| 1  | Supplementary Information.   |
|----|--|
| 2  |  |
| 3  | No evidence of sustained nonzoonotic Plasmodium knowlesi transmission in Malaysia from         |
| 4  | modelling malaria case data  |
| 5  |  |
| 6  | Fornace, et. al.   |
| 7  |  |
| 8  | 1. Malaysian malaria surveillance data.  |
| 9  |  |
| 10 | 1.1 Geolocation and cleaning malaria surveillance data.  |
| 11 |  |
| 12 | Malaria surveillance data included all malaria case notifications in Malaysia reported between |
| 13 | January 2012 to December 2020. All case records included geographic data including the         |
| 14 | names of kampungs (villages) where cases were resident and locations of infections. Exact      |
| 15 | Global Positioning System (GPS) coordinates were not available for over 20% of study           |
| 16 | records, with increasing availability in later years as GPS technology was routinely           |
| 17 | implemented.   |
| 18 |  |

19 To assess the accuracy of recorded GPS coordinates, coordinates with less than 2 decimal 20 points were first mapped onto Google Earth and locations were manually confirmed or GPS 21 coordinates were excluded as unreliable. For each year, GPS coordinates were also imported 22 into Quantum GIS and overlaid with administrative shapefiles for Malaysia. State and district 23 names were extracted from administrative polygons and compared to reported states and 24 districts listed within surveillance data. For all data where administrative units did not match, 25 GPS coordinates were marked as unreliable and manually confirmed. For remaining GPS 26 coordinates, data was mapped onto publicly available satellite data (Google Earth, Open 27 Street Map) to confirm locations. All kampungs were assigned a unique code and records 28 were manually checked to confirm any alternate spellings of names. GPS coordinates reported 29 from the same kampung were compared to ensure locations were consistent. For records with 30 no GPS coordinates, the centroid of the kampung was used to geolocate records. Kampung 31 centroids were either calculated as a mean of all available records from that kampung or 32 manually identified from satellite data, census data or by personnel familiar with the region. 33

To clean nonspatial malaria data, yearly surveillance data was imported into R (R Statistical
Software, v 3.6.2). As the database structure changed slightly during the reporting period, data
was coded with standard headings and merged. Variables not routinely collected throughout

- the study period (e.g. gametocyte presence) were coded as NA for missing records. The dates
  of symptom onset, diagnosis, hospitalisation and notification were extracted for all records.
  Onset dates were excluded as unreliable if they occurred after diagnosis and/or notification
  dates. Due to recall bias, symptom onset dates were additionally excluded as unreliable if they
  occurred more than 30 days prior to diagnosis.
- 42
- 43 1.2 Surveillance data characteristics.
- 44

45 Between 16 December 2011 and 3 January 2021, 32,635 malaria cases were reported to the 46 national surveillance system (Supplementary Tables 1 and 2).

- 47
- 48 Supplementary Table 1. Malaria cases by parasite species in East and West Malaysia\*,
- 49 number (%)
- 50

|          | Р.         | P. knowlesi | P. malariae | P. ovale | P. vivax | Mixed      | Total  |
|----------|------------|-------------|-------------|----------|----------|------------|--------|
|          | falciparum |             |             |          |          | infections |        |
| East     | 2,196      | 19,931      | 1,096       | 99       | 2,566    | 205        | 26,093 |
| Malaysia | (6.7%)     | (61.1%)     | (3.4%)      | (0.3%)   | (7.9%)   | (0.6%)     |        |
| (Borneo) |            |             |             |          |          |            |        |
| West     | 853        | 3,212       | 48          | 20       | 2,375    | 34         | 6,542  |
| Malaysia | (13.0%)    | (49.1%)     | (0.7%)      | (0.3%)   | (36.3%)  | (0.5%)     |        |

- 51 \* Includes all malaria cases, including imported cases and reoccurences
- 52

## 53 Supplementary Table 2. Demographic breakdown of malaria cases, number (%)

54

|                   | P. falciparum | P. knowlesi    | P. vivax      | All Malaria    |
|-------------------|---------------|----------------|---------------|----------------|
| Ν                 | 3,049         | 23,143         | 4,941         | 32,635         |
| Male (n, %)       | 2,484 (81.5%) | 18,701 (80.8%) | 4,209 (85.2%) | 26,651 (81.7%) |
| Age (median, IQR) | 32 (21 – 43)  | 36 (26 – 49)   | 30 (21 – 42)  | 35 (24 – 47)   |
| Indigenous (n, %) | 1,678 (55.0%) | 23,016 (99.4%) | 2,342 (47.4%) | 28,149 (86.3%) |

55

56

## 57 **2. Serial interval estimation.**

58

Assessing the probability of human-mosquito-human transmission of *P. knowlesi* requires
estimating the timing between reported *P. knowlesi* cases from the same transmission chain.
This requires estimating the duration of a series of sequential processes which need to occur

62 in a human-mosquito-human transmission cycle. The generation time (Tg) refers to the 63 duration of time between an infection and the individual infecting another person [1]. The serial 64 interval (SI), the time between clinical presentation of primary and secondary cases, is more 65 commonly used as infection is typically unobserved [2]. While these intervals can be inferred 66 from contract tracing data for directly transmitted diseases, these intervals can only be estimated indirectly for vector-borne diseases [3]. Additionally, there is a lack of empirical 67 68 evidence on human-mosquito-human P. knowlesi transmission as this has only been 69 experimentally observed once. The rapid replication cycle of *P. knowlesi* and weak evidence 70 of adaptation to humans suggests this may differ from other nonzoonotic malaria species [4]. 71

72 To estimate the generation time and serial intervals for human-mosquito-human P. knowlesi 73 transmission, we used a quantitative model-based approach developed by Huber et. al [5]. 74 This models the SI and Tg as random variables based on the sum of random variables 75 representing the sequential steps in the transmission cycle including: the prepatent period, the 76 human to mosquito transmission period, the extrinsic incubation period, the mosquito to 77 human transmission period and infection to detection periods. These were parameterised 78 using a combination of data from secondary literature and the Malaysian malaria surveillance 79 dataset.

80

81 Within this analysis, we made two important assumptions. First, we assumed that 82 asymptomatic human infections did not contribute to transmission. This was based on the very 83 low parasite densities of human *P. knowlesi* infections detected during community surveys 84 within Malaysia, likely insufficient to infect mosquitoes [6-8]. Second, we assumed that 85 individuals became non-infectious on the date of diagnosis and treatment. This is based on the Malaysian national malaria policy of hospitalising malaria cases from diagnosis until 86 87 confirmation as microscopy negative [9]. Within Malaysia, all malaria treatment is free through 88 government healthcare providers.

89

*1. Prepatent period (PREP).* Estimates of the prepatent period for *P. knowlesi* were based on
experimental *P. knowlesi* infections in people following bites from infected *An. balabacensis*,
the main vector in Malaysian Borneo [10]. This was modelled as Normal(10.6 days, 1.15 days)
based on data from Table 1. The time between an individual developing a patent infection and
becoming symptomatic was modelled as Normal(3.5 days, 0.2 days) based on this
experimental data.

96

97 2. Human to mosquito transmission period (HTMP). No data was available on the duration or
98 timing of infectiousness of *P. knowlesi* in humans. However, multiple studies have reported *P*.

99 knowlesi gametocytes in people at both microscopic and submicroscopic densities [11-13]. 100 While *P. knowlesi* gametocytes are highly synchronous in macaques [4, 14], available 101 evidence suggests this is not the case in human infections [13]. Infectiousness to mosquitoes 102 is dependent on the density of malaria gametocytes, with infectious individuals predominantly 103 having gametocyte densities high enough to be detected microscopically [15]. The presence 104 of microscopically detected P. knowlesi gametocytes is captured by the Malaysian 105 surveillance data for cases reported after 2014. These data showed a clear increasing trend 106 in the proportion of cases with observed gametocytes relative to the time since symptom 107 onset. Using this data, we modelled  $Y_{i}$  the number of infectious individuals (defined as having 108 gametocyte densities high enough to be detected by microscopy) out of the total number of 109 cases ( $n_i$ ) as the realisation of a binomial random variable  $Y_i \sim \text{Binomial}(n_i, \pi_i)$ . The probability 110 an individual was infectious ( $\pi_i$ ) was specified as logit( $\pi_i$ ) =  $\beta_0 + \beta_1 x_i$ , where  $\beta_0$  is the intercept and  $\beta_1 x_i$  describes the effect of days since symptom onset. This model was then used to 111 112 simulate the probability and duration of infectiousness using 10,000 simulations. We assumed 113 that no individuals had sufficient gametocyte densities to become infectious prior to developing 114 patent malaria and, if an individual became infectious, they remained infectious until treatment. 115 The duration of patent malaria was estimated as the sum of the time between an individual 116 developing patent malaria and becoming symptomatic and the time between symptom onset 117 and treatment. The time between symptom onset and treatment was fit to Malaysian 118 surveillance data and modelled as a Gamma distribution. As there was insufficient data to 119 estimate the probability of infectiousness two weeks past symptom onset, we assumed a 120 constant probability of infectiousness after 15 days.

121

These probabilities of infectiousness were multiplied by a constant mosquito-to-human ratio estimated from empirical data on human landing catches in Sabah, Malaysia [16, 17]. While other models have used time-varying mosquito-to-human ratios ([5, 18]), Malaysia is equatorial and strong seasonal trends in mosquito densities are not typically observed. This could be expanded in future work to model geographic heterogeneities or uncertainties around this number.

128

3. Extrinsic incubation period (EIP). The next step in the transmission cycle occurs within the
mosquito. The extrinsic incubation period (EIP) is the time between parasite transmission
from an infectious human to a mosquito and the production of sporozoites within the
mosquito. Previous studies have estimated the EIP for *P. knowlesi* as 10 days [17, 19]. An
experimental study of *P. knowlesi* in *An. stephensi* identified the EIP as 6.8 days, however
this is not a natural *P. knowlesi* vector in Malaysia [20]. To represent the uncertainty around

135 these estimates, EIP was modelled as ~ Normal(10 days, 1 day).

*4. Mosquito to human transmission period (MHTP).* Following the approach described by
Huber et. al, we modelled the time between the end of the EIP and the time of a subsequent
human infection as a geometric random variable with probability 1 – p, where p is the
constant probability of daily survival [5]. This assumes no variability in mosquito daily
survival and no association between mosquito survival and probability of human infection.
The probability of daily survival was estimated as 0.85 based on *An. balabacensis* data from
Sabah, Malaysia [17].

144

145 5. Infection to detection period (IDP). As the SI is based on reported clinical cases,

146 estimation of the SI also requires assessing the time between human infection by a mosquito

147 and detection by the health facility. This was modelled as the sum of the prepatent period,

time between patency and symptom onset and time between symptom onset and diagnosis,

149 with variables parameterised based on secondary literature or malaria surveillance data as

- 150 described previously.
- 151

Probabilistic descriptions of the Tg and SI were obtained by summing random variablesusing the approach developed by Huber et. al [5]. First, the Tg was calculated as:

154

155 
$$Tg(i + j + k + l)$$

156

$$= \sum_{i} \sum_{j} \sum_{k} \sum_{l} (\Pr(PREP = i) \times (\Pr(HMTP = j) \times (\Pr(EIP = k) \times (\Pr(MHTP = l))))$$

157 158

Where *i*, *j*, *k* and *l* are dummy variables used to calculate the probability of the Tg for all combinations of *i*, *j*, *k* and *l*. The same approach was used to calculate the SI, accounting for the infection to detection periods of the primary and secondary cases:

163 
$$SI(-i + j + k) = \sum_{i} \sum_{j} \sum_{k} (Pr(IDP = i) \times (Pr(GI = j) \times (Pr(IDP = k))))$$

### 164 **Supplementary Table 3.** Parameters used for SI and Tg estimation

165

| Description                | Estimate                                   | Source                   |  |
|----------------------------|--|--------------------------|--|
| Prepatent period           | ~Normal(10.6 days, 1.14 days)              | [21]                     |  |
| Time from patent infection | ~Normal(3.5 days, 0.2 days)                | [21]                     |  |
| to symptom onset           |  |                          |  |
| Time from symptom onset    | ~Gamma(2.484, 0.473)                       | Fit to Malaysian         |  |
| until treatment            |  | surveillance data        |  |
| Duration of infectiousness | Binomial model fit to presence/ absence of | Fit to Malaysian         |  |
|                            | microscopically observed gametocytes       | surveillance data,       |  |
|                            | from time from infection                   | constant rate of         |  |
|                            |  | infectiousness after day |  |
|                            |  | 15                       |  |
| Mosquito to human ratio    | 4  | Mean biting rate         |  |
|                            |  | reported by [22]         |  |
| Probability of mosquito    | 0.85                                       | [17]                     |  |
| daily survival             |  |                          |  |
| Infection to detection     | Time from infection to symptom onset +     |                          |  |
| period                     | Time from symptom onset to treatment       |                          |  |

166

167

### 168 3. Estimation of R<sub>c</sub>

169

### 170 3.1 Transmission likelihood

171

172 Based on estimates of the duration of infectiousness, we fit shifted Rayleigh distributions to 173 describe a prior distribution of possible serial intervals for nonzoonotic P. knowlesi 174 transmission (Figure S1a) and P. falciparum/ P. vivax transmission (Figure S1b). Using a 175 fixed value for the spatial parameter ( $\delta$ ) of 0.1, we estimated the likelihood of two cases being connected based on the geographic location and time of reporting (Figure S1). The 176 177 fixed value for the spatial parameter corresponded to most cases being infected within a 178 10km radius of their reported residence location; this parameter was obtained based on 179 reported travel history from case investigations reporting most individuals remaining within 180 the same village or district prior to their diagnosis. Individuals with a history of long-range 181 travel were classified as imported cases according to Malaysian Ministry of Health 182 surveillance guidelines.

Supplementary Figure 1. Likelihood of two cases being part of the same transmission chain
based on notification time (X axis) and geographic distance (Y axis) with priors used for a.)
nonzoonotic *P. knowlesi* transmission and b.) *P. falciparum* and *P. vivax* transmission



188 189

190 We used an adapted version of the NetRate model to estimate  $R_C$  [18, 23-25]. Data was 191 input as a series of *n* infections ( $I_1, ..., I_n$ ) reported at times  $t = \{t_1, ..., t_n\}$  with a binary 192 classification of importation status. The serial interval parameters were represented by the 193 function  $f_1$  and the relationship between geographic location of cases and likelihood of 194 transmission was represented by the function  $f_2$ , giving the function:

- 195
- 196

 $f(x_i, t_i | x_j, t_j; \alpha_{i,j}, \beta) = f1(t_i | t_j; \alpha_{i,j}) \ge f2(x_i | x_j; \beta)$ 

197

198 where *t* is the time, *x* is the spatial locations,  $\alpha$  is the transmission rate and  $\beta$  are the spatial 199 parameters. The hazard is defined as the pairwise likelihood divided by the survival function 200 as:

201 
$$H = \frac{f(x_i, t_i | x_j, t_j; \alpha_{i,j}, \beta)}{S(x_i, t_i | x_j, t_j; \alpha_{i,j}, \beta)}$$

202

The pairwise likelihood of a case reported at time  $t_i$  and location  $x_j$  infecting a case reported at time  $t_i$  and location  $x_i$  is:

206 
$$f(x_i, t_i | x_j, t_j; \alpha_{i,j}, \beta) = \alpha(t_i - t_j - \gamma)e^{-\frac{1}{2}\alpha(t_i - t_j - \gamma)}\frac{1}{\beta}$$

207  
208 With the hazard term simplifying to:  
209  
210 
$$H(x_i, t_i | x_j, t_j; a_{i,j}, \beta) = \beta \alpha (t_i - t_j - \gamma) e^{-\beta (x_i - x_j)}$$
211  
212 And the survival term as:  
213  
214 
$$S(x_i, t_i | x_j, t_j; a_{i,j}, \beta) = e^{-\frac{1}{2}\alpha (t_i - t_j - \gamma)} \frac{1}{\beta}$$
215  
216 Integrating the survival term over distances is equivalent to:  
217  
218 
$$S(x_i, t_i | x_j, t_j; a_{i,j}, \beta) = e^{-\frac{1}{2}\alpha (t_i - t_j - \gamma)} \frac{\sqrt{\pi}}{2\sqrt{\beta}}$$
219  
220 And the hazard function is:  
221  
222 
$$H(x_i, t_i | x_j, t_j; a_{i,j}, \beta) = \frac{\alpha (t_i - t_j - \gamma) e^{-\frac{1}{2}\alpha (t_i - t_j - \gamma)} e^{-\beta (x_i - x_j)}}{e^{-\frac{1}{2}\alpha (t_i - t_j - \gamma)} \frac{\sqrt{\pi}}{2\sqrt{\beta}}}$$
223 Which simplifies to:

225 
$$H(x_i, t_i | x_j, t_j; \alpha_{i,j}, \beta) = \frac{2\sqrt{\beta\alpha}(t_i - t_j - \gamma)e^{-\beta(x_i - x_j)^2}}{\sqrt{\pi}}$$

To account for potential unobserved sources of infection, we used Epsilon edges. Within this framework, a high  $\epsilon$  value assumes the case is very likely to be from an unobserved source unless two cases have a high likelihood of being linked while a low  $\epsilon$  assumes unobserved sources of infection are highly unlikely.

With  $\varepsilon$ , this gives:

234 
$$f(\boldsymbol{t},\boldsymbol{x}; \boldsymbol{\epsilon},\boldsymbol{\beta}) = \prod_{t_i \in \boldsymbol{t}} S_0(\boldsymbol{\epsilon}_i) \prod_{I_k: t_k < t_i} S(x_i, t_i | x_j, t_j; \alpha_{i,j}, \beta) \left( H_0(\boldsymbol{\epsilon}_i) + \sum_{I_k: t_k < t_i} H(x_i, t_i | x_j, t_j; \alpha_{i,j}, \beta) \right)$$

With the objective function as:

minimize 
$$\alpha \in -\log f(\mathbf{t}, \mathbf{x}; \in, \boldsymbol{\beta})$$
 subject to  $\alpha, \in, \boldsymbol{\beta} > 0 \quad \forall i, j$ 

Geolocated time series data was used to fit models with varying priors on  $\varepsilon$ , reflecting the uncertainty around the proportion of zoonotic transmission. Model fit was evaluating using the second order AIC (AICc), (Table S3). The best fitting models had no estimates of  $R_c$ greater than one for both East and West Malaysia (Figure S2).

244

Supplementary Table 4. Model selection statistics for *P. knowlesi* models with varying normally distributed priors on  $\varepsilon$  using a fixed value of  $\delta$ =0.1 and priors of Normal(0.002, 0.001) for  $\alpha$ 

248

| Epsilon Priors |        |       |                     |             |  |  |
|----------------|--------|-------|---------------------|-------------|--|--|
| Dataset        | Mean   | SD    | Mean R <sub>c</sub> | AICc*       |  |  |
|                | 0.0001 | 0.001 | 0.945               | -1012488877 |  |  |
|                | 0.001  | 0.01  | 0.796               | -1012540285 |  |  |
| East Malaysia  | 0.01   | 0.1   | 0.431               | -1012612525 |  |  |
|                | 0.1    | 1     | 0.362               | -1012631421 |  |  |
|                | 1      | 1     | 0.074               | -1012730557 |  |  |
|                | 0.0001 | 0.001 | 0.788               | -44737596   |  |  |
|                | 0.001  | 0.01  | 0.450               | -44748816   |  |  |
| West Malaysia  | 0.01   | 0.1   | 0.131               | -44762856   |  |  |
|                | 0.1    | 1     | 0.115               | -44760900   |  |  |
|                | 1      | 1     | 0.015               | -44768972   |  |  |

249

\* Lower AICc values represent improved model fit

250

# 252 Supplementary Figure 2. Estimated *R<sub>c</sub>* values from the best fitting models for *P. knowlesi*. West Malaysia East Malaysia



### 253 254

We additionally conducted sensitivity analyses for *P. falciparum* and *P. vivax* models to assess the impact of varying priors on  $\varepsilon$  while using established distributions for the temporal component. As transmission of these models is known to be nonzoonotic and extensive local transmission was documented during the study period, we excluded unlikely scenarios where all cases were imported (Supplementary Table 4).

260

**Supplementary Table 5.** Model selection statistics for *P. knowlesi* models with varying normally distributed priors on  $\varepsilon$  using a fixed value of  $\delta$ =0.1 and priors of Normal(0.003, 0.001) for  $\alpha$  for a.) *P. falciparum* and b.) *P. vivax* 

264 a.)

|               | Epsilon Pri |         |                     |                   |           |
|---------------|-------------|---------|---------------------|-------------------|-----------|
| Dataset       | Mean        | SD      | Mean R <sub>c</sub> | Percent $R_C > 1$ | AICc*     |
| East Malaysia | 0.0001      | 0.001   | 0.568               | 20.89%            | 250425034 |
|               | 0.00001     | 0.01    | 0.568               | 20.95%            | 250425274 |
|               | 0.000001    | 0.1     | 0.565               | 20.90%            | 250487594 |
| West Malaysia | 0.0001      | 0.001   | 0.386               | 9.14%             | 6401555   |
|               | 0.00001     | 0.0001  | 0.101               | 10.13%            | 6401460   |
|               | 0.000001    | 0.00001 | 0.099               | 9.91%             | 6403864   |

265

266 b.)

|               | Epsilon Priors |         |                     |                   |           |
|---------------|----------------|---------|---------------------|-------------------|-----------|
| Dataset       | Mean           | SD      | Mean R <sub>c</sub> | Percent $R_C > 1$ | AICc*     |
| East Malaysia | 0.0001         | 0.001   | 0.524               | 20.31%            | 652347120 |
|               | 0.00001        | 0.01    | 0.547               | 21.19%            | 652347568 |
|               | 0.000001       | 0.1     | 0.558               | 21.47%            | 652347952 |
| West Malaysia | 0.0001         | 0.001   | 0.230               | 8.61%             | 219812878 |
|               | 0.00001        | 0.0001  | 0.246               | 8.92%             | 219813166 |
|               | 0.000001       | 0.00001 | 0.250               | 9.13%             | 219842030 |

### 268

### 269 3.2 Spatiotemporal models of R<sub>c</sub>

270

271 To visualise the spatial and temporal distributions of P. falciparum and P. vivax cases with 272  $R_{c}$  estimates > 1, we fit geostatistical models using Integrated Nested Laplace 273 Approximation (INLA) in R statistical software. For each species, we classified R<sub>c</sub> estimates 274 into binary classes based on whether R<sub>c</sub> estimates were greater than 1. We resampled all 275 data to 5 km<sup>2</sup> grid cells and calculated the total number of *P. falciparum* or *P. vivax* cases 276 per grid cell  $(m_{i,i})$ ; i = 1...n; t = 1...n; where i indexes location and t indexes year. For each 277 species, we fit separate models for the probability of a malaria case leading to onward 278 transmission ( $R_c > 1$ ) was modelled as:

279

280

 $Y_{i,t} \sim \text{Binomial}(m_{i,t}, \pi_{i,t})$ 

281 Where  $\pi_{i,t}$  is the probability of a malaria case having an  $R_c$  estimate over 1 with the linear 282 predictor for the binomial model specified as:

283

 $\log it(\pi_{i,t}) = \beta_0 + w_i + e_t$ 

Where  $\beta_0$  is the intercept,  $w_i$  is the spatial effect and  $e_t$  is the temporal effect. Candidate 285 286 models including the temporal effect as a random effect, temporally structured random walk 287 models or autoregressive models were evaluated using the Deviance Information Criteria 288 (DIC). The final model included the temporal effect as a temporally structured random walk 289 model of order 2 [26]. The spatial effect  $w_i$  was modelled as a Matern covariance function 290 implemented using the stochastic partial differential equations approach. All models used 291 1,000 samples to estimate posterior probabilities and were visualised in R. As there were no 292 R<sub>c</sub> estimates above 1 for the best fitting P. knowlesi models, models were only fit for P. 293 falciparum and P. vivax. Mean and maximum R<sub>c</sub> estimates per village per year are included 294 below in Figure S3.

- 297 **Supplementary Figure 3.**  $R_c$  estimates for *P. knowlesi* per village per year, including a) 298 mean  $R_c$  estimates and b) maximum  $R_c$  estimates (areas with highest probability of 299 nonzoonotic *P.knowlesi* transmission)
- 300
- 301 a.)



# 303 b.)



313 1. Champredon, D. and J. Dushoff, Intrinsic and realized generation intervals in 314 infectious-disease transmission. Proc Biol Sci, 2015. 282(1821): p. 20152026. 315 2. Fine, P.E., The interval between successive cases of an infectious disease. Am J 316 Epidemiol, 2003. 158(11): p. 1039-47. Park, S.W., D. Champredon, and J. Dushoff, Inferring generation-interval 317 3. 318 distributions from contact-tracing data. J R Soc Interface, 2020. 17(167): p. 319 20190719. 320 4. Hawking, F., M.J. Worms, and K. Gammage, 24 and 48 hour cycles of malaria parasites in the blood; their purpose, production and control. Transactions of the 321 322 Royal Society of Tropical Medicine and Hygiene, 1968. 82(6): p. 731-765. 323 5. Huber, J.H., et al., Quantitative, model-based estimates of variability in the 324 generation and serial intervals of Plasmodium falciparum malaria. Malar J, 2016. 325 **15**(1): p. 490. Fornace, K.M., et al., Environmental risk factors and exposure to the zoonotic malaria 326 6. 327 Plasmodium knowlesi across Northern Sabah, Malaysia: a cross-sectional survey. 328 Lancet Planet Health, 2019. 3(4): p. E179-E186. Fornace, K.M., et al., Exposure and infection to Plasmodium knowlesi in case study 329 7. communities in Northern Sabah, Malaysia and Palawan, The Philippines. PLoS 330 331 Neglected Tropical Diseases [electronic resource], 2018. 12(6): p. e0006432. 332 8. Fornace, K.M., et al., Asymptomatic and Submicroscopic Carriage of Plasmodium 333 knowlesi Malaria in Household and Community Members of Clinical Cases in Sabah, 334 Malaysia. J Infect Dis, 2016. 213(5): p. 784-7. 335 9. Ministry of Health Malaysia, Management guidelines of malaria in Malaysia. 2014, Vector Borne Disease Sector, Disease Control Division, Ministry of Health, Malaysia: 336 337 Putra Java. Chin, W., et al., Experimental mosquito-transmission of Plasmodium knowlesi to man 338 10. 339 and monkey. American Journal of Tropical Medicine & Hygiene, 1968. 17(3): p. 355-340 8. 341 11. Grigg, M.J., et al., Artesunate-mefloquine versus chloroquine for treatment of 342 uncomplicated Plasmodium knowlesi malaria in Malaysia (ACT KNOW): an open-343 label, randomised controlled trial. Lancet Infect Dis, 2016. 16(2): p. 180-8. 344 12. Maeno, Y., et al., Plasmodium knowlesi and human malaria parasites in Khan Phu, 345 Vietnam: Gametocyte production in humans and frequent co-infection of mosquitoes. 346 Parasitology, 2017. 144(4): p. 527-535. 347 Lee, K.S., J. Cox-Singh, and B. Singh, Morphological features and differential counts 13. 348 of Plasmodium knowlesi parasites in naturally acquired human infections. Malar J, 349 2009. 8: p. 73. 350 Anderios, F., A. Noorrain, and I. Vythilingam, In vivo study of human Plasmodium 14. 351 knowlesi in Macaca fascicularis. Exp Parasitol, 2010. 124(2): p. 181-9. 352 Bradley, J., et al., Predicting the likelihood and intensity of mosquito infection from 15. 353 sex specific Plasmodium falciparum gametocyte density. Elife, 2018. 7. 354 16. Fornace, K.M., et al., Local human movement patterns and land use impact exposure to zoonotic malaria in Malaysian Borneo. Elife, 2019. 8. 355 356 17. Wong, M.L., et al., Seasonal and Spatial Dynamics of the Primary Vector of Plasmodium knowlesi within a Major Transmission Focus in Sabah, Malaysia. PLoS 357 358 Negl Trop Dis, 2015. 9(10): p. e0004135. Routledge, I., et al., Estimating spatiotemporally varying malaria reproduction 359 18. 360 numbers in a near elimination setting. Nat Commun, 2018. 9(1): p. 2476. Imai, N., et al., Transmission and control of Plasmodium knowlesi: a mathematical 361 19. 362 modelling study. PLoS Negl Trop Dis, 2014. 8(7): p. e2978. Hawking, F., et al., Transmission of Plasmodium knowlesi by Anopheles stephensi. 363 20. 364 Trans R Soc Trop Med Hyg, 1957. 51(5): p. 397-402. Chin, W., et al., Experimental mosquito-transmission of Plasmodium knowlesi to man 365 21. 366 and monkey. Am J Trop Med Hyg, 1968. 17(3): p. 355-8.

- Fornace, K.M., et al., Local human movement patterns and land use impact exposure
  to zoonotic malaria in Malaysian Borneo. eLife, 2019. 8(10): p. 22.
- Routledge, I., et al., *Tracking progress towards malaria elimination in China: Individual-level estimates of transmission and its spatiotemporal variation using a diffusion network approach.* PLoS Comput Biol, 2020. **16**(3): p. e1007707.
- Routledge, I., H.J.T. Unwin, and S. Bhatt, *Inference of malaria reproduction numbers in three elimination settings by combining temporal data and distance metrics*. Sci Rep, 2021. **11**(1): p. 14495.
- 375 25. Gomez-Rodriguez, M., D. Balduzzi, and B. Scholkpf, *Uncovering the temporal*376 *dynamics of diffusion networks.* Proceedings of the 28th International Conference on
  377 Machine Learning, 2011.
- Lindgren, F. and H. Rue, *Bayesian Spatial Modelling with R-INLA*. Journal of
  Statistical Software, 2015. 63(19).