): δ 7.62–7.55 (m, 2H), 7.50–7.43 (m, 1H), 7.42-7.36 (m, 2H), 5.44 (d, J = 8.1 Hz, 1H), 4.53-4.49 (m, 1H), 3.33 (dd, J = 17.9, 4.6 Hz, 1H), 3.18 (dd, J = 17.9, 4.6 Hz, 1H), 1.46 (s, 9H), 1.45 (s, 9H); ${}^{13}C{}^{1}H$ NMR (CDCl₃, 101 MHz): δ 184.7, 169.8, 155.5, 133.2, 131.0, 128.7, 119.6, 92.0, 87.4, 82.5, 79.9, 50.2, 47.6, 28.3, 27.9; MS (ESI) m/z 396 (M + Na⁺, 100).

tert-Butyl (2S)-(tert-Butoxycarbonylamino)-4-oxo-6-(2'naphthyl)hex-5-ynoate (10b). The reaction was carried out according to the above procedure for the synthesis of 10a using 2ethylnylnaphthalene (0.97 g, 6.4 mmol), n-butyl lithium (2.5 M in hexane, 1.5 mL, 3.8 mmol), and tert-butyl (2S)-(tert-butoxycarbonylamino)-4-[methoxy (methyl)amino]-4-oxobutanoate (9) (0.42 g, 1.3 mmol). Purification by flash column chromatography eluting with 20% diethyl ether in petroleum gave tert-butyl (2S)-(tert-butoxycarbonylamino)-4-oxo-6-(2'-naphthyl)hex-5-ynoate (10b) as a colorless oil (0.41 g, 77%). IR (neat) 3342, 2977, 2197, 1708, 1667, 1496, 1366, 1215, 1080, 746 cm⁻¹; $[\alpha]_D^{24}$ +9.0 (*c* 0.1, CHCl₃); ¹H NMR (CDCl₃, 400 MHz): δ 8.15 (s, 1H), 7.85–7.80 (m, 3H), 7.58–7.50 (m, 3H), 5.49 (d, J = 8.6 Hz, 1H), 4.56 (dt, J = 8.6, 4.6 Hz, 1H), 3.37 (dd, J = 17.9, 4.6 Hz, 1H), 3.23 (dd, J = 17.9, 4.6 Hz, 1H), 1.48 (s, 9H), 1.46 (s, 9H); ${}^{13}C{}^{1}H$ NMR (CDCl₃, 101 MHz): δ 184.7, 169.9, 155.5, 134.6, 134.0, 132.6, 128.5, 128.4, 128.2, 128.1,

127.9, 127.1, 116.8, 92.5, 87.7, 82.5, 79.9, 50.2, 47.6, 28.4, 27.9; MS (ESI) m/z 446 (M + Na⁺, 100); HRMS (ESI) m/z: [M + Na]⁺ Calcd for C₂₅H₂₀NO₅Na 446.1938; found 446.1948.

tert-Butyl (2S)-(tert-Butoxycarbonylamino)-4-oxo-6-(4'methoxyphenyl)hex-5-ynoate (10c). The reaction was carried out according to the above procedure for the synthesis of 10a using 4methoxyphenylacetylene (0.70 mL, 5.40 mmol), n-butyl lithium (2.5 M in hexane, 1.30 mL, 3.24 mmol), and tert-butyl (2S)-(tertbutoxycarbonylamino)-4-[methoxy(methyl)amino]-4-oxobutanoate (9) (0.35 g, 1.1 mmol). Purification by flash column chromatography eluting with 30% diethyl ether in petroleum gave tert-butyl (2S)-(tertbutoxycarbonylamino)-4-oxo-6-(4'-methoxyphenyl)hex-5-ynoate (10c) as a white solid (0.32 g, 74%). Mp 118–120 $^\circ\text{C};$ IR (neat) 3376, 2978, 2361, 2193, 1710, 1665, 1509, 1367, 1253, 1153, 1086, 739 cm⁻¹; $[\alpha]_{D}^{21}$ +16.4 (*c* 0.1, CHCl₃); ¹H NMR (CDCl₃, 400 MHz): δ 7.57–7.51 (m, 2H), 6.92–6.87 (m, 2H), 5.43 (d, J = 8.5 Hz, 1H), 4.50 (dt, J = 8.5, 4.5 Hz, 1H), 3.85 (s, 3H), 3.31 (dd, J = 17.8, 4.5 Hz, 1H), 3.16 (dd, J = 17.8, 4.5 Hz, 1H), 1.46 (s, 9H), 1.44 (s, 9H); $^{13}\text{C}\{^{1}\text{H}\}$ NMR (CDCl₃, 101 MHz): δ 184.6, 169.9, 161.9, 155.5, 135.3, 114.4, 111.4, 93.3, 87.5, 82.4, 79.8, 55.4, 50.2, 47.4, 28.3, 27.9; MS (ESI) m/z 426 (M + Na⁺, 100); HRMS (ESI) m/z: [M + Na]⁺ Calcd for C222H29NO6Na 426.1887; found 426.1881.

tert-Butyl (2S)-(tert-Butoxycarbonylamino)-4-oxo-6-(4'cyanophenyl)hex-5-ynoate (10d). The reaction was carried out according to the above procedure for the synthesis of 10a using 4cyanophenylacetylene (0.21 g, 1.7 mmol), n-butyl lithium (2.5 M in hexane, 0.40 mL, 1.0 mmol), and tert-butyl (2S)-(tert-butoxycarbonylamino)-4-[methoxy(methyl)amino]-4-oxobutanoate (9) (0.12 g, 0.33 mmol). Purification by flash column chromatography eluting with 30% diethyl ether in petroleum gave tert-butyl (2S)-(tertbutoxycarbonylamino)-4-oxo-6-(4'-cyanophenyl)hex-5-ynoate (10d) as a white solid (0.10 g, 61%). Mp 149-151 °C; IR (neat) 3391, 2978, 2366, 2207, 1710, 1678, 1500, 1367, 1252, 1150, 1086, 844, 736 cm⁻¹; $[\alpha]_{D}^{25}$ +9.2 (c 0.1, CHCl₃); ¹H NMR (CDCl₃, 400 MHz): δ 7.71–7.66 (m, 4H), 5.39 (d, J = 8.2 Hz, 1H), 4.53 (dt, J = 8.2, 4.8 Hz, 1H), 3.31 (dd, J = 17.8, 4.8 Hz, 1H), 3.20 (dd, J = 17.8, 4.8 Hz, 1H), 1.46 (s, 9H), 1.45 (s, 9H); ${}^{13}C{}^{1}H$ NMR (CDCl₃, 101 MHz): δ 184.2, 169.6, 155.4, 133.4, 132.3, 124.4, 117.8, 114.3, 89.6, 88.4, 82.8, 80.1, 50.2, 47.6, 28.3, 27.9; MS (ESI) m/z 421 (M + Na⁺, 100); HRMS (ESI) m/z: $[M + Na]^+$ Calcd for $C_{22}H_{26}N_2O_5Na$ 421.1734; found 421.1727.

tert-Butyl (2S)-2-(tert-Butoxycarbonylamino)-3-(2'-phenyl-4'-phenylpyrimidin-6'-yl)propanoate (11a). tert-Butyl (2S)-(tert-butoxycarbonylamino)-4-oxo-6-phenylhex-5-ynoate (10a) (0.046 g, 0.11 mmol) was dissolved in tetrahydrofuran (2 mL), followed by sequential addition of benzamidine hydrochloride (0.026 g, 0.17 mmol), potassium carbonate (0.018 g, 0.13 mmol), and ytterbium triflate (0.0070 g, 0.011 mmol). The mixture was heated to 50 °C for 48 h and then concentrated in vacuo. The residue was redissolved in dichloromethane (10 mL) and washed with a saturated solution of sodium hydrogen carbonate (5 mL) and then brine (5 mL). The organic layer was dried (MgSO₄) and concentrated in vacuo. Purification by flash column chromatography eluting with 30% ethyl acetate in petroleum ether gave tert-butyl (2S)-2-(tertbutoxycarbonylamino)-3-(2'-phenyl-4'-phenylpyrimidin-6'-yl)propanoate (11a) (0.039 g, 69%) as a yellow solid. Mp 95-100 °C; IR (neat) 3429, 2977, 1706, 1583, 1501, 1364, 1250, 1149, 1031, 837, 729 cm⁻¹; $[\alpha]_D^{18}$ +26.0 (*c* 0.1, CHCl₃); ¹H NMR (CDCl₃, 400 MHz): δ 8.61-8.57 (m, 2H), 8.23-8.19 (m, 2H), 7.53-7.49 (m, 6H), 7.47 (s, 1H), 5.94 (d, J = 8.6 Hz, 1H), 4.73 (dt, J = 8.6, 5.1 Hz, 1H), 3.49 (dd, J = 15.7, 5.1 Hz, 1H), 3.36 (dd, J = 15.7, 5.1 Hz, 1H), 1.44 (s, 9H), 1.34 (s, 9H); ¹³C{¹H} NMR (CDCl₃, 101 MHz): δ 170.7, 166.9, 164.03, 163.97, 155.6, 137.7, 137.0, 130.9, 130.7, 128.9, 128.52, 128.46, 127.2, 114.2, 81.9, 79.7, 52.5, 39.3, 28.4, 28.0; MS (ESI) m/z 498 (M + Na⁺, 100); HRMS (ESI) m/z: [M + Na]⁺ Calcd for C28H33N3O4Na 498.2363; found 498.2355.

tert-Butyl (2S)-2-(*tert*-Butoxycarbonylamino)-3-[4'-(2"naphthyl)-2'-phenylpyrimidin-6'-yl]propanoate (11b). *tert*-Butyl (2S)-2-(*tert*-butoxycarbonylamino)-3-[4'-(2"-naphthyl)-2'-phenylpyrimidin-6'-yl]propanoate (11b) was synthesized as described for

11a using tert-butyl (2S)-(tert-butoxycarbonylamino)-4-oxo-6-(2'naphthyl) hex-5-ynoate (10b) (0.076 g, 0.18 mmol), benzamidine hydrochloride (0.042 g, 0.27 mmol), potassium carbonate (0.030 g, 0.22 mmol), and ytterbium triflate (0.010 g, 0.018 mmol). Purification by flash column chromatography eluting with 30% ethyl acetate in petroleum ether gave tert-butyl (2S)-2-(tert-butoxycarbonylamino)-3-[4'-(2"-naphthyl)-2'-phenylpyrimidin-6'-yl]propanoate (11b) (0.067 g, 71%) as a white solid. Mp 130-133 °C; IR (neat) 3366, 2978, 1713, 1571, 1537, 1495, 1367, 1153, 1058, 760, 698 cm⁻¹; $[\alpha]_{D}^{24}$ +10.0 (c 0.1, CHCl₃); ¹H NMR (CDCl₃, 400 MHz): δ 8.72 (br s, 1H), 8.66–8.62 (m, 2H), 8.33 (dd, J = 8.6, 1.7 Hz, 1H), 8.03-7.90 (m, 3H), 7.63 (s, 1H), 7.58-7.50 (m, 5H), 5.95 (d, J = 8.6 Hz, 1H), 4.75 (dt, J = 8.6, 5.2 Hz, 1H), 3.53 (dd, J = 15.7, 5.2 Hz, 1H), 3.40 (dd, J = 15.7, 5.2 Hz, 1H), 1.44 (s, 9H), 1.35 (s, 9H); $^{13}C{^{1}H}$ NMR (CDCl₃, 101 MHz): δ 170.7, 163.9, 137.7, 134.7, 134.3, 133.3, 130.8, 129.0, 128.7, 128.6, 128.5, 127.8, 127.5, 127.4, 126.6, 124.1, 116.8, 114.4, 82.0, 79.7, 52.5, 39.4, 28.4, 28.0; MS (ESI) m/z 548 (M + Na⁺, 100); HRMS (ESI) m/z: [M + Na]⁺ Calcd for C32H35N3O4Na 548.2520; found 548.2520.

tert-Butyl (2S)-2-(tert-Butoxycarbonylamino)-3-[4'-(4"-methoxyphenyl)-2'-phenylpyrimidin-6'-yl]propanoate (11c). tert-Butyl (2*S*)-2-(*tert*-butoxycarbonylamino)-3-[4'-(4"-methoxyphenyl)-2'-phenylpyrimidin-6'-yl]propanoate (11c) was synthesized as described for 11a using tert-butyl (2S)-(tert-butoxycarbonylamino)-4-oxo-6-(4'-methoxyphenyl)hex-5-ynoate (10c) (0.44 g, 1.2 mmol), benzamidine hydrochloride (0.29 g, 1.8 mmol), potassium carbonate (0.20 g, 1.5 mmol), and ytterbium triflate (0.0070 g, 0.012 mmol). Purification by flash column chromatography eluting with 30% ethyl acetate in petroleum ether gave tert-butyl (2S)-2-(tert-butoxycarbonylamino)-3-[4'-(4"-methoxyphenyl)-2'-phenylpyrimidin-6'-yl]propanoate (11c) (0.31 g, 61%) as a white solid. Mp 205-210 °C; IR (neat) 3376, 2982, 1706, 1573, 1533, 1368, 1217, 1172, 763 cm⁻¹; $[\alpha]_{D}^{20}$ +24.5 (c 0.1, CHCl₃); ¹H NMR (CDCl₃, 400 MHz): δ 8.59– 8.57 (m, 2H), 8.20-8.18 (m, 2H), 7.54-7.48 (m, 3H), 7.40 (s, 1H), 7.04-7.02 (m, 2H), 5.98 (d, J = 8.1 Hz, 1H), 4.72-4.68 (m, 1H), 3.89 (s, 3H), 3.46 (dd, J = 15.6, 4.7 Hz, 1H), 3.33 (dd, J = 15.6, 4.5 Hz, 1H), 1.44 (s, 9H), 1.34 (s, 9H); ${}^{13}C{}^{1}H$ NMR (CDCl₃, 101 MHz): δ 170.7, 166.5, 163.9, 163.5, 162.0, 155.6, 137.8, 130.6, 129.4, 128.8, 128.5, 128.4, 114.3, 113.3, 81.9, 79.6, 55.4, 52.5, 39.2, 28.4, 28.0; MS (ESI) m/z 506 (M + H⁺, 100); HRMS (ESI) m/z: [M + H]⁺ Calcd for C₂₉H₃₅N₃O₅H 506.2649; found 506.2647.

tert-Butyl (2S)-2-(tert-Butoxycarbonylamino)-3-[4'-(4"-cyanophenyl)-2'-phenylpyrimidin-6'-yl]propanoate (11d). tert-Butyl (2S)-2-(tert-butoxycarbonylamino)-3-[4'-(4"-cyanophenyl)-2'phenylpyrimidin-6'-yl]propanoate (11d) was synthesized as described for 11a using tert-butyl (2S)-(tert-butoxycarbonylamino)-4-oxo-6-(4'cyanophenyl)hex-5-ynoate (10d) (0.10 g, 0.25 mmol), benzamidine hydrochloride (0.059 g, 0.37 mmol), potassium carbonate (0.041 g, 0.30 mmol), and ytterbium triflate (0.015 g, 0.025 mmol). Purification by flash column chromatography eluting with 30% ethyl acetate in petroleum ether gave tert-butyl (2S)-2-(tert-butoxycarbonylamino)-3-[4'-(4"-cyanophenyl)-2'-phenylpyrimidin-6'-yl]propanoate (11d) (0.11 g, 89%) as a yellow oil. IR (neat) 3425, 2978, 2229, 1705, 1569, 1533, 1367, 1150, 732, 695 cm⁻¹; $\left[\alpha\right]_{\rm D}^{24}$ +30.2 (c 0.1, CHCl₃); ¹H NMR (CDCl₃, 400 MHz): δ 8.60-8.55 (m, 2H), 8.32 (d, J = 8.4 Hz, 2H), 7.83 (d, J = 8.4 Hz, 2H), 7.55–7.51 (m, 4H), 5.80 (d, J = 8.4 Hz, 1H), 4.74 (dt, J = 8.4, 5.2 Hz, 1H), 3.51 (dd, J = 15.9, 5.2 Hz, 1H), 3.41 (dd, J = 15.9, 5.2 Hz, 1H), 1.43 (s, 9H), 1.36 (s, 9H); ${}^{13}C{}^{1}H$ NMR (CDCl₃, 101 MHz): δ 170.6, 167.8, 164.4, 161.8, 155.5, 141.2, 137.2, 132.7, 131.1, 128.6, 128.5, 127.8, 118.4, 114.6, 114.3, 82.1, 79.9, 52.3, 39.5, 28.3, 28.0; MS (ESI) m/z523 (M + Na⁺, 100); HRMS (ESI) m/z: [M + Na]⁺ Calcd for C₂₉H₃₂N₄O₄Na 523.2316; found 523.2316.

tert-Butyl (2S)-2-(tert-Butoxycarbonylamino)-[3-(2'-methyl-4'-phenylpyrimidin-6'-yl)](propanoate) (11e). tert-Butyl (2S)-(tert-butoxycarbonylamino)-4-oxo-6-phenylhex-5-ynoate (10a) (0.34 g, 0.91 mmol) was dissolved in tetrahydrofuran (30 mL), followed by sequential addition of acetamidine hydrochloride (0.13 g, 1.4 mmol), potassium carbonate (0.30 g, 2.2 mmol), and ytterbium triflate (0.056 g, 0.091 mmol). The mixture was heated to 50 °C for 48 h and was

concentrated in vacuo. The residue was redissolved in dichloromethane (30 mL), washed with a saturated solution of sodium hydrogen carbonate (20 mL), brine (20 mL), dried (MgSO₄), and concentrated in vacuo. Purification by flash column chromatography eluting with 30% ethyl acetate in petroleum ether gave tert-butyl (2S)-2-(tert-butoxycarbonylamino)-[3-(2'-methyl-4'-phenylpyrimidin-6'yl)(propanoate)] (11e) (0.27 g, 71%) as a white solid. Mp 98–103 °C; IR (neat) 3665, 2979, 1705, 1580, 1541, 1366, 1149, 1055, 752, 693 cm⁻¹; $[\alpha]_{D}^{25}$ +16.4 (c 0.1, CHCl₃); ¹H NMR (CDCl₃, 400 MHz): δ 8.07–8.03 (m, 2H), 7.51–7.47 (m, 3H), 7.41 (s, 1H), 5.65 (d, J = 8.1 Hz, 1H), 4.67–4.62 (m, 1H), 3.33 (dd, J = 15.1, 5.7 Hz, 1H), 3.27 (dd, J = 15.1, 5.2 Hz, 1H), 2.76 (s, 3H), 1.42 (s, 9H), 1.40 (s, 9H); $^{13}C{^{1}H}$ NMR (CDCl₃, 101 MHz): δ 170.4, 167.7, 166.4, 164.3, 155.4, 136.9, 130.8, 128.9, 127.2, 113.8, 82.0, 79.7, 52.8, 39.4, 28.3, 27.9, 26.0; MS (ESI) m/z 436 (M + Na⁺, 100); HRMS (ESI) m/z: $[M + Na]^+$ Calcd for $C_{23}H_{31}N_3O_4Na$ 436.2207; found 436.2208.

tert-Butyl (2S)-2-(*tert*-Butoxycarbonylamino)-[3-(2'-methyl-4'-phenylpyrimidin-6'-yl)](propanoate) (11e)—Gram-Scale Reaction.²⁰ The reaction was performed as described above using *tert*-butyl (2S)-(*tert*-butoxycarbonylamino)-4-oxo-6-phenylhex-5ynoate (10a) (1.0 g, 2.7 mmol), acetamidine hydrochloride (0.39 g, 4.2 mmol), potassium carbonate (0.90 g, 6.6 mmol), and ytterbium triflate (0.17 g, 0.27 mmol). Purification by flash column chromatography eluting with 30% ethyl acetate in petroleum ether gave *tert*-butyl (2S)-2-(*tert*-butoxycarbonylamino)-[3-(2'-methyl-4'phenylpyrimidin-6'-yl)(propanoate)] (11e) (0.80 g, 69%) as a white solid. Spectroscopic data are as described above.

tert-Butyl (25)-2-(tert-Butoxycarbonylamino)-3-[2'-methyl-4'-(2"-naphthyl)pyrimidin-6'-yl]propanoate (11f). tert-Butyl (2S)-2-(tert-butoxycarbonylamino)-3-[2'-methyl-4'-(2"-naphthyl)pyrimidin-6'-yl]propanoate (11f) was synthesized as described for 11e using tert-butyl (2S)-(tert-butoxycarbonylamino)-4-oxo-6-(2'naphthyl)hex-5-ynoate (10b) (0.17 g, 0.39 mmol), acetamidine hydrochloride (0.092 g, 0.59 mmol), potassium carbonate (0.13 g, 0.94 mmol), and ytterbium triflate (0.024 g, 0.039 mmol). Purification by flash column chromatography eluting with 30% ethyl acetate in petroleum ether gave tert-butyl (2S)-2-(tert-butoxycarbonylamino)-3-[2'-methyl-4'-(2"-naphthyl)pyrimidin-6'-yl]propanoate (11f) (0.14 g, 76%) as a colorless oil. IR (neat) 3361, 2980, 1716, 1587, 1541, 1364, 1254, 1160, 1024, 758 cm⁻¹; $[\alpha]_{D}^{20}$ +20.0 (c 0.1, CHCl₃); ¹H NMR (CDCl₃, 400 MHz): δ 8.61 (br s, 1H), 8.16 (d, J = 8.5 Hz, 1H), 7.98-7.87 (m, 3H), 7.61-7.52 (m, 3H), 5.69 (d, J = 8.0 Hz, 1H), 4.71-4.65 (m, 1H), 3.43-3.31 (m, 2H), 2.81 (s, 3H), 1.42 (br s, 18H); ${}^{13}C{}^{1}H$ NMR (CDCl₃, 101 MHz): δ 170.4, 167.5, 166.2, 164.3, 155.5, 134.6, 134.0, 133.2, 129.1, 128.8, 127.8, 127.5, 126.6, 124.0, 114.0, 82.1, 79.8, 52.8, 39.3, 28.3, 27.9, 25.9; MS (ESI) m/z 464 (M + H⁺, 100); HRMS (ESI) m/z: [M + H]⁺ Calcd for C₂₇H₃₃N₃O₄H 464.2544; found 464.2555.

tert-Butyl (2S)-2-(tert-Butoxycarbonylamino)-3-[4'-(4"-methoxyphenyl)-2'-methylpyrimidin-6'-yl]propanoate (11g). tert-Butyl (2S)-2-(tert-butoxycarbonylamino)-3-[4'-(4"-methoxyphenyl)-2'-methylpyrimidin-6'-yl]propanoate (11g) was synthesized as described for 11e using tert-butyl (2S)-(tert-butoxycarbonylamino)-4-oxo-6-(4'-methoxyphenyl)hex-5-ynoate (10c) (0.093 g, 0.23 mmol), acetamidine hydrochloride (0.033 g, 0.34 mmol), potassium carbonate (0.076 g, 0.55 mmol), and ytterbium triflate (0.014 g, 0.023 mmol). Purification by flash column chromatography eluting with 30% ethyl acetate in petroleum ether gave tert-butyl (2S)-2-(tertbutoxycarbonylamino)-3-[4'-(4"-methoxyphenyl)-2'-methylpyrimidin-6'-yl]propanoate (11g) (0.065 g, 64%) as a white solid. Mp 110-114 °C; IR (neat) 3372, 2976, 1708, 1584, 1516, 1370, 1254, 1159, 840, 731 cm⁻¹; $[\alpha]_{D}^{19}$ +15.0 (c 0.1, CHCl₃); ¹H NMR (CDCl₃, 400 MHz): δ 8.03 (d, J = 8.8 Hz, 2H), 7.32 (s, 1H), 7.01–6.97 (m, 2H), 5.65 (d, J = 8.1 Hz, 1H), 4.62 (dt, J = 8.1, 5.5 Hz, 1H), 3.87 (s, 3H), 3.29 (dd, J = 15.0, 5.5 Hz, 1H), 3.21 (dd, J = 15.0, 5.5 Hz, 1H), 2.71 (s, 3H), 1.42 (s, 9H), 1.39 (s, 9H); ¹³C{¹H} NMR (CDCl₃, 101 MHz): δ 170.5, 167.7, 166.2, 163.6, 161.9, 155.4, 129.4, 128.7, 114.3, 112.8, 81.9, 79.7, 55.4, 52.8, 39.5, 28.3, 27.9, 26.2; MS (ESI) m/z 444 $(M + H^{+}, 100);$ HRMS (ESI) $m/z: [M + H]^{+}$ Calcd for C₂₄H₃₃N₃O₅H 444.2493; found 444.2497.

tert-Butyl (2S)-2-(tert-Butoxycarbonylamino)-3-[4'-(4"-cyanophenyl)-2'-methylpyrimidin-6'-yl]propanoate (11h). tert-Butyl (2S)-2-(tert-butoxycarbonylamino)-3-[4'-(4"-cyanophenyl)-2'methylpyrimidin-6'-yl]propanoate (11h) was synthesized as described for 11e using tert-butyl (2S)-(tert-butoxycarbonylamino)-4oxo-6-(4'-cyanophenyl)hex-5-ynoate (10d) (0.084 g, 0.21 mmol), acetamidine hydrochloride (0.030 g, 0.32 mmol), potassium carbonate (0.070 g, 0.50 mmol), and ytterbium triflate (0.013 g, 0.021 mmol). Purification by flash column chromatography eluting with 30% ethyl acetate in petroleum ether gave tert-butyl (2S)-2-(tertbutoxycarbonylamino)-3-[4'-(4"-cyanophenyl)-2'-methylpyrimidin-6'-yl]propanoate (11h) (0.065 g, 65%) as a colorless oil. IR (neat) 3393, 2885, 2237, 1744, 1592, 1501, 1450, 1247, 1033, 661 cm⁻¹ $[\alpha]_{D}^{24}$ +30.0 (c 0.1, CHCl₃); ¹H NMR (CDCl₃, 400 MHz): δ 8.18 (d, *J* = 8.0 Hz, 2H), 7.81–7.77 (m, 2H), 7.44 (s, 1H), 5.54 (d, *J* = 8.1 Hz, 1H), 4.67-4.62 (m, 1H), 3.37-3.26 (m, 2H), 2.76 (s, 3H), 1.41 (br s, 18H); ${}^{13}C{}^{1}H$ NMR (CDCl₃, 101 MHz): δ 170.3, 168.3, 167.4, 161.9, 155.4, 141.2, 132.6, 127.8, 118.4, 114.1, 82.2, 79.8, 52.7, 39.7, 28.3, 27.9, 26.1; MS (ESI) m/z 439 (M + H⁺, 100); HRMS (ESI) m/ *z*: $[M + H]^+$ Calcd for C₂₄H₃₀N₄O₄H 439.2340; found 439.2341.

tert-Butyl (2S)-2-(tert-Butoxycarbonylamino)-3-(4'-phenylpyrimidin-6'-yl)propanoate (11i). tert-Butyl (2S)-(tert-butoxycarbonylamino)-4-oxo-6-phenylhex-5-ynoate (10a) (0.083 g, 0.22 mmol) was dissolved in tetrahydrofuran (15 mL), followed by sequential addition of formamidine hydrochloride (0.18 g, 2.20 mmol), potassium carbonate (0.61 g, 4.40 mmol), and ytterbium triflate (0.027 g, 0.044 mmol). The mixture was heated to 50 °C for 48 h and was concentrated in vacuo. The residue was redissolved in dichloromethane (20 mL), washed with a saturated solution of sodium hydrogen carbonate (20 mL), brine (20 mL), dried (MgSO₄), and concentrated in vacuo. Purification by flash column chromatography eluting with 30% ethyl acetate in petroleum ether gave tert-butyl (2S)-2-(tert-butoxycarbonylamino)-3-(4'-phenylpyrimidin-6'-yl)propanoate (11i) (0.053 g, 64%) as a colorless oil. IR (neat) 3668, 2977, 1713, 1587, 1512, 1368, 1254, 1156, 1024, 840, 747 cm⁻¹; $[\alpha]_D^{15}$ +40.0 (c 0.1, CHCl₃); ¹H NMR (CDCl₃, 400 MHz): δ 9.15 (s, 1H), 8.09-8.05 (m, 2H), 7.60 (s, 1H), 7.53-7.49 (m, 3H), 5.69 (d, J = 8.5 Hz, 1H), 4.69–4.64 (m, 1H), 3.36 (dd, J = 15.2, 5.7 Hz, 1H), 3.29 $(dd, J = 15.2, 5.0 \text{ Hz}, 1\text{H}), 1.42 (s, 9\text{H}), 1.39 (s, 9\text{H}); {}^{13}\text{C}{}^{1}\text{H}$ NMR (CDCl₂, 101 MHz): δ 170.4, 166.8, 163.9, 158.5, 155.4, 136.6, 131.0, 129.0, 127.1, 116.8, 82.2, 79.8, 52.8, 39.7, 28.3, 27.9; MS (ESI) m/z 400 (M + H⁺, 100): HRMS (ESI) m/z: [M + H]⁺ Calcd for C₂₂H₂₉N₃O₄H 400.2231; found 400.2229.

tert-Butyl (2S)-2-(tert-Butoxycarbonylamino)-3-[4'-(2"naphthyl)pyrimidin-6'-yl]propanoate (11j). tert-Butyl (2S)-2-(tert-butoxycarbonylamino)-3-[4'-(2"-naphthyl)pyrimidin-6'-yl]propanoate (11j) was synthesized as described for 11i using tert-butyl (2S)-(tert-butoxycarbonylamino)-4-oxo-6-(2'-naphthyl)hex-5-ynoate (10b) (0.050 g, 0.12 mmol), formamidine hydrochloride (0.097 g, 1.20 mmol), potassium carbonate (0.33 g, 2.40 mmol), and ytterbium triflate (0.015 g, 0.024 mmol). Purification by flash column chromatography eluting with 30% ethyl acetate in petroleum ether gave tert-butyl (2S)-2-(tert-butoxycarbonylamino)-3-[4'-(2"naphthyl)pyrimidin-6'-yl]propanoate (11j) (0.028 g, 52%) as a white solid. Mp 162-165 °C; IR (neat) 3430, 2979, 1715, 1613, 1497, 1366, 1152, 752, 694 cm⁻¹; $[\alpha]_D^{19}$ +34.0 (c 0.1, CHCl₃); ¹H NMR (CDCl₃, 400 MHz): δ 9.20 (s, 1H), 8.61 (br s, 1H), 8.15 (dd, J = 8.6, 1.5 Hz, 1H), 7.99-7.95 (m, 2H), 7.92-7.87 (m, 1H), 7.75 (s, 1H), 7.59–7.53 (m, 2H), 5.72 (d, J = 8.2 Hz, 1H), 4.72–4.67 (m, 1H), 3.40 (dd, J = 15.1, 5.9 Hz, 1H), 3.33 (dd, J = 15.1, 5.1 Hz, 1H), 1.43 (s, 9H), 1.40 (s, 9H); ${}^{13}C{}^{1}H$ NMR (CDCl₃, 101 MHz): δ 170.5, 166.9, 163.8, 158.5, 155.5, 134.6, 133.7, 133.3, 129.0, 128.84, 128.75, 127.5, 126.7, 123.8, 116.9, 82.2, 79.9, 52.8, 39.7, 28.3, 27.9; MS (ESI) m/z 448 ([M – H]⁻, 100); HRMS (ESI) m/z: [M – H]⁻ Calcd for C₂₆H₃₀N₃O₄ 448.2242; found 448.2230.

tert-Butyl (25)-2-(tert-Butoxycarbonylamino)-3-[4'-(4"methoxyphenyl)pyrimidin-6'-yl]propanoate (11k). tert-Butyl (2S)-2-(tert-butoxycarbonylamino)-3-[4'-(4"-methoxyphenyl)pyrimidin-6'-yl]propanoate (11k) was synthesized as described for 11i using tert-butyl (2S)-(tert-butoxycarbonylamino)-4-oxo-6-(4'- methoxyphenyl)hex-5-ynoate (10c) (0.099 g, 0.25 mmol), formamidine hydrochloride (0.200 g, 2.50 mmol), potassium carbonate (0.69 g, 5.00 mmol), and ytterbium triflate (0.031 g, 0.050 mmol). Purification by flash column chromatography eluting with 30% ethyl acetate in petroleum ether gave tert-butyl (2S)-2-(tert-butoxycarbonylamino)-3-[4'-(4"-methoxyphenyl)pyrimidin-6'-yl]propanoate (11k) (0.054 g, 50%) as a colorless oil. IR (neat) 3368, 2979, 1714, 1592, 1529, 1365, 1254, 1150, 1025 cm⁻¹; $[\alpha]_{\rm D}^{18}$ +19.0 (c 0.1, CHCl₃); ¹H NMR (CDCl₃, 400 MHz): δ 9.08 (s, 1H), 8.07–8.03 (m, 2H), 7.53 (s, 1H), 7.03–7.00 (m, 2H), 5.71 (d, J = 8.2 Hz, 1H), 4.65 (dt, J = 8.2, 5.6 Hz, 1H), 3.88 (s, 3H), 3.33 (dd, J = 15.1, 5.6 Hz, 1H), 3.25 (dd, J = 15.1, 5.6 Hz, 1H), 1.42 (s, 9H), 1.38 (s, 9H); ¹³C{¹H} NMR (CDCl₃, 101 MHz): δ 170.5, 166.4, 163.4, 162.1, 158.4, 155.5, 128.9, 128.7, 115.8, 114.4, 82.1, 79.8, 55.4, 52.8, 39.6, 28.3, 27.9; MS (ESI) m/z 430 (M + H⁺, 100); HRMS (ESI) m/z: [M + H]⁺ Calcd for C₂₃H₃₁N₃O₅H 430.2336; found 430.2339

tert-Butyl (2S)-2-(tert-Butoxycarbonylamino)-3-[4'-(4"cyanophenyl)pyrimidin-6'-yl]propanoate (11l). tert-Butyl (2S)-2-(tert-butoxycarbonylamino)-3-[4'-(4"-cyanophenyl)pyrimidin-6'yl]propanoate (111) was synthesized as described for 11i using tertbutyl (2S)-(tert-butoxycarbonylamino)-4-oxo-6-(4'-cyanophenyl)hex-5-ynoate (10d) (0.053 g, 0.13 mmol), formamidine hydrochloride (0.024 g, 1.30 mmol), potassium carbonate (0.036 g, 2.60 mmol), and ytterbium triflate (0.016 g, 0.026 mmol). Purification by flash column chromatography eluting with 30% ethyl acetate in petroleum ether gave tert-butyl (2S)-2-(tert-butoxycarbonylamino)-3-[4'-(4"cyanophenyl)pyrimidin-6'-yl]propanoate (111) (0.032 g, 60%) as a colorless oil. IR (neat) 3436, 2979, 2230, 1710, 1588, 1499, 1367, 1253, 1153, 1024, 903, 725 cm⁻¹; $[\alpha]_{D}^{20}$ +20.0 (c 0.1, CHCl₃); ¹H NMR (CDCl₃, 400 MHz): δ 9.19 (s, 1H), 8.20 (d, J = 8.5 Hz, 2H), 7.82-7.79 (m, 2H), 7.65 (s, 1H), 5.61 (d, J = 8.0 Hz, 1H), 4.70-4.65 (m, 1H), 3.41–3.31 (m, 2H), 1.41 (s, 9H), 1.40 (s, 9H); ¹³C{¹H} NMR (CDCl₃, 101 MHz): δ 170.3, 167.7, 161.7, 158.7, 155.4, 140.7, 132.8, 127.7, 118.3, 117.2, 114.5, 82.4, 79.9, 52.7, 39.9, 28.3, 27.9; MS (ESI) m/z 425 (M + H⁺, 100); HRMS (ESI) m/z: [M + H]⁺ Calcd for C₂₃H₂₈N₄O₄H 425.2183; found 425.2183.

(25)-2-Amino-3-(2'-phenyl-4'-phenylpyrimidin-6'-yl)-propanoic Acid Hydrochloride (12a). *tert*-Butyl (2S)-2-(*tert*butoxycarbonylamino)-3-(2'-phenyl-4'-phenylpyrimidin-6'-yl)propanoate (11a) (0.11 g, 0.23 mmol) was dissolved in acetonitrile (1 mL). To the reaction mixture was added 1 M hydrochloric acid (10 mL) and the mixture was stirred at room temperature for 18 h. The reaction mixture was then concentrated in vacuo. Purification by recrystallization from diethyl ether (5 mL) gave (2S)-2-amino-3-(2'phenyl-4'-phenylpyrimidin-6'-yl)propanoic acid hydrochloride (12a) as a white solid (0.077 g, 95%). Mp 195-200 °C; IR (neat) 3357, 2932, 2158, 1717, 1574, 1537, 1377, 1029, 749, 688 cm⁻¹; $[\alpha]_{D}^{20}$ +19.8 (c 0.1, MeOH); ¹H NMR (CD₃OD, 400 MHz): δ 8.60–8.53 (m, 2H), 8.34-8.28 (m, 2H), 7.84 (s, 1H), 7.60-7.49 (m, 6H), 4.65 (t, J = 5.4 Hz, 1H), 3.63 (d, J = 5.4 Hz, 2H); ¹³C{¹H} NMR (CD₃OD, 101 MHz): δ 169.9, 164.9, 164.6, 164.2, 137.4, 136.5, 131.0, 130.7, 128.7, 128.2, 127.0, 114.1, 50.7, 35.8; MS (ESI) m/z342 (M + Na⁺, 100); HRMS (ESI) m/z: [M + Na]⁺ Calcd for C19H17N3O2Na 342.1213; found 342.1212.

(2S)-2-Amino-3-[4'-(2"-naphthyl)-2'-phenylpyrimidin-6'-yl]propanoic Acid Hydrochloride (12b). (2S)-2-Amino-3-[4'-(2"naphthyl)-2'-phenylpyrimidin-6'-yl]propanoic acid hydrochloride (12b) was prepared as described for 12a using tert-butyl (2S)-2-(tert-butoxycarbonylamino)-3-[4'-(2"-naphthyl)-2'-phenylpyrimidin-6'-yl]propanoate (11b) (0.11 g, 0.23 mmol). Following completion, the reaction mixture was concentrated in vacuo. This gave (2S)-2amino-3-[4'-(2"-naphthyl)-2'-phenylpyrimidin-6'-yl]propanoic acid hydrochloride (12b) as a colorless oil (0.077 g, 95%). IR (neat) 3305, 2826, 1746, 1571, 1537, 1377, 1082, 760, 697 cm⁻¹; $[\alpha]_{\rm D}^{24}$ +19.1 (c 0.1, MeOH); ¹H NMR (CD₃OD, 400 MHz): δ 8.86 (br s, 1H), 8.64–8.59 (m, 2H), 8.42 (dd, J = 8.0, 3.6 Hz, 1H), 8.08–8.00 (m, 3H), 7.96–7.93 (m, 1H), 7.62–7.52 (m, 5H), 4.67 (t, J = 5.4 Hz, 1H), 3.67 (d, J = 5.4 Hz, 2H); ${}^{13}C{}^{1}H$ NMR (CD₃OD, 101 MHz): δ 169.8, 164.8, 164.7, 164.0, 137.1, 135.0, 133.6, 133.3, 130.9, 128.8, 128.4, 128.33, 128.26, 127.6, 127.44, 127.42, 126.5, 123.7, 114.5, 50.7,

35.8; MS (ESI) m/z 370 (M + H⁺, 100); HRMS (ESI) m/z: [M + H]⁺ Calcd for C₂₃H₁₉N₃O₂H 370.1550; found 370.1558.

(2S)-2-Amino-3-[4'-(4"-methoxyphenyl)-2'-phenylpyrimidin-6'-yl]propanoic Acid Hydrochloride (12c). (2S)-2-Amino-3-[4'-(4"-methoxyphenyl)-2'-phenylpyrimidin-6'-yl]propanoic acid hydrochloride (12c) was prepared as described for 12a using tert-butyl (2S)-2-(tert-butoxycarbonylamino)-3-[4'-(4"-methoxyphenyl)-2'phenylpyrimidin-6'-yl]propanoate (11c) (0.124 g, 0.280 mmol). This gave (2S)-2-amino-3-[4'-(4"-methoxyphenyl)-2'-phenylpyrimidin-6'yl]propanoic acid hydrochloride (12c) as a white solid (0.0764 g, 95%). Mp 175-180 °C; IR (neat) 3354, 2835, 1745, 1588, 1631, 1563, 1428, 1254, 1176, 1081, 840, 758 cm⁻¹; $[\alpha]_{\rm D}^{18}$ +22.4 (c 0.1, MeOH); ¹H NMR (CD₃OD, 400 MHz): δ 8.53–8.50 (m, 2H), 8.34 (d, J = 8.6 Hz, 2H), 7.97 (s, 1H), 7.62–7.55 (m, 3H), 7.13 (d, J = 8.6 Hz, 2H), 4.71 (t, J = 5.6 Hz, 1H), 3.91 (s, 3H), 3.70 (d, J = 5.6 Hz, 2H); ¹³C{¹H} NMR (CD₃OD, 101 MHz): δ 169.4, 165.4, 163.7, 163.4, 162.1, 134.6, 131.9, 129.9, 128.6, 127.1, 114.4, 114.3, 54.9, 50.8, 35.1; MS (ESI) *m*/*z* 350 (M + H⁺, 100); HRMS (ESI) *m*/*z*: [M + H]⁺ Calcd for $C_{20}H_{19}N_3O_3H$ 350.1499; found 350.1500.

(2S)-2-Amino-3-[4'-(4"-cyanophenyl)-2'-phenylpyrimidin-6'-yl]propanoic Acid Hydrochloride (12d). (2S)-2-Amino-3-[4'-(4"-cyanophenyl)-2'-phenylpyrimidin-6'-yl]propanoic acid hydrochloride (12d) was prepared as described for 12a using tert-butyl (2S)-2-(tert-butoxycarbonylamino)-3-[4'-(4"-cyanophenyl)-2'-phenylpyrimidin-6'-yl]propanoate (11d) (0.088 g, 0.18 mmol). Following completion, the reaction mixture was concentrated in vacuo. This gave (2S)-2-amino-3-[4'-(4"-cyanophenyl)-2'-phenylpyrimidin-6'yl]propanoic acid hydrochloride (12d) as a yellow oil (0.057 g, 92%). IR (neat) 3354, 2936, 1712, 1646, 1588, 1516, 1366, 1251, 1173, 770 cm⁻¹; $[\alpha]_{D}^{22}$ +15.0 (*c* 0.1, MeOH); ¹H NMR (CD₃OD, 400 MHz): δ 8.58–8.54 (m, 2H), 8.49 (d, J = 7.9 Hz, 2H), 7.96 (s, 1H), 7.92 (d, J = 7.9 Hz, 2H), 7.55–7.50 (m, 3H), 4.69 (t, J = 4.6 Hz, 1H), 3.69 (d, J = 4.6 Hz, 2H); ¹³C{¹H} NMR (CD₃OD, 101 MHz): δ 169.8, 165.7, 164.4, 162.6, 140.8, 137.0, 132.6, 130.9, 128.3, 128.2, 127.9, 117.9, 115.0, 114.2, 50.6, 36.0; MS (ESI) m/z 345 (M + H⁺, 100); HRMS (ESI) m/z: $[M + H]^+$ Calcd for $C_{20}H_{16}N_4O_2H$ 345.1346; found 345.1349.

(2S)-2-Amino-3-(2'-methyl-4'-phenylpyrimidin-6'-yl)propanoic Acid Hydrochloride (12e). (2S)-2-Amino-3-(2'-methyl-4'-phenylpyrimidin-6'-yl)propanoic acid hydrochloride (12e) was prepared as described for 12a using tert-butyl (2S)-2-(tertbutoxycarbonylamino)-3-(2'-methyl-4'-phenylpyrimidin-6'-yl)propanoate (11e) (0.22 g, 0.53 mmol). This gave (2S)-2-amino-3-(2'-methyl-4'-phenylpyrimidin-6'-yl)propanoic acid hydrochloride (12e) as a white solid (0.13 g, 95%). Mp 145-149 °C; IR (neat) 3371, 2918, 1745, 1612, 1593, 1505, 1440, 1368, 1223, 1055, 752, 695 cm⁻¹; $[\alpha]_D^{24}$ +17.6 (c 0.1, MeOH); ¹H NMR (CD₃OD, 400 MHz): δ 8.36 (s, 1H), 8.26-8.22 (m, 2H), 7.75-7.62 (m, 3H), 4.81-4.75 (m, 1H), 3.80 (dd, J = 16.4, 6.1 Hz, 1H), 3.72 (dd, J = 16.4, 7.1 Hz, 1H), 2.98 (s, 3H); ¹³C{¹H} NMR (CD₃OD, 101 MHz): δ 168.9, 166.7, 164.61, 164.55, 133.2, 132.4, 129.3, 128.5, 116.6, 50.6, 35.4, 22.0; MS (ESI) *m*/*z* 258 (M + H⁺, 100); HRMS (ESI) *m*/*z*: [M $+ H^{+}$ Calcd for C₁₄H₁₅N₂O₂H 258.1237; found 258.1238.

(2S)-2-Amino-3-[2'-methyl-4'-(2"-naphthyl)pyrimidin-6'yl]propanoic Acid Hydrochloride (12f). (2S)-2-Amino-3-[2'methyl-4'-(2"-naphthyl)pyrimidin-6'-yl]propanoic acid hydrochloride (12f) was prepared as described for 12a using tert-butyl (2S)-2-(tertbutoxycarbonylamino)-3-[2'-methyl-4'-(2"-naphthyl)pyrimidin-6'yl]propanoate (11f) (0.0650 g, 0.14 mmol). Following completion, the reaction mixture was concentrated in vacuo. This gave (2S)-2amino-3-[2'-methyl-4'-(2"-naphthyl)pyrimidin-6'-yl]propanoic acid hydrochloride (12f) as a colorless oil (0.040 g, 93%). IR (neat) 3354, 2952, 1724, 1512, 1347, 1214, 1056, 812, 743 cm⁻¹; $[\alpha]_{\rm D}^{22}$ +18.0 (c 0.1, MeOH); ¹H NMR (CD₃OD, 400 MHz): δ 8.87 (s, 1H), 8.54 (s, 1H), 8.21 (d, J = 8.9 Hz, 1H), 8.10-8.03 (m, 2H), 7.94 (d, J = 8.9 Hz, 1H), 7.67-7.56 (m, 2H), 4.82-4.79 (m, 1H), 3.82 (dd, J = 16.5, 4.9 Hz, 1H), 3.78 (dd, I = 16.5, 6.9 Hz, 1H), 2.97 (s, 3H); ¹³C{¹H} NMR (CD₃OD, 101 MHz): δ 168.7, 168.0, 165.4, 163.7, 135.8, 132.9, 130.9, 129.4, 129.3, 129.2, 128.9, 127.6, 127.3, 123.8, 117.2, 50.7, 35.0, 21.5; MS (ESI) m/z 308 (M + H⁺, 100); HRMS

(ESI) m/z: $[M + H]^+$ Calcd for $C_{18}H_{17}N_3O_2H$ 308.1394; found 308.1399.

(2S)-2-Amino-3-[4'-(4"-methoxyphenyl)-2'-methylpyrimidin-6'-yl]propanoic Acid Hydrochloride (12g). (2S)-2-Amino-3-[4'-(4"-methoxyphenyl)-2'-methylpyrimidin-6'-yl]propanoic acid hydrochloride (12g) was prepared as described for 12a using *tert*-butyl (2S)-2-(tert-butoxycarbonylamino)-3-[4'-(4"-methoxyphenyl)-2'methylpyrimidin-6'-yl]propanoate (11g) (0.12 g, 0.28 mmol). This gave (2S)-2-amino-3-[4'-(4"-methoxyphenyl)-2'-methylpyrimidin-6'yl]propanoic acid hydrochloride (12g) as a white solid (0.077 g, 95%). Mp 174-180 °C; IR (neat) 3262, 2830, 1742, 1584, 1251, 1180, 1031, 894, 621 cm⁻¹; $[\alpha]_D^{18}$ +13.0 (c 0.1, MeOH); ¹H NMR (CD₃OD, 400 MHz): δ 8.23 (dd, J = 7.9, 2.1 Hz, 2H), 8.16 (s, 1H), 7.17 (dd, J = 7.9, 2.1 Hz, 2H), 4.70 (t, J = 6.7 Hz, 1H), 3.93 (s, 3H), 3.68 (dd, J = 16.2, 6.7 Hz, 1H), 3.60 (dd, J = 16.2, 6.7 Hz, 1H), 2.90 (s, 3H); ${}^{13}C{}^{1}H$ NMR (CD₃OD, 101 MHz): δ 168.8, 164.9, 163.83, 163.82, 130.8, 124.0, 115.4, 114.9, 55.0, 50.7, 35.1, 21.7; MS (ESI) m/z 288 (M + H⁺, 100); HRMS (ESI) m/z: [M + H]⁺ Calcd for C15H17N3O3H 288.1343; found 288.1349.

(2S)-2-Amino-3-[4'-(4"-cyanophenyl)-2'-methylpyrimidin-6'-yl]propanoic Acid Hydrochloride (12h). (2S)-2-Amino-3-[4'-(4"-cyanophenyl)-2'-methylpyrimidin-6'-yl]propanoic acid hydrochloride (12h) was prepared as described for 12a using tert-butyl (2S)-2-(tert-butoxycarbonylamino)-3-[4'-(4"-cyanophenyl)-2'-methylpyrimidin-6'-yl]propanoate (11h) (0.065 g, 0.15 mmol). Following completion, the reaction mixture was concentrated in vacuo. This gave (2S)-2-amino-3-[4'-(4"-cyanophenyl)-2'-methylpyrimidin-6'-yl]propanoic acid (12h) as a yellow oil (0.040 g, 94%). IR (neat) 3325, 2922, 1641, 1592, 1526, 1490, 1362, 1011 cm⁻¹; $[\alpha]_D^{25}$ +14.0 (c 0.1, MeOH); ¹H NMR (CD₃OD, 400 MHz): δ 8.36–8.32 (m, 2H), 7.94–7.89 (m, 3H), 4.61 (dd, J = 7.4, 4.7 Hz, 1H), 3.60 (dd, J = 16.7, 4.7 Hz, 1H), 3.51 (dd, J = 16.7, 7.4 Hz, 1H), 2.79 (s, 3H); ${}^{13}C{}^{1}H{}$ NMR (CD₃OD, 101 MHz): δ 169.4, 167.8, 165.7, 162.9, 140.2, 132.5, 128.0, 117.8, 114.6, 114.5, 50.8, 35.5, 24.1; MS (ESI) m/z 283 $(M + H^+, 100)$; HRMS (ESI) m/z: $[M + H]^+$ Calcd for C15H14N4O2H 283.1190; found 283.1189.

(25)-2-Amino-3-(4'-phenylpyrimidin-6'-yl)propanoic Acid Hydrochloride (12i). (2S)-2-Amino-3-(4'-phenylpyrimidin-6'-yl)propanoic acid hydrochloride (12i) was prepared as described for 12a using *tert*-butyl (2S)-2-(*tert*-butoxycarbonylamino)-3-(4'-phenylpyrimidin-6'-yl)propanoate (11i) (0.12 g, 0.31 mmol). This gave (2S)-2-amino-3-(4'-phenylpyrimidin-6'-yl)propanoic acid hydrochloride (12i) as a white solid (0.073 g, 95%). Mp 68–70 °C; IR (neat) 3440, 2891, 1741, 1631, 1587, 1466, 1325, 1228, 1160, 970, 693 cm⁻¹; [α]₁^B +21.0 (*c* 0.1, MeOH); ¹H NMR (CD₃OD, 400 MHz): δ 9.33 (s, 1H), 8.34 (s, 1H), 8.22 (d, *J* = 8.0 Hz, 2H), 7.71– 7.57 (m, 3H), 4.73–4.68 (m, 1H), 3.74 (dd, *J* = 16.5, 5.3 Hz, 1H), 3.66 (dd, *J* = 16.5, 6.9 Hz, 1H); ¹³C{¹H} NMR (CD₃OD, 101 MHz): δ 169.1, 166.9, 163.9, 155.0, 133.4, 132.7, 129.2, 127.9, 118.5, 50.7, 35.7; MS (ESI) *m/z* 244 (M + H⁺, 100); HRMS (ESI) *m/z*: [M + H]⁺ Calcd for C₁₃H₁₃N₃O₂H 244.1081; found 244.1085.

(25)-2-Amino-3-[4'-(2"-naphthyl)pyrimidin-6'-yl]propanoic Acid Hydrochloride (12j). (2S)-2-Amino-3-[4'-(2"-naphthyl)pyrimidin-6'-yl]propanoic acid hydrochloride (12j) was prepared as described for 12a using tert-butyl (2S)-2-(tert-butoxycarbonylamino)-3-[4'-(2"-naphthyl)pyrimidin-6'-yl]propanoate (11j) (0.029 g, 0.070 mmol). This gave (2S)-2-amino-3-[4'-(2"-naphthyl)pyrimidin-6'yl]propanoic acid hydrochloride (12j) as a white solid (0.019 g, 95%). Mp 153-155 °C; IR (neat) 3429, 2977, 1706, 1583, 1364, 1250, 1149, 729 cm⁻¹; $[\alpha]_D^{19}$ +18.2 (c 0.1, MeOH); ¹H NMR (CD₃OD, 400 MHz): δ 9.29 (s, 1H), 8.78 (br s, 1H), 8.42 (s, 1H), 8.21 (d, J = 8.4 Hz, 1H), 8.05–8.00 (m, 2H), 7.91 (d, J = 8.9 Hz, 1H), 7.64–7.53 (m, 2H), 4.72–4.67 (m, 1H), 3.71 (dd, J = 16.8, 4.7 Hz, 1H), 3.64 (dd, J = 16.8, 7.2 Hz, 1H); ¹³C{¹H} NMR (CD₃OD, 101 MHz): δ 169.2, 166.2, 164.1, 155.3, 135.4, 133.1, 130.8, 129.2, 129.1, 129.0, 128.4, 127.5, 126.9, 123.5, 118.4, 50.8, 35.6; MS (ESI) m/z 292 ([M – H]⁻, 100); HRMS (ESI) m/z: [M – H]⁻ Calcd for C17H14N3O2 292.1092; found 292.1093.

(25)-2-Amino-3-[4'-(4"-methoxyphenyl)pyrimidin-6'-yl]propanoic Acid Hydrochloride (12k). (2S)-2-Amino-3-[4'-(4"- methoxyphenyl)pyrimidin-6'-yl]propanoic acid hydrochloride (12k) was prepared as described for 12a using *tert*-butyl (2*S*)-2-(*tert*-butoxycarbonylamino)-3-[4'-(4"-methoxyphenyl)pyrimidin-6'-yl]-propanoate (11k) (0.73 g, 1.7 mmol). Following completion, the reaction mixture was concentrated *in vacuo*. This gave (2*S*)-2-amino-3-[4'-(4"-methoxyphenyl)pyrimidin-6'-yl]propanoic acid hydrochloride (12k) as a yellow oil (0.44 g, 95%); IR (neat) 3401, 2844, 1749, 1591, 1462, 1260, 1181, 1020, 841 cm⁻¹; [*α*]_D²³ +15.0 (*c* 0.1, MeOH); ¹H NMR (CD₃OD, 400 MHz): *δ* 9.24 (s, 1H), 8.25 (d, *J* = 8.6 Hz, 2H), 8.22 (s, 1H), 7.15 (d, *J* = 8.6 Hz, 2H), 4.72–4.64 (m, 1H), 3.92 (s, 3H), 3.74–3.56 (m, 2H); ¹³C{¹H} NMR (CD₃OD, 101 MHz): *δ* 169.1, 165.9, 164.3, 163.4, 154.4, 130.0, 125.0, 117.2, 114.7, 54.9, 50.7, 35.5; MS (ESI) *m*/*z* 274 (M + H⁺, 100); HRMS (ESI) *m*/*z*: [M + H]⁺ Calcd for C₁₄H₁₅N₃O₃H 274.1186; found 274.1192.

(2S)-2-Amino-3-[4'-(4"-cyanophenyl)pyrimidin-6'-yl]propanoic Acid Hydrochloride (12l). (2S)-2-Amino-3-[4'-(4"cyanophenyl)pyrimidin-6'-yl]propanoic acid hydrochloride (12l) was prepared as described for 12a using tert-butyl (2S)-2-(tertbutoxycarbonylamino)-3-[4'-(4"-cyanophenyl)pyrimidin-6'-yl]propanoate (111) (0.068 g, 0.16 mmol). Following completion, the reaction mixture was concentrated in vacuo. This gave (2S)-2-amino-3-[4'-(4"-cyanophenyl)pyrimidin-6'-yl]propanoic acid hydrochloride (121) as a colorless oil (0.041 g, 95%). IR (neat) 3372, 2977, 1739, 1595, 1505, 1185, 1139, 837 cm⁻¹; $[\alpha]_{D}^{22}$ +12.0 (c 0.1, MeOH); ¹H NMR (CD₃OD, 400 MHz): δ 9.25 (s, 1H), 8.36 (d, J = 8.1 Hz, 2H), 8.25 (s, 1H), 7.88 (d, J = 8.1 Hz, 2H), 4.67-4.62 (m, 1H), 3.67 (dd, J = 16.6, 4.0 Hz, 1H), 3.59 (dd, J = 16.6, 7.2 Hz, 1H); ¹³C{¹H} NMR (CD₃OD, 101 MHz): δ 169.3, 165.9, 162.8, 157.1, 139.5, 132.7, 128.2, 118.2, 117.8, 114.7, 50.8, 35.5; MS (ESI) m/z 269 (M + H⁺, 100); HRMS (ESI) m/z: $[M + H]^+$ Calcd for $C_{14}H_{12}N_4O_2H$ 269.1033; found 269.1037.

ASSOCIATED CONTENT

Data Availability Statement

The data underlying this study are available in the published article and its online Supporting Material.

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.joc.3c01437.

Photophysical data for α -amino acids **12a–l** and ¹H and ¹³C NMR spectra for all compounds (PDF)

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Notes

The authors declare no competing financial interest.

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REFERENCES

(1) For reviews see: (a) Wagner, I.; Musso, H. New Naturally Occurring Amino Acids. Angew. Chem., Int. Ed. **1983**, 22, 816–828. (b) Greenstein, J. P.; Winitz, M. Chemistry of the Amino Acids; Krieger, R. E.: FL, 1984; Vol. 1–3. (c) Barrett, G. E., Ed. Chemistry and Biochemistry of the Amino Acids, Chapman and Hall: London, 1985. (d) Breslow, R.; Chmielewski, J.; Foley, D.; Johnson, B.; Kumabe, N.; Varney, M.; Mehra, R. Optically Active Amino Acid Synthesis by Artificial Transaminase Enzymes. Tetrahedron **1988**, 44, 5515–5524. (2) For example, see: (a) Easton, C. J. Free-Radical Reactions in the Synthesis of α -Amino Acids and Derivatives. Chem. Rev. **1997**, 97, 53–82. (b) Nájera, C.; Sansano, J. M. Catalytic Asymmetric Synthesis of α -Amino Acids. Chem. Rev. **2007**, 107, 4584–4671. (c) Noisier, A. F. M.; Brimble, M. A. C-H Functionalization in the Synthesis of Amino Acids and Peptides. Chem. Rev. **2014**, 114, 8775–8806.

(3) (a) Ohfune, Y. Stereoselective Routes Towards the Synthesis of Unusual Amino Acids. Acc. Chem. Res. **1992**, 25, 360–366. (b) Dougherty, D. A. Unnatural Amino Acids as Probes of Protein Structure and Function. Curr. Opin. Chem. Biol. **2000**, 4, 645–652. (c) Vogt, H.; Bräse, S. Recent Approaches Towards the Asymmetric Synthesis of α, α -Disubstituted α -Amino Acids. Org. Biomol. Chem. **2007**, 5, 406–430. (d) Smits, R.; Cadicamo, C. D.; Burger, K.; Koksch, B. Synthetic Strategies to α -Trifluoromethyl and α -Difluoromethyl Substituted α -Amino Acids. Chem. Soc. Rev. **2008**, 37, 1727–1739. (e) Mortensen, M.; Husmann, R.; Veri, E.; Bolm, C. Synthesis and Applications of Silicon-Containing α -Amino Acids. Chem. Soc. Rev. **2009**, 38, 1002–1010. (f) Lang, K.; Chin, J. W. Cellular Incorporation of Unnatural Amino Acids and Bioorthogonal Labeling of Proteins. Chem. Rev. **2014**, 114, 4764–4806.

(4) For reviews, see: (a) Sinkeldam, R. W.; Greco, N. J.; Tor, Y. Fluorescent Analogs of Biomolecular Building Blocks: Design, Properties and Applications. *Chem. Rev.* **2010**, *110*, 2579–2619. (b) Niu, L.-Y.; Chen, Y.-Z.; Zheng, H.-R.; Wu, L.-Z.; Tung, C.-H.; Yang, Q.-Z. Design Strategies of Fluorescent Probes for Selective Detection Among Biothiols. *Chem. Soc. Rev.* **2015**, *44*, 6143–6160. (c) Singh, H.; Tiwari, K.; Tiwari, R.; Pramanik, S. K.; Das, A. Small Molecule as Fluorescent Probes for Monitoring Intracellular Enzymatic Transformations. *Chem. Rev.* **2019**, *119*, 11718–11760.

(5) For reviews, see: (a) Krueger, A. T.; Imperiali, B. Fluorescent Amino Acids: Modular Building Blocks for the Assembly of New Tools for Chemical Biology. *ChemBioChem* **2013**, *14*, 788–799. (b) Kubota, R.; Hamachi, I. Protein Recognition using Synthetic Small-Molecular Binders Toward Optical Protein Sensing *In Vitro* and in Live Cells. *Chem. Soc. Rev.* **2015**, *44*, 4454–4471. (c) Harkiss, A. H.; Sutherland, A. Recent Advances in the Synthesis and Application of Fluorescent α -Amino Acids. *Org. Biomol. Chem.* **2016**, *14*, 8911– 8921. (d) Cheng, Z.; Kuru, E.; Sachdeva, A.; Vendrell, M. Fluorescent Amino Acids as Versatile Building Blocks for Chemical Biology. *Nat. Rev.* **2020**, *4*, 275–290.

(6) (a) Heim, R.; Cubitt, A. B.; Tsien, R. Y. Improved Green Fluorescence. *Nature* **1995**, 373, 663–664. (b) Tsien, R. Y. The Green Fluorescent Protein. *Annu. Rev. Biochem.* **1998**, 67, 509–544.

(7) Chen, S.; Fahmi, N. E.; Wang, L.; Bhattacharya, C.; Benkovic, S. J.; Hecht, S. M. Detection of Dihydrofolate Reductase Conformational Change by FRET Using Two Fluorescent Amino Acids. *J. Am. Chem. Soc.* **2013**, *135*, 12924–12927.

(8) Cheruku, P.; Huang, J.-H.; Yen, H.-J.; Iyer, R. S.; Rector, K. D.; Martinez, J. S.; Wang, H.-L. Tyrosine-Derived Stimuli Responsive, Fluorescent Amino Acids. *Chem. Sci.* **2015**, *6*, 1150–1158.

(9) For examples of cyanotryptophans, see: (a) Talukder, P.; Chen, S.; Roy, B.; Yakovchuk, P.; Spiering, M. M.; Alam, M. P.; Madathil, M. M.; Bhattacharya, C.; Benkovic, S. J.; Hecht, S. M. Cyanotryptophans as Novel Fluorescent Probes for Studying Protein Conformational Changes and DNA-Protein Interaction. *Biochemistry* **2015**, *54*, 7457–7469. (b) Bartoccini, F.; Bartolucci, S.; Mari, M.; Piersanti, G. A Simple, Modular Synthesis of C4-Substituted Tryptophan Derivatives. *Org. Biomol. Chem.* **2016**, *14*, 10095–10100. (c) Hilaire, M. R.; Ahmed, I. A.; Lin, C.-W.; Jo, H.; DeGrado, W. F.; Gai, F. Blue Fluorescent Amino Acid for Biological Spectroscopy and Microscopy. *Proc. Natl. Acad. Sci. U.S.A.* **2017**, *114*, 6005–6009. (d) Boville, C. E.; Romney, D. K.; Almhjell, P. J.; Sieben, M.; Arnold, F. H. Improved Synthesis of 4-Cyanotryptophan and other Tryptophan Analogues in Aqueous Solvents Using Variants of TrpB from *Thermotoga maritima*. *J. Org. Chem.* **2018**, *83*, 7447–7452. (e) Zhang, K.; Ahmed, I. A.; Kratochvil, H. T.; DeGrado, W. F.; Gai, F.; Jo, H. Synthesis and Application of the Blue Fluorescent Amino Acid L-4-Cyanotryptophan to Assess Peptide-Membrane Interactions. *Chem. Commun.* **2019**, *55*, 5095–5098.

(10) For example, see: (a) Wen, J.; Zhu, Bi. L.-L.; Bi, Q.-W.; Shen, Z.-Q.; Li, X.-X.; Li, X.; Wang, Z.; Chen, Z. Highly N²-Selective Coupling of 1,2,3-Triazoles with Indole and Pyrrole. Chem. - Eur. J. 2014, 20, 974-978. (b) Williams, T. J.; Reay, A. J.; Whitwood, A. C.; Fairlamb, I. J. S. A Mild and Selective Pd-Mediated Methodology for the Synthesis of Highly Fluorescent 2-Arylated Tryptophans and Tryptophan-Containing Peptides: A Catalytic Role for Pd⁰ Nanoparticles. Chem. Commun. 2014, 50, 3052-3054. (c) Mendive-Tapia, L.; Zhao, C.; Akram, A. R.; Preciado, S.; Alberico, F.; Lee, M.; Serrels, A.; Kielland, N.; Read, N. D.; Lavilla, R.; Vendrell, M. Spacer-Free BODIPY Fluorogens in Antimicrobial Peptides for Direct Imaging of Fungal Infection in Human Tissue. Nat. Commun. 2016, 7, No. 10940. (d) Subiros-Funosas, R.; Mendive-Tapia, L.; Sot, J.; Pound, J. D.; Barth, N.; Varela, Y.; Goñi, F. M.; Paterson, M.; Gregory, C. D.; Albericio, F.; Dransfield, I.; Lavilla, R.; Vendrell, M. A Trp-BODIPY Cyclic Peptide for Fluorescence Labelling of Apoptotic Bodies. Chem. Commun. 2017, 53, 945-948. (e) Subiros-Funosas, R.; Ho, V. C. L.; Barth, N. D.; Medive-Tapia, L.; Pappalardo, M.; Barril, X.; Ma, R.; Zhang, C.-B.; Qian, B.-Z.; Sintes, M.; Ghashghaei, O.; Lavilla, R.; Vendrell, M. Fluorogenic Trp(redBODIPY) Cyclopeptide Targeting Keratin 1 for Imaging of Aggressive Carcinomas. Chem. Sci. 2020, 11, 1368-1374.

(11) (a) Gilfillan, L.; Artschwager, R.; Harkiss, A. H.; Liskamp, R. M. J.; Sutherland, A. Synthesis of Pyrazole Containing α -Amino Acids via a Highly Regioselective Condensation/aza-Michael Reaction of β -Aryl α , β -Unsaturated Ketones. *Org. Biomol. Chem.* **2015**, *13*, 4514–4523. (b) Bell, J. D.; Harkiss, A. H.; Nobis, D.; Malcolm, E.; Knuhtsen, A.; Wellaway, C. R.; Jamieson, A. G.; Magennis, S. W.; Sutherland, A. Conformationally Rigid Pyrazoloquinazoline α -Amino Acids: One and Two-Photon Induced Fluorescence. *Chem. Commun.* **2020**, *56*, 1887–1890.

(12) (a) Bell, J. D.; Morgan, T. E. F.; Buijs, N.; Harkiss, A. H.; Wellaway, C. R.; Sutherland, A. Synthesis and Photophysical Properties of Benzotriazole-Derived Unnatural α -Amino Acids. J. Org. Chem. 2019, 84, 10436–10448. (b) Riley, L. M.; Mclay, T. N.; Sutherland, A. Synthesis and Fluorescent Properties of Alkynyl- and Alkenyl-Fused Benzotriazole-Derived α -Amino Acids. J. Org. Chem. 2023, 88, 2453–2463.

(13) Harkiss, A. H.; Bell, J. D.; Knuhtsen, A.; Jamieson, A. G.; Sutherland, A. Synthesis and Fluorescent Properties of β -Pyridyl α -Amino Acids. J. Org. Chem. **2019**, 84, 2879–2890.

(14) Möschwitzer, V. D.; Kariuki, B. M.; Redman, J. E. Asymmetric Synthesis of Aminopyrimidine and Cyclic Guanidine Amino Acids. *Tetrahedron Lett.* **2013**, *54*, 4526–4528. A pyrimidine-derived α amino acid has also been prepared by Heck coupling of an iodopyrimidine with methyl 2-acetamidoacrylate, followed by Rh(I)catalyzed asymmetric hydrogenation

(15) Vu, H.-D.; Renault, J.; Toupet, L.; Uriac, P.; Gouault, N. From Aspartic Acid to Dihydropyridone-2-carboxylates: Access to Enantiopure 6-Substituted 4-Oxo- and 4-Hydroxypipecolic Acid Derivatives. *Eur. J. Org. Chem.* **2013**, 2013, 6677–6686.

(16) Bowden, K.; Jones, E. R. H. Researches on Acetylenic Compounds. Part IX. Heterocyclic Compounds Derived from Ethynyl Ketones. J. Chem. Soc. **1946**, 953–954.

(17) (a) Karpov, A. S.; Muller, T. J. J. New Entry to a Three-Component Pyrimidine Synthesis by TMS-Ynones via Sonogashira Coupling. *Org. Lett.* **2003**, *5*, 3451–3454. (b) Romanov, A. R.; Rulev, A. Y.; Ushakov, I. A.; Muzalevskiy, V. M.; Nenajdenko, V. G. One-Pot, Atom and Step Economy (PASE) Assembly of Trifluoromethylated Pyrimidines from CF_3 -Ynones. *Eur. J. Org. Chem.* **2017**, 2017, 4121–4129.

(18) (a) Adlington, R. M.; Baldwin, J. E.; Catterick, D.; Pritchard, G. J. A Versatile Approach to Pyrimidin-4-yl Substituted α -Amino Acids from Alkynyl Ketones; The Total Synthesis of L-Lathyrine. *Chem. Commun.* **1997**, 1757–1758. (b) Adlington, R. M.; Baldwin, J. E.; Catterick, D.; Pritchard, G. J. The Synthesis of Pyrimidin-4-yl Substituted α -Amino Acids. A Versatile Approach from Alkynyl Ketones. J. Chem. Soc., Perkin Trans. 1 **1999**, 855–866.

(19) (a) Bagley, M. C.; Dale, J. W.; Hughes, D. D.; Ohnesorge, M.; Phillipas, N. G.; Bower, J. Synthesis of Pyridines and Pyrido[2,3d]pyrimidines by the Lewis Acid Catalysed Bohlmann-Rahtz Heteroannulation Reaction. Synlett 2001, 2001, 1523–1526.
(b) Hughes, D. D.; Bagley, M. C. One or Two-Step Bohlmann-Rahtz Heteroannulation of 6-Aminouracil Derivatives for the Synthesis of Pyrido[2,3-d]pyrimidines. Synlett 2002, 1332–1334.
(c) Bagley, M. C.; Brace, C.; Dale, J. W.; Ohnesorge, M.; Phillips, N. G.; Xiong, X.; Bower, J. Synthesis of Tetrasubstituted Pyridines by the Acid-Catalysed Bohlmann-Rahtz Reaction. J. Chem. Soc., Perkin Trans. 1 2002, 1663–1671.

(20) The ytterbium-catalyzed heterocyclization could also be performed on gram scale as demonstrated by the synthesis of **11e**. See Experimental Section for full details.

(21) For example, see: (a) Achelle, S.; Nouira, I.; Pfaffinger, B.; Ramondenc, Y.; Plé, N.; Rodríquez-López, J. V-Shaped 4,6-Bis(arylvinyl)pyrimidine Oligomers: Synthesis and Optical Properties. *J. Org. Chem.* **2009**, *74*, 3711–3717. (b) Hadad, C.; Achelle, S.; García-Martinez, J. C.; Rodríquez-López, J. 4-Arylvinyl-2,6-di-(pyridine-2-yl)pyrimidines: Synthesis and Optical Properties. *J. Org. Chem.* **2011**, *76*, 3837–3845. (c) Achelle, S.; Barsella, A.; Baudequin, C.; Caro, B.; Robin-le Guen, F. Synthesis and Photophysical Investigation of a Series of Push-Pull Arylvinyldiazine Chromophores. *J. Org. Chem.* **2012**, *77*, 4087–4096. (d) Aydıner, B.; Seferoglu, Z. Proton Sensitive Functional Organic Fluorescent Dyes Based on Coumarin-imidazo[1,2-a]pyrimidine; Syntheses, Photophysical Properties, and Investigation of Protonation Ability. *Eur. J. Org. Chem.* **2018**, *2018*, 5921–5934.