Title

Cataract, abnormal electroretinogram and visual evoked potentials in a child with SMA-LED2 - extending the phenotype

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Abstract

This case report describes a girl who presented antenatal arthrogryposis and postnatal hypotonia, generalized and respiratory weakness, joint deformities particularly affecting the lower limbs and poor swallow. By 5 months, cataracts, abnormal electroretinograms, visual evoked potentials and global developmental impairments were recognized. No causative variants were identified on targeted gene panels. After her unexpected death at 11 months, gene-agnostic trio whole exome sequencing revealed a likely pathogenic *de novo BICD2* missense variant, NM_001003800.1, c.593T>C, p.(Leu198Pro), confirming the diagnosis of spinal muscular atrophy lower extremity predominant type 2 (SMA-LED2). We propose that cataracts and abnormal electroretinograms are novel features of SMA-LED2.

Introduction

Arthrogryposis describes congenital contractures in two or more different joints and affects 1 in 3,000 live births. Over 400 conditions present with arthrogryposis; however, regardless of the specific cause, the underlying pathogenesis includes a reduction in fetal movements, leading to contractures (1). Next generation sequencing now enables a genetic diagnosis to be reached in up to 60% of cases (2).

We describe an infant with antenatal arthrogryposis, post-natal hypotonia and cataracts, abnormal electroretinograms (ERGs), visual evoked potentials (VEPs) and global developmental impairments recognized over time. Gene-agnostic trio whole exome sequencing (WES) identified a *de novo* likely pathogenic variant in *BICD2*, which, to our knowledge, has not been described as causing cataract, ERG or VEP abnormalities.

Case report

This female baby was born at 36+2 weeks of gestational age to a healthy non-consanguineous Scottish couple. Antenatal ultrasound at 21 weeks revealed intrauterine growth restriction and arthrogryposis. She was delivered by elective caesarean section with Apgar scores of 5 and 8 at 1 minute and 5 minutes, respectively, and weighing 1.78kg (-4.45 SDS). She had frontal bossing, upslanting palpebral fissures (Figure 1A,B) and generalized hypotonia with reduced muscle bulk, particularly in the lower limbs. Finger creases were present and there was no dimpling over joints. Her shoulders were internally rotated with mild adduction deformities. She had mild elbow flexion contractures (10˚) and restricted extension (to 150˚). Her wrists could not be passively flexed beyond the neutral position. She had mild flexion deformities of proximal interphalangeal joints on the left and left fifth finger clinodactyly. Her hips were dislocated bilaterally, there was a flexion contracture (20˚) of the right knee, bilateral vertical talus and ankle hyper-dorsiflexion bilaterally. There were little movements in facial muscles

and flickers of movement only in the lower limbs, whilst upper limbs movements were part-range antigravity. She required extensive respiratory support, tracheostomy and percutaneous endoscopic gastrostomy due to unsafe swallow and gastro-oesophageal reflux.

Ophthalmology review at three months, delayed by illness, found exotropia, roving nystagmus and bilateral cataracts (dense lamellar on the left and more peripheral on the right). Ultrasound showed grossly normal features of both eyes and the dimensions were within normal range. VEPs at five months were present binocularly and monocularly, but with delay of the predominant peak; the right eye VEP was more complex and robust than the left eye VEP. ERGs under light-adapted conditions were present, but small and delayed with an atypical waveshape (Figure 2). Given the electrodiagnostic results and the differences in the appearance of both cataracts, bilateral simultaneous surgery was performed. She was aphakic post-operatively with refractive correction initially provided with contact lenses and then prescription glasses. Visual impairment remained; however, her nystagmus lessened in frequency. There was light perception post-operatively in both eyes.

Over time, global developmental delay became evident. Although auditory brainstem evoked potentials were normal, she remained non-verbal with a pre-intentional level of communication. She did not achieve head control, nor independent sitting. After intensive physiotherapy, she was able to bring both hands together for finger play while supine. She developed flickers of hip adduction, abduction, and flexion.

A trial of pyridostigmine was performed without any clinical benefit. Nerve conduction studies were within normal limits (Supplementary Table 1).

Creatinine kinase was normal (28 IU/L). There were no atypical transferrin glycoforms at 4.5 months. Brain MRI aged 1 month showed normal brain parenchyma, splaying of the frontal horns of lateral ventricles and absent myelination in the posterior limb of internal capsule

consistent with delayed myelination due to prematurity (Figure 1C). Echocardiography revealed a small apical ventricular septal defect.

She died unexpectedly at 11 months, due to refractory status epilepticus. Despite broad-spectrum antibiotics, antiviral therapy and inotropes she suffered two cardiac arrests. Her seizures failed to respond to appropriate, timely anti-seizure medication management and after discussion with her parents, active treatment was discontinued and she passed away peacefully.

Post-mortem findings were consistent with infection at the time of death. There was acute bronchopneumonia and positive norovirus antibody titres (Ct value 30). Cortisol levels were very high (>3,300nmol/l). Macroscopic examination of the brain was normal.

SNP-array showed a paternally-inherited 3.7 Mb deletion at 13q31.3-q32.1 containing eleven protein-coding HGNC genes, none associated with cataract. Only four of these were morbid genes, all for autosomal recessive disorders: *CLDN10* (Helix Syndrome, OMIM #617671), *DNAJC3* (cerebellar and peripheral ataxia with hearing loss and diabetes mellitus, OMIM #616192), *GPC6* (omodysplasia type 1, OMIM #258315) and *TGDS* (Catel-Manzke syndrome, OMIM #616145). She tested negative for the myotonic dystrophy type 1 repeat expansion, and sequencing of congenital myasthenic syndrome genes revealed no causative variants. Testing of a clinical exome panel of 107 neuromuscular arthrogryposis genes was initiated and customized, when the cataract was recognized, to include genes known to cause both cataracts and arthogryposis (*ALG14, ALG3, COG6, COG7, COG8, DPACGT1, DYNC1H1, ERCC5, HRAS, INPP5K*), but no causative gene variants were found. She was heterozygous for a pathogenic *LAMA2* variant c.2049_2050del, p.(Arg683Serfs*21)). After her death, trio WES using an inheritance-based, gene-agnostic approach revealed a heterozygous *de novo BICD2* missense variant (NM_001003800.1, c.593T>C, p.(Leu198Pro)), confirmed on Sanger sequencing and classified likely pathogenic (ACMG criteria: PS2 moderate, PM2 moderate,

PP3 supporting, PP4 supporting (3)). Analysis of the PanelApp Cataracts version 2.3 gene panel was applied and excluded a reduced penetrance heterozygous inherited variant (4).

Discussion

Dominant variants in *BICD2* (BICD Cargo Adaptor 2, #OMIM 609797) cause spinal muscular atrophy, lower extremity-predominant type 2 (SMA-LED2, #OMIM 615290) (5-7). Clinical severity varies and includes severe, antenatally lethal forms, congenital forms with or without contractures, and childhood and adult-onset lower extremity weakness (8-11). *De novo* variants in *BICD2* have been specifically shown to be correlated with more severe clinical features including arthrogryposis and ealy fatal outcome (8, 10, 12).

Retrospectively, the disparity between our patient's severe lower limb weakness and much milder upper limb weakness is characteristic of SMA-LED and similar disparity is evident in her neurophysiology, where CMAPs were much smaller in lower than upper limbs, despite being within normal limits for age.

Upper motor neuron involvement and neurocognitive involvement were highlighted in the initial description of SMA-LED2 (6). Subsequently, cerebellar hypoplasia and perisylvian polymicrogyria were described (10). Although no structural brain changes were identified in our patient, we presume her neurodevelopmental impairment relates to central involvement.

Our patient's abnormal light-adapted single flash ERGs were similar in morphology to those seen in congenital disorder of glycosylation type Ia (CDG1a, OMIM #212065) (13), however is unlikely that this child had co-existing CDG1a because her neurological features were atypical, transferrin glycoforms were normal and no pathogenic variants in *PMM2* (OMIM #601785) were identified on WES.

We propose that cataract and abnormal retinal function may be features of *BICD2*-related disorders, as no alternative cause of these was found on extensive genetic testing. Furthermore, early onset cataracts have been described in patients with variants in *DYNC1H1* (OMIM #600112) (14), which encodes the cytoplasmic heavy chain 1 of dynein, an important binding partner of BICD2. *DYNC1H1* variants cause a range of phenotypes including SMA-LED1 (14, 15), which is clinically strikingly similar to SMA-LED2 (11), therefore it seems reasonable that this similarity could extend to include cataract, especially since our patient's variant lies in the *BICD2's* dynein binding domain (CC1) (16).

Moreover, as a golgin and cargo adaptor protein (7), linking the dynein-dynactin motor complex with its intracellular cargo (18), BICD2 plays a role in retrograde axonal transport, vesicular transport, Golgi integrity and synaptic vesicle recycling (7), any or all which could be affected by our patient's variant.

Although no studies have directly shown that, both *DYNC1H1* and *BICD2* are expressed in the retina (20) and during retinal development (21). *DYNC1H1* is expressed in the elongating fiber cells of rats' lens where it is suggested to play a role in trafficking of vesicles along microtubules during development (19) and a non-sense *DYNC1H1* variant has been shown to impair photoreceptor development in zebrafish (22). The role of Bic-D in microtubule-based transport was also shown to be essential for photoreceptor nuclear migration during eye development in *Drosophila* (23). A role for *BICD2* in eye development is further suggested by the finding that another of its binding partners, Rab6 (24), is involved in trafficking rhodopsin in post-Golgi vesicles during photoreceptor development and maintenance (25, 26).

Finally, since eye involvement has not been described in SMA-LED2 before, it may be a rare feature. It is possible that in more severely affected cases their clinical severity has precluded detailed formal assessment.

The intractable seizures at the end of our patient's life were unexpected and occurred in the context of a pyrexial illness. Although seizures have been described in three other SMA-LED2 cases, two had structural brain malformations and one had severe ischaemic brain injury (8, 10, 12). Seizures have also been described in patients with *DYNC1H1* variants (14, 28). Our patient's seizures could be entirely unrelated to the SMA-LED2 or reflect a non-specifically reduced seizure threshold because of her central involvement. Nonetheless, it is notable that the seizures were resistant to standard therapy and it may be reasonable to maintain a low clinical index of suspicion for seizures in SMA-LED2.

The rapid terminal decompensation in our patient could raise the possibility of a second, metabolic, disorder (29). However, neither biochemical testing, nor WES found any evidence for this.

This patient had extensive genetic testing in life, with only a heterozygous pathogenic *LAMA2* variant detected, which is presumed to represent incidental carrier status since her clinical phenotype and normal CK make merosin-deficient congenital muscular dystrophy (MDC1A, OMIM #607855) most unlikely (30). This illustrates the difficulty in selecting targeted gene panels in cases where the clinical phenotype is still evolving and highlights the utility of gene-agnostic trio WES as a first line investigation in such cases.

In conclusion, we believe this is the first report of eye involvement in SMA-LED2 and suggest that cataract and abnormal ERG waveforms may be features of the condition in some cases.

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Conflicts of interest

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References

1. Hall JG. Arthrogryposis (multiple congenital contractures): Diagnostic approach to etiology, classification, genetics, and general principles. European Journal of Medical Genetics. 2014;57(8):464-72. doi: 10.1016/j.ejmg.2014.03.008

2. Bayram Y, Karaca E, Coban Akdemir Z, Yilmaz E, Tayfun G, Aydin H, et al. Molecular etiology of arthrogryposis in multiple families of mostly Turkish origin. Journal of Clinical Investigation. 2016;126(2):762-78. doi: 10.1172/Jci84457

3. Ellard S, Baple EL, Callaway A, Berry I, Forrester N, Turnbull C, et al. ACGS Best Practice Guidelines for Variant Classification in Rare Disease 2020: Association for Clinical Genomic Science (ACGS); 2020 [09/04/2021]. Available from: https://www.acgs.uk.com/media/11631/uk-practice-guidelines-for-variant-classification-v4-01-2020.pdf.

4. Martin AR, Williams E, Foulger RE, Leigh S, Daugherty LC, Niblock O, et al. PanelApp crowdsources expert knowledge to establish consensus diagnostic gene panels. Nature Genetics. 2019;51(11):1560-5. doi: 10.1038/s41588-019-0528-2

5. Peeters K, Litvinenko I, Asselbergh B, Almeida-Souza L, Chamova T, Geuens T, et al. Molecular defects in the motor adaptor BICD2 cause proximal spinal muscular atrophy with autosomal-dominant inheritance. American journal of human genetics. 2013;92(6):955-64. doi: 10.1016/j.ajhg.2013.04.013

6. Oates Emily C, Rossor Alexander M, Hafezparast M, Gonzalez M, Speziani F, MacArthur Daniel G, et al. Mutations in BICD2 Cause Dominant Congenital Spinal Muscular Atrophy and Hereditary Spastic Paraplegia. Am J Hum Genet. 2013;92(6):965-73. doi: 10.1016/j.ajhg.2013.04.018

7. Neveling K, Martinez-Carrera LA, Hölker I, Heister A, Verrips A, Hosseini-Barkooie SM, et al. Mutations in BICD2, which encodes a golgin and important motor adaptor, cause congenital autosomal-dominant spinal muscular atrophy. American journal of human genetics. 2013;92(6):946-54. doi: 10.1016/j.ajhg.2013.04.011

8. Koboldt DC, Waldrop MA, Wilson RK, Flanigan KM. The Genotypic and Phenotypic Spectrum of BICD2 Variants in Spinal Muscular Atrophy. Annals of Neurology. 2020;87(4):487-96. doi: 10.1002/ana.25704

9. Ravenscroft G, Di Donato N, Hahn G, Davis MR, Craven PD, Poke G, et al. Recurrent de novo BICD2 mutation associated with arthrogryposis multiplex congenita and bilateral perisylvian polymicrogyria. Neuromuscul Disord. 2016;26(11):744-8. doi: 10.1016/j.nmd.2016.09.009

10. Storbeck M, Horsberg Eriksen B, Unger A, Hölker I, Aukrust I, Martínez-Carrera LA, et al. Phenotypic extremes of BICD2-opathies: from lethal, congenital muscular atrophy with arthrogryposis to asymptomatic with subclinical features. European journal of human genetics : EJHG. 2017;25(9):1040-8. doi: 10.1038/ejhg.2017.98

11. Rossor AM, Oates EC, Salter HK, Liu Y, Murphy SM, Schule R, et al. Phenotypic and molecular insights into spinal muscular atrophy due to mutations in BICD2. Brain : a journal of neurology. 2015;138(Pt 2):293-310. doi: 10.1093/brain/awu356

12. Koboldt DC, Kastury RD, Waldrop MA, Kelly BJ, Mosher TM, McLaughlin H, et al. In-frame de novo mutation in BICD2 in two patients with muscular atrophy and arthrogryposis. Cold Spring Harbor molecular case studies. 2018;4(5):a003160. doi: 10.1101/mcs.a003160

13. Thompson DA, Lyons RJ, Liasis A, Russell-Eggitt I, Jägle H, Grünewald S. Retinal On-Pathway Deficit in Congenital Disorder of Glycosylation Due to Phosphomannomutase Deficiency. Archives of Ophthalmology. 2012;130(6):712-9. doi: 10.1001/archophthalmol.2012.130

14. Amabile S, Jeffries L, McGrath JM, Ji W, Spencer-Manzon M, Zhang H, et al. DYNC1H1-related disorders: A description of four new unrelated patients and a comprehensive review of previously reported variants. American Journal of Medical Genetics Part A. 2020;182(9):2049-57. doi: 10.1002/ajmg.a.61729

15. Strickland AV, Schabhüttl M, Offenbacher H, Synofzik M, Hauser NS, Brunner-Krainz M, et al. Mutation screen reveals novel variants and expands the phenotypes associated with DYNC1H1. J Neurol. 2015;262(9):2124-34. doi: 10.1007/s00415-015-7727-2

16. Matanis T, Akhmanova A, Wulf P, Del Nery E, Weide T, Stepanova T, et al. Bicaudal-D regulates COPI-independent Golgi–ER transport by recruiting the dynein–dynactin motor complex. 2002;4(12):986-92. doi: 10.1038/ncb891

17.	Hoang HT, Schlager MA, Carter AP, Bullock SL. DYNC1H1 mutations associated with neurological diseases compromise processivity of dynein-dynactin-cargo adaptor complexes. Proceedings of the National Academy of Sciences of the United States of America. 2017;114(9):E1597-E606. doi: 10.1073/pnas.1620141114

18.	Hoogenraad CC, Akhmanova A, Howell SA, Dortland BR, De Zeeuw CI, Willemsen R, et al. Mammalian Golgi-associated Bicaudal-D2 functions in the dynein-dynactin pathway by interacting with these complexes. The EMBO journal. 2001;20(15):4041-54. doi: 10.1093/emboj/20.15.4041

19.	Lo W-K, Wen X-J, Zhou C-J. Microtubule configuration and membranous vesicle transport in elongating fiber cells of the rat lens. Exp Eye Res. 2003;77(5):615-26. doi: 10.1016/s0014-4835(03)00176-3

20.	Papatheodorou I, Moreno P, Manning J, Fuentes AM-P, George N, Fexova S, et al. Expression Atlas update: from tissues to single cells. Nucleic Acids Res. 2020;48(D1):D77-D83. doi: 10.1093/nar/gkz947

21.	Mansergh FC, Carrigan M, Hokamp K, Farrar GJ. Gene expression changes during retinal development and rod specification. Molecular vision. 2015;21:61-87.

22.	Insinna C, Baye LM, Amsterdam A, Besharse JC, Link BA. Analysis of a zebrafish dync1h1mutant reveals multiple functions for cytoplasmic dynein 1 during retinal photoreceptor development. Neural Development. 2010;5(1):12. doi: 10.1186/1749-8104-5-12

23.	Houalla T, Hien Vuong D, Ruan W, Suter B, Rao Y. The Ste20-like kinase misshapen functions together with Bicaudal-D and dynein in driving nuclear migration in the developing drosophila eye. Mechanisms of Development. 2005;122(1):97-108. doi: 10.1016/j.mod.2004.08.005

24.	Will L, Portegies S, van Schelt J, van Luyk M, Jaarsma D, Hoogenraad CC. Dynein activating adaptor BICD2 controls radial migration of upper-layer cortical neurons in vivo. 2019;7(1):162. doi: 10.1186/s40478-019-0827-y

25.	Deretic D. Post-Golgi trafficking of rhodopsin in retinal photoreceptors. Eye. 1998;12(3):526-30. doi: 10.1038/eye.1998.141

26.	Deretic D, Papermaster DS. Rab6 is associated with a compartment that transports rhodopsin from the trans-Golgi to the site of rod outer segment disk formation in frog retinal photoreceptors. Journal of Cell Science. 1993;106(3):803-13. doi: 10.1242/jcs.106.3.803

27.	Matsuto M, Kano F, Murata M. Reconstitution of the targeting of Rab6A to the Golgi apparatus in semi-intact HeLa cells: A role of BICD2 in stabilizing Rab6A on Golgi membranes and a concerted role of Rab6A/BICD2 interactions in Golgi-to-ER retrograde transport. Biochimica et Biophysica Acta (BBA) - Molecular Cell Research. 2015;1853(10, Part A):2592-609. doi: 10.1016/j.bbamcr.2015.05.005

28.	Gelineau-Morel R, Lukacs M, Weaver KN, Hufnagel RB, Gilbert DL, Stottmann RW. Congenital Cataracts and Gut Dysmotility in a DYNC1H1 Dyneinopathy Patient. Genes. 2016;7(10):85. doi: 10.3390/genes7100085

29.	FitzPatrick DR. Genetic Metabolic Disease. Fetal and Neonatal Pathology London: Springer London; 2007. p. 162-83.

30.	Philpot J, Cowan F, Pennock J, Sewry C, Dubowitz V, Bydder G, et al. Merosin-deficient congenital muscular dystrophy: the spectrum of brain involvement on magnetic resonance imaging. Neuromuscul Disord. 1999;9(2):81-5. doi: 10.1016/s0960-8966(98)00110-2

Figure 1.

A

B

C

Figure 1. A, B. Clinical photographs aged 4 months. The patient has a frontal bossing (A) and upslanting palpebral fissures (B), the latter shared with her mother (not shown). Permission for publication of these photos has been granted by the parents of the patient. **C.** MRI Head: T2 sequence. There is absence of myelination in the posterior internal capsule (white arrow). There is splaying of the frontal horns of the lateral ventricles (black arrow).
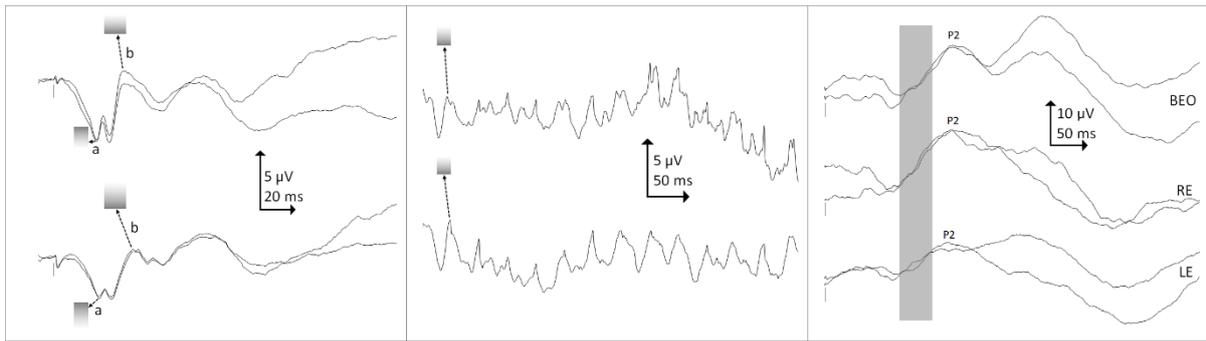
Figure 2

Figure 2. Visual electrodiagnostic findings. Left: light-adapted single flash ERGs, right eye above, left eye below. Shaded zones indicate reference limits for a- and b-waves: the patient's ERGs are smaller and slower than normal, with a pronounced oscillation at the a-wave trough. Centre: light-adapted 30 Hz flicker ERGs, right eye above, left eye below: the patient's ERGs are smaller than normal. Shaded zones indicate reference limits for flicker peak. ERGs were recorded during natural sleep using skin electrodes on the lower eyelid, with a high forehead reference (Fz). Reference limits have been adjusted to compensate for the known effect of closed lids. Right: binocular (top), right eye (middle) and left eye (bottom) flash VEPs. Shaded bar indicates normal timing for the predominant (P2) peak: the patient's VEPs are delayed. VEPs were recorded during natural sleep, active electrode at Oz referred to Fz. The stimulus was a strong (40 cd s m-2) diffused xenon flash held a few centimetres from the closed eyelids. Positive is plotted upwards.

Supplementary Table 1

	CMAPs	Velocities
Upper limb	5.7mV/8.7 mV	35/43 m/s
Lower limb	1.1 mV	30 m/s

Supplementary Table 1. Results of nerve conduction studies at 6 months of age. CMAPs - compound muscle action potentials.