

Hoes, M. F., Arany, Z., Bauersachs, J., Hilfiker-Kleiner, D., Petrie, M. C., Sliwa, K. and van der Meer, P. (2022) Pathophysiology and risk factors of peripartum cardiomyopathy. *Nature Reviews Cardiology*, (doi: 10.1038/s41569-021-00664-8)

The material cannot be used for any other purpose without further permission of the publisher and is for private use only.

There may be differences between this version and the published version. You are advised to consult the publisher's version if you wish to cite from it.

http://eprints.gla.ac.uk/263329/

Deposited on 14 January 2022

Enlighten – Research publications by members of the University of Glasgow

http://eprints.gla.ac.uk

Pathophysiology and risk factors of peripartum

cardiomyopathy

3	
4	Authors:
5 6 7 8 9 10 11	Martijn F. Hoes ^{1†} Zoltan Arany ² Johann Bauersachs ³ Denise Hilfiker-Kleiner ⁴ Mark C. Petrie ⁵ Karen Sliwa ⁶ Peter van der Meer ¹
13 14 15 16 17 18 19 20 21	 Department of Cardiology, University Medical Center Groningen, University of Groningen, Groningen, The Netherlands Department of Medicine, Cardiovascular Institute, Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA, USA Department of Cardiology and Angiology, Hannover Medical School, Hannover, Germany Dean of the medical faculty of the Philipps University Marburg, Marburg, Germany Institute of Cardiovascular & Medical Sciences, University of Glasgow, Glasgow, UK Department of Medicine & cardiology, Cape Heart Institute, Faculty of Heath Sciences, University of Cape Town, Cape Town, South Africa
23 24 25 26	† Corresponding author: Martijn Hoes (m.hoes@umcg.nl)

Abstract

1

2

4

5

6

7

9

10

11

12

13

14

15

16

Peripartum cardiomyopathy (PPCM) is a potentially fatal form of idiopathic heart failure with varying incidences among countries and races. The cause of PPCM is uncertain but it may result from a combination of environmental and genetic factors, as well as pregnancy associated conditions such as pre-eclampsia. Animal studies suggested that impaired vascular and metabolic function may be central to the development of PPCM. Clarifying the pathogenic mechanisms is necessary to establish new therapies to improve the outcomes of patients with PPCM. Pregnancy hormones tightly coordinate a plethora of maternal adaptive responses, including haemodynamic, vascular, structural, and metabolic changes of the cardiovascular system. While pregnancy is considered to be a cardiovascular challenge, hormonal effects uniquely drive systemic insulin resistance and mostly fatty acid-dependent cardiac metabolism. In PPCM, the peripartum period is associated with profound and rapid hormonal changes that result in a brief period of disrupted cardiovascular (metabolic) homeostasis prone to secondary perturbations. This review summarizes and reflects on recent literature on the potential pathophysiological mechanisms and risk factors for PPCM with a focus on the maternal cardiovascular changes associated with pregnancy. We provide an updated framework to improve understanding of PPCM pathogenesis, which may lead to a better disease definition.

18

19

20

21

22

23

24

25

17

Introduction

Peripartum cardiomyopathy (PPCM) is a form of heart failure associated with pregnancy and the postpartum period^{1,2}. PPCM is defined as an idiopathic cardiomyopathy presenting with heart failure secondary to left ventricular (LV) systolic dysfunction in the peripartum phase (i.e., towards the end of pregnancy, during delivery or in the months following delivery) where no other cause of heart failure is found^{1,3–5}. Diagnosis generally follows the exclusion of other (similar) conditions and differential diagnoses include pre-existing dilated cardiomyopathy, Takotsubo cardiomyopathy,

myocarditis, familial cardiomyopathy and valvular heart disease^{1,2,5,6}. Furthermore, outcomes varied greatly in the European Society of Cardiology (ESC) EURObservational Research Programme (EORP) PPCM Registry⁴. Myocardial recovery (i.e., LV ejection fraction [LVEF] >50%) was observed in 46% of patients 6 months after diagnosis and persisting severe LV dysfunction or death was seen in 28% of patients worldwide⁴. Previous studies have indicated that PPCM patients often suffer from hypertension and palpitations, and may have a persisting higher risk for sudden death, arrhythmia, and other cardiovascular complications. Long-term prescribed drug use is common, even in patients with fully recovered LV function^{7,8}. PPCM incidence appears to vary markedly among geographical regions, but differing definitions prevent direct comparison of studies. Countries with the lowest reported incidence (i.e., per live birth) include Japan (1 in 16 667)^{9,10}, Denmark (1 in 10 000)¹¹, and Sweden (1 in 5882)¹². In contrast, those with higher rates appear to be Nigeria (1 in 100)¹³, Haiti (1 in 333)¹⁴, Pakistan (1 in 840)¹⁵, and South Africa (1 in 1000)¹⁶. In comparison, estimated incidences in Germany are 1 in 1000 to 1500 live births¹⁷. Studies in the USA suggest an increasing incidence over the past 20 years¹⁸. Various pathophysiological mechanisms have been suggested 19-23, but their clinical relevance remains to be confirmed. A common hypothesis states that PPCM is a multifactorial syndrome where several known and unknown factors in the setting of pregnancy may lead to PPCM, i.e., a "multiple-hit model". This hypothesis is supported by the onset of PPCM in mice with a cardiac specific knockout for either the signal transducer and activator of transcription 3 (Stat3) gene or the peroxisome proliferator-activated receptor y coactivator 1α (*Pparqc1a*) gene^{19,20}. These mice developed severe heart failure postpartum, but did not present any heart failure-related symptoms before pregnancy^{19,20}. A recent study in human induced pluripotent stem cells (hiPSC) derived from patients with PPCM highlighted a role of cardiomyocyte metabolism in the pathogenesis of PPCM²⁴. Several pregnancy-associated hormones, including progesterone, oestrogens, prolactin, soluble fmslike tyrosine kinase 1 (sFLT1), and fibroblast growth factor 21 (FGF21) play roles in the coordination of cardiac metabolism^{25–27}. Impaired metabolism in PPCM patient-derived cardiomyocytes and

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

47

48

49

50

metabolic effects of pregnancy hormones may indicate that the hearts of patients with PPCM cannot cope with the profound fluctuations of hormones and downstream metabolic changes that occur in the peripartum period. This review summarizes and reflects on recent literature on the potential pathophysiological mechanisms and risk factors for PPCM with a focus on the physiological maternal changes associated with pregnancy. We provide an updated framework to improve understanding of PPCM pathogenesis, which may lead to a better disease definition.

Cardiovascular adaptations in pregnancy

PPCM is hypothesized to occur due to the interaction of an external trigger and a predisposition: a "two-hit model". While a putative predisposition remains elusive (but is likely to be genetic), far more is known about the challenges of pregnancy and the effects on the cardiovascular system that could trigger PPCM pathogenesis. Hormones are the key regulatory elements that drive the different stages and related adaptations during and after pregnancy. Maternal adaptations to the cardiovascular system include hemodynamic and structural changes, vascular remodelling, and bioenergetic shifts. These adaptive processes are necessary to prevent diseases like PPCM.

Currently, it is unknown which adaptive processes fail in the pathogenesis of PPCM. More research is needed to identify these potentially insufficient mechanisms in PPCM. However, this section provides a basis for such studies by summarizing what is known about physiological pregnancy-related adaptation from a cardiovascular perspective.

Haemodynamic changes

Pregnancy is associated with an increasing blood volume that leads to a chronically elevated cardiac volume load²⁸. As a result, cardiac output increases to a prolonged peak from the second trimester to term and corresponds to an increased heart rate by \sim 20% and stroke volume by \sim 25%^{29–32}. Increases in stroke volume were also found to be higher in subsequent pregnancies compared with

the first pregnancy²⁹. Vascular resistance also falls by ~30% in the first trimester and recovers after delivery^{29–32}. Gestational blood pressures were previously undefined, but a recent multicentre, longitudinal study in 4,279 women demonstrated that median systolic and diastolic pressures briefly declined during early pregnancy, but rose by 7 mmHg and 9 mmHg, respectively, above nominal pressures by late gestation³². These changes appear minor and could explain the inconsistency of previous studies. It is unknown whether haemodynamic changes could lead to PPCM, but low systolic blood pressure and elevated heart rate were associated with worse outcome in patients with PPCM³³.

85

86

87

88

89

90

91

92

93

94

95

96

97

98

99

100

101

102

77

78

79

80

81

82

83

84

Structural changes

Parallel to haemodynamic changes, the human maternal heart undergoes substantial remodelling. Both left and right ventricular end diastolic diameters (LVEDD and RVEDD, respectively) increased by ~20%, whereas the left end systolic diameter (LVESD) did not change between the third trimester and postpartum³³. A meta-analysis of 48 studies indicated that LV mass was about 28% higher in the last trimester of normotensive pregnancy³⁴. These observations are indicative of gestational cardiac hypertrophy. Of note, cardiac dimensions and estimated weights were often compared to postpartum time points. While these structural changes are known to be transient, it is unknown whether heart dimensions can fully return to baseline (i.e., pre-pregnant) or the time required to do so. Additionally, several histological studies in rodents have indicated that the extensive cardiac remodelling does not involve fibrosis during or after pregnancy^{35–37}. However, similar histological studies have not been performed in healthy women pre- or postpartum as these are limited by the requirement of cardiac biopsies and the associated risks. PPCM can have various cardiac phenotypes including ventricular dilation¹, borderline non-compaction cardiomyopathy^{38,39}, and peripartum takotsubo cardiomyopathy^{40,41}, whereas normal pregnancy is associated with reversible eccentric cardiac hypertrophy⁴². This disparity may indicate that regulatory mechanisms involved in physiological cardiac remodelling during and after pregnancy could be impaired, leading to a

decompensated phenotype. Genetic variants of structural genes have been associated with PPCM and are discussed in detail in the *Risk factors* section.

105

106

107

108

109

110

111

112

113

114

115

116

117

118

119

120

121

122

123

124

125

103

104

Vascular remodelling

The balance between cardiac hypertrophy and vascular growth is crucial to maintain adequate cardiac function during pregnancy. In concert with increased ventricular mass, the vasculature is required to adapt accordingly. Like fibrosis, data on vascular changes is mostly available from rodent studies. It was shown that capillary density is transiently increased in mice in late pregnancy^{19,35}. Specifically, pro-angiogenic gene (including Vegf, Ppargc1a, angiopoietin-1, and Fgf2) are activated in early and mid-gestation, but return to non-pregnant levels in late gestation⁴³. This is in line with the observed antiangiogenic environment associated with late gestation²⁰. These findings in rodents corresponded with serum levels of PIGF, which reached a peak in the second trimester as well before returning to baseline levels in the last trimester in humans⁴⁴. In contrast, circulating VEGF appears to be stable in the first two trimesters before increasing near term⁴⁵. Like VEGF, serum levels of soluble VEGF receptor-1 (sFlt-1) were elevated in late pregnancy⁴⁶⁻⁴⁸. Since sFlt-1 readily binds circulating VEGF, it is unknown whether the elevated levels of VEGF reflected levels of free VEGF or inactivated VEGF that is bound to sFlt-1. Hence, this may be a physiological response to maintain an angiogenic balance systemically and locally²⁰. Disruption of this delicate balance is a key factor in the development of pre-eclampsia and is likely also involved in the pathogenesis of PPCM. Specifically, mice with cardiac ablation of the Ppargc1a gene (which encodes the transcription factor PGC-1α) developed a PPCM-like phenotype following inhibition of VEGF signalling²⁰. Vascular function and remodelling in the peripartum period are key aspects of PPCM pathophysiology and is discussed in more detail in the following sections.

126

127

Maternal cardiac metabolism during pregnancy

The maternal heart undergoes unique bioenergetic changes during pregnancy, which is tightly regulated during each gestational phase. In a normal, non-pregnant, fasted state, the human heart primarily utilizes free fatty acids (FFA) as a source of fuel^{49,50}. Other metabolic substrates include ketones, lactate, and amino acids^{49,50}. While glucose is one of the principal metabolic substrates for most human tissues, recent studies demonstrated that the heart consumes very little in the average population, at least in the fasting state^{49,50}. As pregnancy progresses, maternal metabolism shifts from a predominant anabolic state with increased fat stores to a catabolic state with reduced fat mass and elevated levels of circulating FFA to meet the energetic needs of the foetus⁵¹. The transition from an anabolic state to a catabolic state is characterized by a profound increase of basal metabolic rates in mothers by up to 60%⁵². Insulin signalling plays a pivotal role in coordinating this shift. Insulin resistance gradually develops with gestation and results in hyperglycaemia and hyperinsulinemia in late pregnancy^{53,54}. Consequently, glucose uptake is limited in the maternal body and is shunted to the foetus. Little is known about how cardiac metabolism changes during pregnancy in humans, but animal studies have provided insight into the associated molecular mechanisms. Early studies in rats showed a reduction in cardiac glucose oxidation by ~75% during pregnancy⁵⁵, and studies in dogs indicated similar suppression of glucose use and a near doubling of FFA oxidation during late pregnancy⁵⁶. Despite this metabolic shift towards FFA oxidation, and in contrast to insulin resistance in the liver and skeletal muscle, the hearts of mice in late pregnancy retain insulin sensitivity (defined as activation of signalling cascade)⁵⁷. The causes for these metabolic changes is incompletely understood, but likely include inhibition of glycolysis by high levels of FFA in late pregnancy according to the Randle cycle⁵⁸, and specific cellular reprogramming through hormonal signalling, such as induction by progesterone of PDK4, an endogenous inhibitor of PDH and thus of carbohydrate use (Figure 1)⁵⁹. Understanding how cardiac bioenergetics are regulated is crucial to understanding the underlying mechanisms of heart diseases in general. Deletion of PGC- 1α resulted in angiogenic imbalance, but PGC1 α is also a key regulator of major metabolic pathways, especially related to fatty acid oxidation²⁰. In vitro studies have also indicated

128

129

130

131

132

133

134

135

136

137

138

139

140

141

142

143

144

145

146

147

148

149

150

151

152

that PPCM patient-derived cardiomyocytes demonstrated reduced viability and metabolic flexibility upon inhibition of lipid metabolism²⁴. Hence, impaired metabolic regulation may be a central aspect of the development of PPCM.

157

158

159

160

161

162

163

164

165

166

167

168

169

170

171

172

173

174

175

176

177

178

179

156

154

155

Cardiovascular effects of hormones

Sex and pregnancy-related hormones are the key modulators of the various stages of pregnancy. Several hormones are known to profoundly affect the cardiovascular system, but their specific molecular mechanisms and pathways are largely unknown. A plethora of association studies are available on hormone levels and effects regarding the pregnancy and foetal status, but most cardiovascular mechanisms have been demonstrated in animal models (Figure 2). Oestrogens are a class of sex hormones that govern the development of the female reproductive system as well as pregnancy. Oestrogen levels increase progressively during pregnancy and instantly decrease after delivery⁶⁰. Cardiovascular effects of oestrogens are pleiotropic and exert mainly cardiovascular protective effects⁶¹. Oestrogens induce angiogenesis and vasodilation through increased NO synthesis^{62,63} and secretion of VEGF and PIGF^{64,65}. Additionally, oestrogens were found to reduce inflammatory signalling, attenuate cardiac hypertrophy, and are protective against oxidative stress in endothelial cells and cardiomyocytes^{66–68}. Many of its protective effects are derived from the potent inhibition of apoptosis in cardiomyocytes and endothelial cells^{69,70}. It was recently demonstrated that a related class of receptors, the oestrogen-related receptors (ERRs), widely regulate cardiac metabolism, contractility, and conduction properties⁷¹. Like oestrogens, progesterone is primarily produced by the placenta during pregnancy with increasing serum levels toward delivery. Progesterone was shown to protect against apoptosis by direct inhibition of the L-type voltage dependent Ca2+ channel (in dogs) and via induction of the BCL2 Like 1 gene (Bcl2l1; in mice)^{72,73}. Furthermore, eNOS mediated NO synthesis is enhanced after progesterone stimulation in the endothelium, causing a marked reduction in vascular resistance in pregnant rats and humans^{74,75}. Recent studies in animals have demonstrated that progesterone can

inhibit glycolysis via Forkhead box protein O1 (FOXO1)-mediated mechanisms in tumors^{76,77}. In cardiomyocytes, progesterone induced pyruvate dehydrogenase kinase (PDK4) activity, which inhibits pyruvate dehydrogenase, an essential step in glycolysis⁵⁷. Prolactin has been widely associated with PPCM pathogenesis and is discussed in detail in the section on Pathophysiology. Serum levels peak at term and rapidly fall to pre-pregnancy levels after delivery if it is not repeatedly stimulated by breastfeeding⁷⁸. Cardiovascular effects of prolactin include a blunted response to angiotensin in rats⁷⁹, endothelial pro-survival signalling via the Janus activator kinase (JAK)-signal transducer and activator of transcription (STAT) signalling pathway⁸⁰, and reversed phenylephrine-induced vascular tone in rat aortic rings⁸¹. Clearly such potential effects of prolactin are dose-related, and high dosages/concentrations often used in those studies preclude firm conclusions on the role of prolactin in humans. A lesser-known pregnancy-related hormone is FGF21, which is mainly produced by the liver during pregnancy under the control of PPAR- α^{82-84} . The vast majority of studies were done in animals, as reflected in the following section. In addition, the heart is a target and a source of FGF2185. Downstream effects of FGF21 signalling in the heart are related to protection against pathological hypertrophy and damage following myocardial infarction^{85,86}. Remarkably, cardiac remodelling was absent in pregnant FGF21 knockout mice and FFA oxidation was significantly reduced²⁶. Most of these mechanisms remain to be confirmed in humans, but FGF21 has been correlated to maternal body mass index and adiposity⁸⁷. Moreover, fasting glucose levels also inversely correlated with FGF21, which may reflect maternal nutrient status in pregnancy87. By extension, FGF21 has also been suggested as a biomarker for gestational diabetes mellitus and type 2 diabetes mellitus⁸⁸.

Biomarkers and risk factors

180

181

182

183

184

185

186

187

188

189

190

191

192

193

194

195

196

197

198

199

200

201

202

203

204

205

This section briefly discusses biomarkers that support the diagnosis of PPCM and could result from underlying disease mechanisms. Furthermore, while the cause of PPCM is currently unknown, several risk factors have been proposed, including heart failure-associated genetic defects⁸⁹,

ethnicity^{4,90}, hypertensive disorders⁹¹, infections⁹², twin and subsequent pregnancies⁹³, and previous cancer⁹⁴.

Biomarkers

Several studies have determined whether specific biomarkers were associated with PPCM, which were summarized by Cherubin et al⁹⁵. The authors evaluated 117 biomarkers from 31 case-control studies. Several biomarkers were identified as be independent risk factors for PPCM. See **Table 1** for an overview of biomarkers. A quantitative meta-analysis suggested that patients with PPCM had higher levels of natriuretic peptides, troponin, CRP, and white blood cell counts, but reduced levels of albumin and selenium compared with healthy controls⁹⁵. Note that these biomarkers mostly reflect the presence of cardiomyopathy and appear to be unspecific for PPCM. However, a few studies investigated potential PPCM-specific biomarkers by comparing patients with PPCM to patients with other types of heart disease. Increased levels of prolactin⁹⁶, miR-146a^{21,97}, and PIGF⁴⁸ were found in patients with PPCM relative to non-pregnancy-related heart failure. Additionally, the ratio between circulating sFlt-1 levels and PIGF was suggested to have significant diagnostic value for PPCM⁴⁸. Identifying more PPCM-specific biomarkers is a great unmet need and will significantly improve diagnosis and prognosis as targeted treatments can be started sooner.

Table 1 - Biomarkers as risk factors for PPCM.

Biomarker	Odds Ratio	95% confidence interval
B1R and M2R ⁹⁸	18.786	1.926 – 183.262
antimyocardial IgG ⁹⁹	2.68	1.19 – 4.85
NT-proBNP ¹⁰⁰	1.92	1.12 – 4.15
CRP ^{99,100}	1.86	1.08-4.02
Uric acid ¹⁰¹	1.3	1.049 – 1.614
ACE polymorphism ¹⁰²	0.253	0.114 – 0.558

B1R: Bradykinin B1 receptor, M2R: M2 muscarinic receptor, IgG: Immunoglobulin G, NT-proBNP: N-terminal pro b-type natriuretic peptide, CRP: C-reactive, ACE: angiotensin converting enzyme.

Genetics

As PPCM and dilated cardiomyopathy (DCM) have similar clinical characteristics, and it may be that PPCM is part of the spectrum of DCM. Some patients diagnosed with PPCM may have had a previously unrecognised dilated cardiomyopathy, although in the few documented cases where echocardiography was incidentally available prior to clinical diagnosis of PPCM, ejection fraction was normal¹⁰³. Recent studies indicated that genetic variants of in Titin (TTN)^{89,104–107}, cardiac Troponin C (TNNC1)¹⁰⁸, Desmoplakin (DSP)^{89,107}, Lamin A/C (LMNA)^{89,107}, BAG Cochaperone 3 (BAG3)^{89,107}, Filamin C (FLNC), Myosin Heavy Chain 6 and 7 (MYH6 and MYH7)⁸⁹, and Vinculin (VCL)^{89,107} were identified in both PPCM and DCM. Truncating variants of TTN were found in 10% of patients with PPCM. Mutations in DSP, FLNC, and BAG3 which were previously associated with DCM have now also been confirmed in PPCM patients^{89,106,107}. in addition, the frequencies with which mutations in each of these genes are found in patients with PPCM closely mirrors the same frequencies in patients with DCM, underscoring the similarity of genetic predispositions to both diseases. The association of BAG3 variants and PPCM has also prompted the hypothesis that various classes of molecular chaperones (e.g., heat-shock proteins) could be involved in the pathogenesis of PPCM¹⁰⁹. A small genome wide association study identified enrichment of a single nucleotide polymorphism near the Parathyroid Hormone Like Hormone (PTHLH) gene in 79 PPCM patients, although this observation requires confirmation¹¹⁰. PPCM and DCM may be caused by similar gene variants, of which TTN mutations seem to be most prevalent. However, how these various mutations converge and lead to PPCM remains to be investigated.

248

249

250

251

252

253

254

229

230

231

232

233

234

235

236

237

238

239

240

241

242

243

244

245

246

247

Geographical variation

Very little is known about the geographical variation on PPCM, despite global initiatives like the ESC EORP. Most studies did not select patients using consecutive screening (i.e., patients were selected based on PPCM diagnosis) and too few countries were affiliated with these studies. Ideally, a registry could be started that includes patients based on consecutive screening (e.g., include all pregnant women and note incidence of PPCM) and is performed consistently in as many countries as possible.

However, this is an ambitious endeavour that is has not been initiated yet and conclusive data on incidence rates remains limited by regional studies. Consequently, this section summarizes what is known from local studies (with non-consecutive screening) and indicates which factors could be considered if a global registry is initiated with consecutive patient inclusion. Recent studies demonstrated that the incidence of PPCM varies among geographical regions, with the lowest reported rates in several European and Asian countries¹⁰. PPCM incidence is highest in Nigeria and Haiti^{13,14,111}. These geographical hotspots support the hypothesis that a specific genetic background may underlie the disease although environmental factors are also likely. The recently concluded PEACE registry in Nigeria was a national consecutive study and indicated that selenium deficiency and malnutrition were significantly associated with PPCM^{111,112} and selenium supplementation could be beneficial in the treatment of PPCM¹¹³. Micronutrient deficiency and malnutrition in general are examples environmental factors that could predispose to heart failure¹¹⁴ and could trigger PPCM. Selenium deficiency is also common in neighbouring regions of Nigeria and in the Keshan region in China, but PPCM incidence rates are unknown for these regions. In contrast, studies from the USA that encompass a diverse population within the same healthcare system corroborate that race is an important risk factor. Multiple nationwide studies from the USA have demonstrated that over 40% of patients were African-American and 35% were Caucasian 18,90,115,116, in contrast to population estimates of 60.3% non-Hispanic Caucasian and 13.4% African American¹¹⁷. Two independent US studies demonstrated fundamental differences between Caucasian and African American patients, as African American patients were younger, had a higher prevalence of gestational hypertension, had a lower LVEF at diagnosis, and functional recovery was less likely or more slowly in African American patients compared with Caucasians 118,119.

277

278

279

280

255

256

257

258

259

260

261

262

263

264

265

266

267

268

269

270

271

272

273

274

275

276

Socio-economic status

Since the ESC EORP is a global study, healthcare systems differ among included countries which may skew the results. An interim report of the ESC EORP indicated that patients from countries with

middle to high health expenditure were 64.6% Caucasian compared with 5.1% Black¹²⁰. However, patients from countries with a predominantly low health expenditure were Black or Asian (45.2% and 39.4% respectively)¹²⁰. In this regard, low socio-economic status (specifically lower education) was associated with worse outcomes independent of race¹²¹. Despite marked differences in socio-economic background, the mode of presentation was largely similar. Isogai et al. compared PPCM incidence with all-cause maternal mortality per country and found a significant correlation (Spearman correlation: 0.80)¹⁰. PPCM is a major cause for maternal death and likely drives such increased maternal mortality rates. In general, maternal mortality rates are relatively high in low-income countries due to suboptimal treatment regimens and low hospitalization rates. Additionally, birth rates are lower for high-income countries versus low-income countries. Suboptimal healthcare (i.e., lack of genetic screening), more subsequent pregnancies, and a lack of contraception and family planning may indirectly contribute to the high incidence of PPCM in specific regions^{122,123}.

Pre-eclampsia and vascular dysfunction

Pregnancy-associated hypertension and its more severe form pre-eclampsia, and PPCM are both cardiovascular diseases that can affect women during late-gestation. Pre-eclampsia is defined as new-onset hypertension and proteinuria or new-onset hypertension with end-organ dysfunction with or without proteinuria after 20 weeks of gestation. The exact relationship between PPCM and pre-eclampsia is not fully understood, but pre-eclampsia strongly predisposes to PPCM^{124,125}.

Whether the increased cardiovascular risk is due to direct consequences of the underlying cause of pre-eclampsia or due to shared risk factors is currently unknown^{91,126,127}. A substudy of the ESC EORP described the differences in phenotypes and outcomes of PPCM patients with and without hypertensive disorders, including pre-eclampsia⁹¹. Patients with PPCM and pre-eclampsia presented with worse symptoms, but the LVEF of women with both diseases was more likely to recover than in PPCM patients without hypertension⁹¹. Patients with PPCM and pre-eclampsia were more likely to have peripheral oedema, pulmonary rales, a high body mass index (BMI), short QRS durations, and

New York Heart Association (NYHA) class IV symptoms⁹¹. One reason for improved outcomes in patients with PPCM and pre-eclampsia may be that these patients are diagnosed and treated earlier in disease progression¹²⁸.

Pre-eclampsia is in part caused by impaired placental function resulting in excessive levels of circulating angiostatic factors such as sFlt-1 and placental growth factor (PIGF) secreted by the placenta^{46,129}. An excess of sFlt-1 was shown to inhibit vascular endothelial growth factor (VEGF)-induced vasodilation, reduce capillary density, and cause endothelial dysfunction^{20,46,130}. Similarly, increased levels of sFlt-1 and 16kDa prolactin were also associated with PPCM patients^{20,131}. Mouse models for PPCM have indicated that the levels of 16kDa prolactin induced similar vascular dysfunction^{19,20}. However, this mechanism remains to be confirmed in humans.

Immune responses

PPCM is also associated with specific immune responses (possibly following viral infections¹³²) that may increase susceptibility or result in worse outcomes. Serum markers related to inflammation (i.e., C-reactive protein [CRP], tumor necrosis factor-alpha [TNF-α], and interleukin 6 [IL-6]) were significantly increased in PPCM patients compared with controls¹³³. A recent substudy of the IPAC study determined that IL-22 and TNF-α were associated with adverse outcome and IL-22 and IL-17 corresponded with disease severity, whereas IL-2 and IL-4 correlated with recovered LVEF at 12 months postpartum¹³⁴. Circulating NK cells were reduced while specific subsets of T cells were increased early postpartum in PPCM patients versus pregnancy matched controls¹³⁵. Recovery of immune cell levels was generally quick, but recovery of NK cells was delayed particularly in black women¹³⁵. Additionally, autoantibodies against troponin I or cardiac sarcomeric myosin were also found in patients and correlated with lower LVEF and reduced cardiac recovery at follow-up¹³⁶. Notably, the cause for (auto)immune responses can be variable and remains to be specified.

Subsequent pregnancies

Subsequent pregnancies pose an increased risk for recurrence or worsening of heart failure in patients with PPCM^{137,138}. Unrecovered left ventricular function at the time of a subsequent pregnancy was associated with a higher risk of a fatal outcome regardless of age, gravidity, parity, hypertension, and smoking¹³⁸. Study parameters varied among studies, but the consensus is that all subsequent pregnancies were associated with significantly reduced LVEF regardless of LVEF recovery after the index pregnancy^{138,139}. Mortality following the subsequent pregnancy was significantly higher in women with persistently impaired LVEF (<50%) compared with women with recovered LVEF^{138,139}. Therefore, PPCM could result in persisting subclinical cardiac dysfunction and subsequent pregnancies may aggravate cardiac function recurrently. It is not clear to which extent this deterioration continues, but is likely that cardiac function will decline continuously with each subsequent pregnancy^{140,141}.

Cancer

The prevalence of cancer was also indicated to be 16-fold higher in PPCM patients compared to agematched women⁹⁴. 57% of patients were diagnosed with cancer prior to PPCM presentation of which 92% were treated with cardiotoxic cancer therapies, which likely contributed to deterioration of LV function when PPCM developed and delayed full cardiac recovery therafter⁹⁴. Whole exome sequencing revealed that 6 out of 14 screened patients carried potential pathogenic gene variants associated with cardiomyopathy or cancer predisposition syndromes⁹⁴. However, in a large South African PPCM cohort there was no association of cancer diagnoses with PPCM diagnosis (unpublished data). Thus, it is not yet clear whether screening for genetic variants and for cancer in PPCM patients is warranted.

Pathophysiology

Very little is known about the pathophysiology of PPCM in humans. The previously described risk factors provide some insight into the state of patients with PPCM at the time of diagnosis and at different times of follow up, but these data do not support inference regarding pathological mechanisms that eventually precipitate into PPCM. Most mechanistic data were obtained from animal models that presented a phenotype that is similar to PPCM in humans. While these models helped shape some of the clinical guidelines and pathogenic hypotheses, most putative mechanisms remain to be confirmed in humans.

364

365

366

367

368

369

370

371

372

373

374

375

376

377

378

379

380

381

382

357

358

359

360

361

362

363

Mouse models for PPCM

The two main mouse models used to study PPCM were based on cardiac-specific deletion of the Stat3 or Pparac1a gene^{19,20}. Mice with either genotype developed severe heart failure postpartum that closely resembled PPCM with increasing severity in subsequent pregnancies 19,20. Abrogation of STAT3 or PGC-1α-mediated signalling pathways resulted in an impaired response to oxidative stress related to late pregnancy and early postpartum^{19,20,142}. The PI3K-Akt pathway is thought to be cardioprotective during, but transgenic overexpression of Akt in concert with Stat3 knockout could not prevent the onset of PPCM¹⁴³. Consequently, stressed cardiomyocytes secreted the ubiquitous lysosomal protease cathepsin D following hypoxic stress, mechanical stretch, and oxidative stress in addition to regulated exocytosis 19,144,145. Extracellular cathepsin D exhibited proteolytic cleavage of the nursing hormone prolactin during the peripartum period^{19,146,147}. The produced fragment is a peptide known as 16 kDa prolactin and is classified as a vasoinhibin and part of a family of peptides that elicit antiangiogenic effects¹⁴⁸. Subsequently, the 16 kDa prolactin fragment interacted with the urokinase plasminogen activator surface receptor (uPAR) on the cell membrane of adjacent endothelial cells and endocytosis was induced by circulating plasminogen activator inhibitor-1 (PAI-1)^{21,23,149}. This mechanism effectively inhibited migration and cell cycle progression, and induced apoptosis in endothelial cells, subsequently disrupting the cardiac microvasculature 150-152. Consequently, endothelial cells secreted exosomes loaded with microRNA-146a (miR-146a), which

were taken up by surrounding cardiomyocytes²¹. MiR1-146a effectively decreased protein levels of Erb-B2 Receptor Tyrosine Kinase 4 (ERBB4) and neuroblastoma RAS viral (v-ras) oncogene homolog (NRAS) in cardiomyocytes^{21,153}. The effects on NRAS-mediated mechanisms were minimal as NRAS expression is low in cardiomyocytes. In contrast, ERBB4 mediates cardiac development and metabolic processes, but its role in PPCM pathogenesis remain undefined^{154,155}. In addition, PGC-1 α regulates the expression and production of VEGF and, therefore, facilitates angiogenesis. This pathway likely offsets the antiangiogenic effects of high sFlt-1 levels at term and cardiac deletion of PGC-1 α predisposes mice to cardiomyopathy even in the absence of pregnancy^{20,156}. Thus, impaired STAT3 and PGC-1 α -mediated mechanisms resulted in striking PPCM phenotypes in mice via an induced angiogenic imbalance and abnormal metabolic regulation. See **Figure 3** for a summary of these molecular mechanisms.

Translating pathophysiology from mice to humans

The previous section focused on the potential pathogenic pathways leading to PPCM in mouse models, but these mechanisms remain to be confirmed in humans. Results from these mouse models provided a strong basis for clinical trials of bromocriptine to study the inhibition of prolactin release in PPCM patients, which showed promising outcomes^{157,158}. Bromocriptine is a dopamine agonist that supresses prolactin release from the pituitary gland and was hypothesized to ameliorate the adverse effects of 16 kDa prolactin. However, translating the findings from mice to the human situation has proven difficult as several caveats exist. For example, the mouse models were based on cardiac-specific deletion of *Stat3* and *Ppargc1a*, which is not representative for PPCM patients. However, STAT3 is a main regulator of inflammation and STAT3 activation and protein levels were greatly reduced in hearts of patients with dilated cardiomyopathy or PPCM, which suggests that STAT3 is essential to mount an adequate response upon cardiac stress^{19,159}. One study also showed that circulating prolactin levels were elevated in PPCM patients versus pregnancy-matched control while both groups were nursing⁹⁶, but these results remain to be replicated. A study in a German

cohort demonstrated that serum levels of cathepsin D and miR-146a were also significantly elevated in PPCM patients⁹⁷. Normal levels of circulating miR-146a were observed in all PPCM patients who had already received early bromocriptine treatment⁹⁷. Additionally, the antiangiogenic effects of 16 kDa prolactin is central to the proposed pathophysiology and increased circulating levels have been shown in a few PPCM patients¹⁹. Due to the lack of quantitative assays for vasoinhibins, no reference ranges or serum levels in disease have been determined. While prolactin, cathepsin D, and miR-146a were shown to be elevated in PPCM, the role of 16 kDa prolactin remains a topic of debate since cathepsin D produces five distinct vasoinhibins of which four are potent antiangiogenic agents and should be investigated further ^{147,160}. Moreover, human prolactin is not readily cleaved by cathepsin D in most extracellular conditions, mostly dependent on pH^{161,162}. Extrapolations from animal models may be difficult regarding the specific role of prolactin, but the notion of angiogenic imbalance in PPCM pathophysiology remains promising¹⁶³. The systemic antiangiogenic state during late pregnancy and early postpartum is negated by local VEGF production in the hearts of mice²⁰. The effects of excessive levels of sFlt-1 were significantly associated with vascular dysfunction and pre-eclampsia in humans¹³⁰, which supports the hypothesis that PPCM may be a predominantly vascular disease of the heart. While angiogenic therapies were beneficial in pre-eclamptic rats and PPCM mouse models, clinical studies cannot be conducted yet as human pregnancies take much longer and trials will be complicated by vast interindividual differences^{20,164}. PPCM onset is generally later than pre-eclampsia, but angiogenic imbalance may be pivotal in both diseases. Serum levels of sFlt-1, PIGF, and the sFlt-1/PIGF ratio are used to diagnose pre-eclampsia and may also be used in the diagnosis of PPCM^{47,48}. Therefore, from a vascular perspective, PPCM and pre-eclampsia may be part of a spectrum of cardiovascular diseases associated with a vascular dysfunction.

432

433

409

410

411

412

413

414

415

416

417

418

419

420

421

422

423

424

425

426

427

428

429

430

431

Metabolic contribution to PPCM

Pregnancy has evolved as a tightly regulated process with major consequences for both the mother and child when certain aspects are disrupted. Insight into specific aberrant metabolic pathways in PPCM patients is scarce. Specific genetic factors have recently been associated with PPCM recently and it is hypothesized that an underlying (genetic) factor may cause cardiovascular distress that results in PPCM. A recent study examined the differences between hiPSC-derived cardiomyocytes from typical PPCM patients and their respective familial controls²⁴. To mimic pregnancy-related cardiac volume overload, cyclic mechanical stretch was applied to cultured cardiomyocytes. While mechanical stretch caused differential expression of 2647 genes, of which 1248 specific to the PPCM cardiomyocytes, computational pathway analysis was ambiguous. This suggested that mechanical stretch did not elicit pathological effects in PPCM cardiomyocytes. In contrast, 95 genes were differentially expressed in all stretched cardiomyocytes and in static PPCM cardiomyocytes, but not static cardiomyocytes derived from controls. The majority of enriched pathways was found to be related to lipid metabolism. Cardiac lipid metabolism is known to be reduced during cardiac stress and disease^{49,50,165}. However, aberrant pathways related to lipid metabolism in static PPCM cardiomyocytes indicated a specific predisposition that was also functionally confirmed in vitro in these hiPSC-derived cardiomyocytes and in isolated cardiomyocytes from cardiac specific STAT3 knockout mice. Further analysis indicated that the majority of differentially expressed genes are controlled by several shared transcription factors, including nuclear transcription factor Y (NFY), Sp1 transcription factor (SP1), and sterol regulatory element-binding transcription factor 1 (SREBP1). Considering these experimental results, there might be a link between unstable metabolism and endocrine regulation of cardiac metabolism during pregnancy. Most PPCM patients had no indication of cardiovascular disease prior to the onset of PPCM. Moreover, mutations in the TTN gene may result in metabolic abnormalities as well. Truncating variants of TTN mutations have been associated with PPCM^{89,106,166} and were shown to have pleiotropically detrimental to cardiac function¹⁶⁷. A recent study compared cardiomyopathy patients with and without truncating *TTN* variants and showed that these TTN variants were associated with cardiac fibrosis and mitochondrial

434

435

436

437

438

439

440

441

442

443

444

445

446

447

448

449

450

451

452

453

454

455

457

458

dysfunction¹⁶⁸. Various were enriched based on genome-wide transcriptome analysis in patients with truncating TTN variants versus patients without, including oxidative phosphorylation, carbon metabolism, pyruvate metabolism, glycolysis, and PPAR signalling¹⁶⁸. Additionally, mutations in the sarcomeric proteins troponin T and C have been shown to modify the calcium binding affinity during contraction, which resulted altered ATP consumption and increased energetic demands 169,170. Destabilizing mutations in the MYH7 gene were shown to have detrimental effects on cardiac function as well, but specifically on metabolic remodelling, glycolysis, and overall mitochondrial function¹⁷¹. It is unknown how these sarcomeric alterations might induce PPCM, but they could be considered to predispose to the disease. Pregnancy gradually introduces various cardiovascular stresses. The steady increase of stress might be slow enough for the cardiovascular system to cope with these changes, but perhaps the sudden reversal of most pregnancy-related changes after delivery presents an overwhelming challenge. In contrast, the conditions of late pregnancy may present a specific challenge in itself, which might explain the onset of PPCM during the last trimester. For example, high levels of progesterone and FGF21 could disrupt the metabolic balance in heart, leading to cardiac distress^{26,57}. Fluctuations of prolactin-derived vasoinhibins might impair the delicate angiogenic balance as well, leading to impaired vascular function that may be mediated through miR-146a^{20,21}. Since vascular function is related to the supply of metabolic substrates and hypoxia, this could induce cardiac metabolic stress as well¹⁷².

478

479

480

481

482

483

484

485

460

461

462

463

464

465

466

467

468

469

470

471

472

473

474

475

476

477

Translational opportunities and future studies

The majority of studies pertaining to PPCM resulted in correlations of clinical characteristics and biomarkers. A great unmet need remains to determine how these correlations are related to the underlying pathophysiology of PPCM. Since pregnancy is a defining aspect of PPCM, future studies will be limited to an *in vivo* design. However, it is very difficult to develop a representative animal model for a putative "multiple-hit" disease; especially PPCM considering the variety of risk factors. Determining pathological genetic factors are also limited by a varying disease definition among

countries, which will dictate the nature of patients included in cohort studies. This will be a dynamic and adaptive process before a definitive disease definition can be reached. However, studies performed by Mizuno et al. and Murashige et al. determined which metabolic substrates are used in the heart in a healthy and a diseased state^{49,50}. Such studies could be repeated in PPCM patients at the time of diagnosis, and analyses could be expanded beyond metabolomics to also include (targeted) proteomics to determine hormone levels and levels of other endocrine factors. This could be repeated during and after recovery of PPCM in order to elucidate the changes in molecular circulating profiles. Such studies would be mildly invasive and will likely be limited to postpartum PPCM patients. Additionally, since cardiac biopsies are often unobtainable, molecular mechanisms could be studied in other tissues instead, like in the skeletal muscle. While significantly different in various respects, some essential pathways are shared among skeletal tissues and could provide valuable information. Further genetic screening can be done to discover genetic variants in coding and non-coding genes. Currently, such studies are hampered by high costs, sample availability, technological, and statistical limitations. Several genetic variations have been associated with PPCM, but the combination of genetic variation differs among individuals, number of identified genes, and scarcity of PPCM patients greatly limits the statistical power to improve genetic screening at this time. Once a potential pathological factor has been identified, animal models could be developed depending on the nature of that factor. For example, some animal models are more suitable for epigenetic studies, pregnancy studies, or hormone homology. Importantly, most molecular aspects of pregnancy are conserved in placental mammals, but duration and placentation differs greatly among species. Mouse models may be unsuitable to study analogous mechanisms from human and animals with a longer gestation would be required.

508

509

510

511

486

487

488

489

490

491

492

493

494

495

496

497

498

499

500

501

502

503

504

505

506

507

Conclusions

PPCM is a complex disease with many risk factors and hypothesized aetiologies. Clear guidelines have been proposed and are regularly updated to reflect novel insights and observations. Since the

pathogenesis of PPCM is still largely unconfirmed in humans, diagnosis is difficult and targeted screening is advised to be started early upon suspicion of PPCM. Extensive cohort studies like the ESC EORP are crucial to gain a better understanding of the clinical presentation, risk factors, and prognosis, 4 but a registry on a global scale with consecutive patient selection will be highly incremental to our knowledge regarding geographical variation and incidence rates among countries. Moreover, recent insights into associated genetic factors and predisposition to PPCM indicated that these may predict a worse prognosis and have relevant clinical implications. Genetic evaluation may, therefore, be advisable for patients with a family history of cardiomyopathies. Moreover, little is known about the relationship between PPCM and pre-eclampsia. Disease definitions suggest that the diseases are fundamentally different, but the initial clinical symptoms may be indicative for both and have historically been a reason for a delayed diagnosis. However, while high blood pressure is required for pre-eclampsia, it is not for PPCM, which is characterized by a significant decline in LV function. Typically, PPCM occurs in the first months after delivery, whereas pre-eclampsia is seen in the second half of gestation and is effectively treated by removing the placenta. Pre-eclampsia may be a distinct entity, but it was repeatedly shown to be a risk factor to develop PPCM, which may suggest that pre-eclampsia may cause lasting damage to the maternal vasculature. The cardiovascular system undergoes fundamentally different changes in late pregnancy compared with the early postpartum period and the underlying cause for PPCM may be related to each period or the transition caused by delivery. From a mechanistic perspective, PPCM appears to result in heart failure secondary to vascular dysfunction. The two principal mouse models have demonstrated that regulation of angiogenesis during the peripartum period is tightly regulated and remarkably sensitive to detrimental stimuli 19,20. Currently, these animal models are the basis for several clinical studies and clinical recommendations, despite the uncertainty of how well findings in animals can be extrapolated to human patients. Hence, there is a great need for mechanistic studies in humans with PPCM in order to gain more insight into the pathophysiology of PPCM. In light of this, it was recently shown that hiPSC-derived cardiomyocytes from PPCM patients were

512

513

514

515

516

517

518

519

520

521

522

523

524

525

526

527

528

529

530

531

532

533

534

535

536

metabolically impaired *in vitro*²⁴. Pregnancy hormones extensively orchestrate maternal metabolism and angiogenesis during and after pregnancy. Alternatively, mutations in several genes have been associated with PPCM, some of which may also cause metabolic distress. However, it is unclear how these dysfunctional gene variants could interact with the mechanisms of pregnancy at specific times. It is known that placental hormones can cause pre-eclampsia; it remains unclear if a specific hormone profile can be distinctly linked to PPCM pathogenesis. Taken together, pregnancy hormones might link the delicately balanced angiogenic state to potentially unstable metabolic processes in the heart of PPCM patients.

References

- 1. Bauersachs, J. *et al.* Pathophysiology, diagnosis and management of peripartum

 cardiomyopathy: a position statement from the Heart Failure Association of the European

 Society of Cardiology Study Group on peripartum cardiomyopathy. *Eur. J. Heart Fail.* **21**, 827–843 (2019).
- Sliwa, K. *et al.* Current state of knowledge on aetiology, diagnosis, management, and therapy
 of peripartum cardiomyopathy: a position statement from the Heart Failure Association of
 the European Society of Cardiology Working Group on peripartum cardiomyopathy. *Eur. J.*Heart Fail. **12**, 767–78 (2010).
- 555 3. Sliwa, K. *et al.* Risk stratification and management of women with cardiomyopathy/heart
 556 failure planning pregnancy or presenting during/after pregnancy: a position statement from
 557 the Heart Failure Association of the European Society of Cardiology Study Group on
 558 Peripartum . *Eur. J. Heart Fail.* **23**, 527–540 (2021).
- 559 4. Sliwa, K. *et al.* Clinical presentation, management, and 6-month outcomes in women with peripartum cardiomyopathy: an ESC EORP registry. *Eur. Heart J.* **41**, 3787–3797 (2020).
- 5. Davis, M. B., Arany, Z., McNamara, D. M., Goland, S. & Elkayam, U. Peripartum

 Cardiomyopathy. *J. Am. Coll. Cardiol.* **75**, 207–221 (2020).
- 6. Honigberg, M. C. & Givertz, M. M. Peripartum cardiomyopathy. BMJ k5287 (2019).
- 7. Moulig, V. *et al.* Long-term follow-up in peripartum cardiomyopathy patients with

 contemporary treatment: low mortality, high cardiac recovery, but significant cardiovascular

 co-morbidities. *Eur. J. Heart Fail.* **21**, 1534–1542 (2019).
- 8. Sliwa, K. *et al.* Long-term prognosis, subsequent pregnancy, contraception and overall
 management of peripartum cardiomyopathy: practical guidance paper from the Heart Failure
 Association of the European Society of Cardiology Study Group on Peripartum
 Cardiomyopathy. *Eur. J. Heart Fail.* **20**, 951–962 (2018).
- 9. Kamiya, C. A. et al. Different characteristics of peripartum cardiomyopathy between patients

- complicated with and without hypertensive disorders. -Results from the Japanese Nationwide survey of peripartum cardiomyopathy-. *Circ. J.* **75**, 1975–81 (2011).
- 10. Isogai, T. & Kamiya, C. A. Worldwide Incidence of Peripartum Cardiomyopathy and Overall

 Maternal Mortality. *Int. Heart J.* **60**, 503–511 (2019).
- 576 11. Ersbøll, A. S. *et al.* Peripartum cardiomyopathy in Denmark: a retrospective, population-based 577 study of incidence, management and outcome. *Eur. J. Heart Fail.* **19**, 1712–1720 (2017).
- 12. Barasa, A., Rosengren, A., Sandström, T. Z., Ladfors, L. & Schaufelberger, M. Heart Failure in
 Late Pregnancy and Postpartum: Incidence and Long-Term Mortality in Sweden From 1997 to
 2010. *J. Card. Fail.* **23**, 370–378 (2017).
- 13. Isezuo, S. A. & Abubakar, S. A. Epidemiologic profile of peripartum cardiomyopathy in a tertiary care hospital. *Ethn. Dis.* **17**, 228–33 (2007).
- Fett, J. D., Christie, L. G., Carraway, R. D. & Murphy, J. G. Five-year prospective study of the incidence and prognosis of peripartum cardiomyopathy at a single institution. *Mayo Clin.*Proc. **80**, 1602–6 (2005).
- Hasan, J. A., Qureshi, A., Ramejo, B. B. & Kamran, A. Peripartum cardiomyopathy

 characteristics and outcome in a tertiary care hospital. *J. Pak. Med. Assoc.* **60**, 377–80 (2010).
- Desai, D., Moodley, J. & Naidoo, D. Peripartum Cardiomyopathy: Experiences at King Edward

 VIII Hospital, Durban, South Africa and a Review of the Literature. *Trop. Doct.* **25**, 118–123

 (1995).
- 591 17. Koenig, T., Hilfiker-Kleiner, D. & Bauersachs, J. Peripartum cardiomyopathy. *Herz* **43**, 431–437 592 (2018).
- 18. Kolte, D. *et al.* Temporal Trends in Incidence and Outcomes of Peripartum Cardiomyopathy in the United States: A Nationwide Population-Based Study. *J. Am. Heart Assoc.* **3**, (2014).
- Hilfiker-Kleiner, D. *et al.* A Cathepsin D-Cleaved 16 kDa Form of Prolactin Mediates
 Postpartum Cardiomyopathy. *Cell* 128, 589–600 (2007).
- 597 20. Patten, I. S. et al. Cardiac angiogenic imbalance leads to peripartum cardiomyopathy. Nature

- **485**, 333–338 (2012).
- Halkein, J. *et al.* MicroRNA-146a is a therapeutic target and biomarker for peripartum cardiomyopathy. *J. Clin. Invest.* **123**, 2143–2154 (2013).
- Stapel, B. *et al.* Low STAT3 expression sensitizes to toxic effects of β-adrenergic receptor stimulation in peripartum cardiomyopathy. *Eur. Heart J.* **38**, ehw086 (2016).
- Ricke-Hoch, M. *et al.* In peripartum cardiomyopathy plasminogen activator inhibitor-1 is a potential new biomarker with controversial roles. *Cardiovasc. Res.* **116**, 1875–1886 (2020).
- Hoes, M. F. *et al.* Human iPSC-Derived Cardiomyocytes of Peripartum Patients With
 Cardiomyopathy Reveal Aberrant Regulation of Lipid Metabolism. *Circulation* **142**, 2288–2291
 (2020).
- Liu, L. X. & Arany, Z. Maternal cardiac metabolism in pregnancy. *Cardiovasc. Res.* **101**, 545–553 (2014).
- Redondo-Angulo, I. *et al.* Fgf21 is required for cardiac remodeling in pregnancy. *Cardiovasc.*Res. **113**, 1574–1584 (2017).
- Kodogo, V., Azibani, F. & Sliwa, K. Role of pregnancy hormones and hormonal interaction on the maternal cardiovascular system: a literature review. *Clin. Res. Cardiol.* **108**, 831–846 (2019).
- Longo, L. D. Maternal blood volume and cardiac output during pregnancy: a hypothesis of endocrinologic control. *Am. J. Physiol. Integr. Comp. Physiol.* **245**, R720–R729 (1983).
- Clapp, J. F. & Capeless, E. Cardiovascular Function Before, During, and After the First and
 Subsequent Pregnancies. *Am. J. Cardiol.* 80, 1469–1473 (1997).
- Melchiorre, K., Sharma, R., Khalil, A. & Thilaganathan, B. Maternal Cardiovascular Function in
 Normal Pregnancy. *Hypertension* **67**, 754–762 (2016).
- Loerup, L. *et al.* Trends of blood pressure and heart rate in normal pregnancies: a systematic review and meta-analysis. *BMC Med.* **17**, 167 (2019).
- 623 32. Green, L. J. et al. Gestation-Specific Vital Sign Reference Ranges in Pregnancy. Obstet.

624 *Gynecol.* **135**, 653–664 (2020).

Reson. 16, 1 (2014).

- Ducas, R. A. *et al.* Cardiovascular magnetic resonance in pregnancy: Insights from the cardiac hemodynamic imaging and remodeling in pregnancy (CHIRP) study. *J. Cardiovasc. Magn.*
- De Haas, S. *et al.* Cardiac remodeling in normotensive pregnancy and in pregnancy complicated by hypertension: systematic review and meta-analysis. *Ultrasound Obstet. Gynecol.* **50**, 683–696 (2017).
- Umar, S. *et al.* Cardiac structural and hemodynamic changes associated with physiological heart hypertrophy of pregnancy are reversed postpartum. *J. Appl. Physiol.* **113**, 1253–9 (2012).
- 634 36. Chung, E., Yeung, F. & Leinwand, L. A. Akt and MAPK signaling mediate pregnancy-induced 635 cardiac adaptation. *J. Appl. Physiol.* **112**, 1564–1575 (2012).
- Aljabri, M. B. *et al.* Pregnancy protects against antiangiogenic and fibrogenic effects of angiotensin II in rat hearts. *Acta Physiol.* **201**, 445–456 (2011).
- 38. Peters, F. *et al.* Peripartum Cardiomyopathy Associated With Left Ventricular Noncompaction

 Phenotype and Reversible Rigid Body Rotation. *Circ. Hear. Fail.* **6**, (2013).
- Lea, B., Bailey, A. L., Wiisanen, M. E., Attili, A. & Rajagopalan, N. Left ventricular noncompaction presenting as peripartum cardiomyopathy. *Int. J. Cardiol.* **154**, e65–e66 (2012).
- 40. Haghikia, A. *et al.* Prognostic implication of right ventricular involvement in peripartum
 cardiomyopathy: a cardiovascular magnetic resonance study. *ESC Hear. Fail.* 2, 139–149
 (2015).
- Yang, W.-I. *et al.* Clinical features differentiating Takotsubo cardiomyopathy in the peripartum period from peripartum cardiomyopathy. *Heart Vessels* **35**, 665–671 (2020).
- Eghbali, M., Wang, Y., Toro, L. & Stefani, E. Heart Hypertrophy During Pregnancy: A Better
 Functioning Heart? *Trends Cardiovasc. Med.* 16, 285–291 (2006).

- 650 43. Chung, E., Yeung, F. & Leinwand, L. A. Calcineurin activity is required for cardiac remodelling 651 in pregnancy. *Cardiovasc. Res.* **100**, 402–410 (2013).
- 652 44. Saffer, C. *et al.* Determination of placental growth factor (PIGF) levels in healthy pregnant
 653 women without signs or symptoms of preeclampsia. *Pregnancy Hypertens. An Int. J. Women's*654 *Cardiovasc. Heal.* **3**, 124–132 (2013).
- 45. Hunter, A. *et al.* Serum Levels of Vascular Endothelial Growth Factor in Preeclamptic and
 Normotensive Pregnancy. *Hypertension* 36, 965–969 (2000).
- Maynard, S. E. *et al.* Excess placental soluble fms-like tyrosine kinase 1 (sFlt1) may contribute to endothelial dysfunction hypertension, and proteinuria in preeclampsia. *J. Clin. Invest.* **111**, 649–658 (2003).
- Zeisler, H. *et al.* Predictive Value of the sFlt-1:PIGF Ratio in Women with Suspected
 Preeclampsia. *N. Engl. J. Med.* 374, 13–22 (2016).
- 662 48. Mebazaa, A. *et al.* Imbalanced Angiogenesis in Peripartum Cardiomyopathy Diagnostic
 663 Value of Placenta Growth Factor . *Circ. J.* **81**, 1654–1661 (2017).
- 664 49. Mizuno, Y. *et al.* The diabetic heart utilizes ketone bodies as an energy source. *Metabolism*665 **77**, 65–72 (2017).
- 666 50. Murashige, D. *et al.* Comprehensive quantification of fuel use by the failing and nonfailing 667 human heart. *Science (80-.).* **370**, 364–368 (2020).
- Lain, K. Y. & Catalano, P. M. Metabolic Changes in Pregnancy. *Clin. Obstet. Gynecol.* **50**, 938–948 (2007).
- Lof, M. *et al.* Changes in basal metabolic rate during pregnancy in relation to changes in body weight and composition, cardiac output, insulin-like growth factor I, and thyroid hormones and in relation to fetal growth. *Am. J. Clin. Nutr.* **81**, 678–685 (2005).
- Buchanan, T. A., Metzger, B. E., Freinkel, N. & Bergman, R. N. Insulin sensitivity and B-cell responsiveness to glucose during late pregnancy in lean and moderately obese women with normal glucose tolerance or mild gestational diabetes. *Am. J. Obstet. Gynecol.* **162**, 1008–

- 676 **1014 (1990)**.
- 677 54. Catalano, P. M., Tyzbir, E. D., Roman, N. M., Amini, S. B. & Sims, E. A. H. Longitudinal changes 678 in insulin release and insulin resistance in nonobese pregnant women. *Am. J. Obstet. Gynecol.* 679 **165**, 1667–1672 (1991).
- Sugden, M. C., Changani, K. K., Bentley, J. & Holness, M. J. Cardiac glucose metabolism during pregnancy. *Biochem. Soc. Trans.* **20**, 195S (1992).
- 682 56. Williams, J. G. *et al.* Coronary nitric oxide production controls cardiac substrate metabolism during pregnancy in the dog. *Am. J. Physiol. Circ. Physiol.* **294**, H2516–H2523 (2008).
- 57. Liu, L. X. *et al.* PDK4 Inhibits Cardiac Pyruvate Oxidation in Late Pregnancy. *Circ. Res.* **121**, 1370–1378 (2017).
- Randle, P. J., Garland, P. B., Hales, C. N. & Newsholme, E. A. the Glucose Fatty-Acid Cycle Its

 Role in Insulin Sensitivity and the Metabolic Disturbances of Diabetes Mellitus. *Lancet*(London, England) 1, 785–9 (1963).
- 689 59. Whittaker, P. G., Macphail, S. & Lind, T. Serial hematologic changes and pregnancy outcome.
 690 *Obstet. Gynecol.* **88**, 33–9 (1996).
- 691 60. Abbassi-Ghanavati, M., Greer, L. G. & Cunningham, F. G. Pregnancy and Laboratory Studies.
 692 *Obstet. Gynecol.* **114**, 1326–1331 (2009).
- 693 61. Zafirovic, S. *et al.* 17β-Estradiol protects against the effects of a high fat diet on cardiac glucose, lipid and nitric oxide metabolism in rats. *Mol. Cell. Endocrinol.* **446**, 12–20 (2017).
- 62. Caulin-Glaser, T., García-Cardeña, G., Sarrel, P., Sessa, W. C. & Bender, J. R. 17 beta-estradiol 696 regulation of human endothelial cell basal nitric oxide release, independent of cytosolic Ca2+ 697 mobilization. *Circ. Res.* **81**, 885–92 (1997).
- 63. Simoncini, T., Genazzani, A. R. & Liao, J. K. Nongenomic mechanisms of endothelial nitric

 oxide synthase activation by the selective estrogen receptor modulator raloxifene. *Circulation*105, 1368–73 (2002).
- 64. Johnson, M. L., Grazul-Bilska, A. T., Redmer, D. A. & Reynolds, L. P. Effects of estradiol-17beta

- on expression of mRNA for seven angiogenic factors and their receptors in the endometrium of ovariectomized (OVX) ewes. *Endocrine* **30**, 333–42 (2006).
- 65. Hervé, M. A. J. *et al.* Regulation of the vascular endothelial growth factor (VEGF) receptor Flk
 1/KDR by estradiol through VEGF in uterus. *J. Endocrinol.* **188**, 91–9 (2006).
- 706 66. Straub, R. H. The Complex Role of Estrogens in Inflammation. *Endocr. Rev.* **28**, 521–574 (2007).
- 708 67. van Eickels, M. *et al.* 17beta-estradiol attenuates the development of pressure-overload 709 hypertrophy. *Circulation* **104**, 1419–23 (2001).
- Satoh, M. *et al.* Inhibition of Apoptosis-Regulated Signaling Kinase-1 and Prevention of
 Congestive Heart Failure by Estrogen. *Circulation* 115, 3197–3204 (2007).
- Fortini, F. *et al.* Estrogen receptor β –dependent Notch1 activation protects vascular endothelium against tumor necrosis factor α (TNF α)-induced apoptosis. *J. Biol. Chem.* **292**, 18178–18191 (2017).
- 70. Patten, R. D. *et al.* 17β-Estradiol Reduces Cardiomyocyte Apoptosis In Vivo and In Vitro via
 Activation of Phospho-Inositide-3 Kinase/Akt Signaling. *Circ. Res.* 95, 692–699 (2004).
- 71. Wang, T. *et al.* Estrogen-Related Receptor α (ERRα) and ERRγ Are Essential Coordinators of
 718 Cardiac Metabolism and Function. *Mol. Cell. Biol.* 35, 1281–1298 (2015).
- 719 72. Morrissy, S., Xu, B., Aguilar, D., Zhang, J. & Chen, Q. M. Inhibition of apoptosis by progesterone in cardiomyocytes. *Aging Cell* **9**, 799–809 (2010).
- 73. Ramírez-Rosas, M. B., Cobos-Puc, L. E., Sánchez-López, A., Gutiérrez-Lara, E. J. & Centurión, D.
 Pharmacological characterization of the mechanisms involved in the vasorelaxation induced
 by progesterone and 17β-estradiol on isolated canine basilar and internal carotid arteries.
 Steroids 89, 33–40 (2014).
- 74. Amaral, L. M. *et al.* Progesterone supplementation attenuates hypertension and the

 autoantibody to the angiotensin II type I receptor in response to elevated interleukin-6 during

 pregnancy. *Am. J. Obstet. Gynecol.* **211**, 158.e1-158.e6 (2014).

- 75. Nelson, S. H. *et al.* Increased Nitric Oxide Synthase Activity and Expression in the Human

 Uterine Artery During Pregnancy. *Circ. Res.* **87**, 406–411 (2000).
- 730 76. Atif, F., Yousuf, S., Espinosa-Garcia, C., Sergeeva, E. & Stein, D. G. Progesterone Treatment

 731 Attenuates Glycolytic Metabolism and Induces Senescence in Glioblastoma. *Sci. Reports 2019*732 *91* **9**, 1–12 (2019).
- 733 77. Kyo, S. *et al.* Forkhead Transcription Factor FOXO1 is a Direct Target of Progestin to Inhibit

 734 Endometrial Epithelial Cell Growth. *Clin. Cancer Res.* **17**, 525–537 (2011).
- 735 78. Freeman, M. E., Kanyicska, B., Lerant, A. & Nagy, G. Prolactin: Structure, Function, and Regulation of Secretion. *Physiol. Rev.* **80**, 1523–1631 (2000).
- 737 79. Mills, D. E. & Ward, R. P. Effect of Prolactin on Blood Pressure and Cardiovascular
 738 Responsiveness in the Rat. *Exp. Biol. Med.* **181**, 3–8 (1986).
- Hsieh, D. J.-Y. *et al.* Prolactin protects cardiomyocytes against intermittent hypoxia-induced cell damage by the modulation of signaling pathways related to cardiac hypertrophy and proliferation. *Int. J. Cardiol.* **181**, 255–266 (2015).
- 742 81. Gonzalez, C. *et al.* The prolactin family hormones regulate vascular tone through NO and
 743 prostacyclin production in isolated rat aortic rings. *Acta Pharmacol. Sin. 2015 365* **36**, 572–
 744 586 (2015).
- 745 82. Cui, Y. *et al.* Hepatic FGF21 production is increased in late pregnancy in the mouse. *Am. J. Physiol. Integr. Comp. Physiol.* **307**, R290–R298 (2014).
- 83. Badman, M. K. *et al.* Hepatic Fibroblast Growth Factor 21 Is Regulated by PPARα and Is a Key
 Mediator of Hepatic Lipid Metabolism in Ketotic States. *Cell Metab.* 5, 426–437 (2007).
- Response by PPARα-Mediated Induction
 of Fibroblast Growth Factor 21. *Cell Metab.* 5, 415–425 (2007).
- Planavila, A. *et al.* Fibroblast growth factor 21 protects against cardiac hypertrophy in mice.
 Nat. Commun. 4, 2019 (2013).
- 753 86. Planavila, A. et al. Fibroblast growth factor 21 protects the heart from oxidative stress.

- 754 *Cardiovasc. Res.* **106**, 19–31 (2015).
- Sutton, E. F., Morrison, C. D., Stephens, J. M. & Redman, L. M. Fibroblast growth factor 21, adiposity, and macronutrient balance in a healthy, pregnant population with overweight and obesity. *Endocr. Res.* **43**, 275 (2018).
- 758 88. Yuan, D., Wu, B. J., Henry, A., Rye, K.-A. & Ong, K. L. Role of fibroblast growth factor 21 in 759 gestational diabetes mellitus: A mini-review. *Clin. Endocrinol. (Oxf).* **90**, 47–55 (2019).
- 760 89. Goli, R. *et al.* Genetic and Phenotypic Landscape of Peripartum Cardiomyopathy. *Circulation* 761 **143**, 1852–1862 (2021).
- McNamara, D. M. *et al.* Clinical Outcomes for Peripartum Cardiomyopathy in North America:
 Results of the IPAC Study (Investigations of Pregnancy-Associated Cardiomyopathy). *J. Am. Coll. Cardiol.* 66, 905–14 (2015).
- Jackson, A. M. *et al.* Hypertensive disorders in women with peripartum cardiomyopathy:
 insights from the ESC Peripartum Cardiomyopathy Registry. *Eur. J. Heart Fail.* ejhf.2264
 (2021).
- 768 92. Cénac, A., Gaultier, Y., Devillechabrolle, A. & Moulias, R. Enterovirus infection in peripartum
 769 cardiomyopathy. *Lancet (London, England)* **2**, 968–9 (1988).
- 93. Elkayam, U. et al. Pregnancy-Associated Cardiomyopathy. Circulation 111, 2050–2055 (2005).
- 94. Pfeffer, T. J. *et al.* Increased Cancer Prevalence in Peripartum Cardiomyopathy. *JACC* 772 CardioOncology 1, 196–205 (2019).
- 95. Cherubin, S. *et al.* Systematic review and meta-analysis of prolactin and iron deficiency in peripartum cardiomyopathy. *Open Hear.* **7**, e001430 (2020).
- 96. Forster, O. *et al.* Reversal of IFN-γ, oxLDL and prolactin serum levels correlate with clinical
 improvement in patients with peripartum cardiomyopathy. *Eur. J. Heart Fail.* 10, 861–868
 (2008).
- Haghikia, A. *et al.* Phenotyping and outcome on contemporary management in a German cohort of patients with peripartum cardiomyopathy. *Basic Res. Cardiol.* **108**, (2013).

- 780 98. Liu, J. *et al.* The correlation between peripartum cardiomyopathy and autoantibodies against cardiovascular receptors. *PLoS One* **9**, e86770 (2014).
- Huang, G.-Y., Zhang, L.-Y., Bai, T.-F., Wang, R.-K. & Zhang, X.-S. Effect of inflammation and autoimmunity in peripartum cardiomyopathy. *Clin. Res. Geriatr Cardiol* **7**, 106–109 (2010).
- Huang, G. Y., Zhang, L. Y., Long-Le, M. A. & Wang, L.-X. Clinical characteristics and risk factors for peripartum cardiomyopathy. *Afr. Health Sci.* **12**, 26–31 (2012).
- 101. Sagy, I. *et al.* Peripartum cardiomyopathy is associated with increased uric acid concentrations: A population based study. *Heart Lung* **46**, 369–374 (2017).
- Yaqoob, I. *et al.* Insertion/deletion polymorphism of ACE gene in females with peripartum cardiomyopathy: A case-control study. *Indian Heart J.* **70**, 66–70 (2018).
- Tamrat, R. *et al.* Women with peripartum cardiomyopathy have normal ejection fraction, but abnormal systolic strain, during pregnancy. *ESC Hear. Fail.* **8**, 3382–3386 (2021).
- Morales, A. *et al.* Rare Variant Mutations in Pregnancy-Associated or Peripartum
 Cardiomyopathy. *Circulation* 121, 2176–2182 (2010).
- van Spaendonck-Zwarts, K. Y. *et al.* Titin gene mutations are common in families with both peripartum cardiomyopathy and dilated cardiomyopathy. *Eur. Heart J.* **35**, 2165–2173 (2014).
- Ware, J. S. *et al.* Shared Genetic Predisposition in Peripartum and Dilated Cardiomyopathies.
 N. Engl. J. Med. **374**, 233–41 (2016).
- 798 107. Mazzarotto, F. *et al.* Reevaluating the Genetic Contribution of Monogenic Dilated
 799 Cardiomyopathy. *Circulation* **141**, 387–398 (2020).
- van Spaendonck-Zwarts, K. Y. *et al.* Peripartum Cardiomyopathy as a Part of Familial Dilated
 Cardiomyopathy. *Circulation* 121, 2169–2175 (2010).
- Spracklen, T. F. *et al.* Genetics of Peripartum Cardiomyopathy: Current Knowledge, Future

 Directions and Clinical Implications. *Genes (Basel).* **12**, 103 (2021).
- Horne, B. D. *et al.* Genome-wide significance and replication of the chromosome 12p11.22 locus near the PTHLH gene for peripartum cardiomyopathy. *Circ. Cardiovasc. Genet.* **4**, 359–

- 806 66 (2011).
- 111. Karaye, K. M. *et al.* Incidence, clinical characteristics, and risk factors of peripartum cardiomyopathy in Nigeria: results from the PEACE Registry. *ESC Hear. Fail.* **7**, (2020).
- 112. Karaye, K., Yahaya, I., Lindmark, K. & Henein, M. Serum Selenium and Ceruloplasmin in

 Nigerians with Peripartum Cardiomyopathy. *Int. J. Mol. Sci.* **16**, 7644–7654 (2015).
- 113. Karaye, K. M. *et al.* Selenium supplementation in patients with peripartum cardiomyopathy: a proof-of-concept trial. *BMC Cardiovasc. Disord.* **20**, 457 (2020).
- 813 114. Bomer, N. et al. Selenium and outcome in heart failure. Eur. J. Heart Fail. (2019).
- Mielniczuk, L. M. *et al.* Frequency of Peripartum Cardiomyopathy. *Am. J. Cardiol.* **97**, 1765–
 1768 (2006).
- 116. Krishnamoorthy, P. *et al.* Epidemiology and outcomes of peripartum cardiomyopathy in the
 United States. *J. Cardiovasc. Med.* **17**, 756–761 (2016).
- 117. US Census Bureau July 1 2019 Estimates. *United States Census Bureau*. Retrieved: Aug 17,

 2021 https://www.census.gov/quickfacts/fact/table/US/PST045219 (2019).
- 118. Irizarry, O. C. *et al.* Comparison of Clinical Characteristics and Outcomes of Peripartum

 Cardiomyopathy Between African American and Non–African American Women. *JAMA*Cardiol. **2**, 1256 (2017).
- 119. Goland, S., Modi, K., Hatamizadeh, P. & Elkayam, U. Differences in clinical profile of AfricanAmerican women with peripartum cardiomyopathy in the United States. *J. Card. Fail.* **19**,

 214–8 (2013).
- 120. Sliwa, K. *et al.* Clinical characteristics of patients from the worldwide registry on peripartum cardiomyopathy (PPCM). *Eur. J. Heart Fail.* **19**, 1131–1141 (2017).
- 121. Getz, K. D. *et al.* Neighborhood education status drives racial disparities in clinical outcomes in PPCM. *Am. Heart J.* **238**, 27–32 (2021).
- Azibani, F. *et al.* Outcome in German and South African peripartum cardiomyopathy cohorts associates with medical therapy and fibrosis markers. *ESC Hear. Fail.* **7**, 512–522 (2020).

- 123. Karaye, K. M. *et al.* Clinical Features and Outcomes of Peripartum Cardiomyopathy in Nigeria.
- 33 J. Am. Coll. Cardiol. **76**, 2352–2364 (2020).
- 124. Amos, A. M., Jaber, W. A. & Russell, S. D. Improved outcomes in peripartum cardiomyopathy with contemporary. *Am. Heart J.* **152**, 509–513 (2006).
- 336 125. Gunderson, E. P. *et al.* Epidemiology of peripartum cardiomyopathy: Incidence, predictors, and outcomes. *Obstet. Gynecol.* **118**, 583–591 (2011).
- Bello, N., Rendon, I. S. H. & Arany, Z. The relationship between pre-eclampsia and peripartum cardiomyopathy: a systematic review and meta-analysis. *J. Am. Coll. Cardiol.* **62**, 1715–1723 (2013).
- Young, B. C., Levine, R. J. & Karumanchi, S. A. Pathogenesis of preeclampsia. *Annual Review of Pathology: Mechanisms of Disease* vol. 5 173–192 (2010).
- Lewey, J., Levine, L. D., Elovitz, M. A., Irizarry, O. C. & Arany, Z. Importance of Early Diagnosis in Peripartum Cardiomyopathy. *Hypertens. (Dallas, Tex. 1979)* **75**, 91–97 (2020).
- Steegers, E. A. P., Von Dadelszen, P., Duvekot, J. J. & Pijnenborg, R. Pre-eclampsia. in *The Lancet* vol. 376 631–644 (Elsevier B.V., 2010).
- 130. Levine, R. J. *et al.* Circulating Angiogenic Factors and the Risk of Preeclampsia. *N. Engl. J. Med.*848 **350**, 672–683 (2004).
- Damp, J. *et al.* Relaxin-2 and Soluble Flt1 Levels in Peripartum Cardiomyopathy. Results of the
 Multicenter IPAC Study. *JACC Hear. Fail.* **4**, 380–388 (2016).
- Bültmann, B. D., Klingel, K., Näbauer, M., Wallwiener, D. & Kandolf, R. High prevalence of viral genomes and inflammation in peripartum cardiomyopathy. *Am. J. Obstet. Gynecol.* **193**, 363–5 (2005).
- 133. Sarojini, A., Sai Ravi Shanker, A. & Anitha, M. Inflammatory Markers-Serum Level of C
 Reactive Protein, Tumor Necrotic Factor-α, and Interleukin-6 as Predictors of Outcome for

 Peripartum Cardiomyopathy. *J. Obstet. Gynecol. India* **63**, 234–239 (2013).
- 857 134. Koczo, A. et al. Proinflammatory TH17 cytokine activation, disease severity and outcomes in

- 858 peripartum cardiomyopathy. *Int. J. Cardiol.* (2021).
- 135. McTiernan, C. F. *et al.* Circulating T-Cell Subsets, Monocytes, and Natural Killer Cells in

 Peripartum Cardiomyopathy: Results From the Multicenter IPAC Study. *J. Card. Fail.* **24**, 33–

 42 (2018).
- Haghikia, A. *et al.* Evidence of autoantibodies against cardiac troponin I and sarcomeric myosin in peripartum cardiomyopathy. *Basic Res. Cardiol.* **110**, 60 (2015).
- 137. Elkayam, U. *et al.* Maternal and Fetal Outcomes of Subsequent Pregnancies in Women with

 Peripartum Cardiomyopathy. *N. Engl. J. Med.* **344**, 1567–1571 (2001).
- Hilfiker-Kleiner, D. *et al.* Outcome of subsequent pregnancies in patients with a history of peripartum cardiomyopathy. *Eur. J. Heart Fail.* **19**, 1723–1728 (2017).
- Elkayam, U. *et al.* Maternal and fetal outcomes of subsequent pregnancies in women with peripartum cardiomyopathy. *N. Engl. J. Med.* **344**, 1567–71 (2001).
- 140. Lampert, M. B. *et al.* Contractile reserve in patients with peripartum cardiomyopathy and recovered left ventricular function. *Am. J. Obstet. Gynecol.* **176**, 189–95 (1997).
- 141. Goland, S. *et al.* Angiogenic Imbalance and Residual Myocardial Injury in Recovered

 Peripartum Cardiomyopathy Patients. *Circ. Heart Fail.* **9**, (2016).
- Toescu, V., Nuttall, S. L., Martin, U., Kendall, M. J. & Dunne, F. Oxidative stress and normal pregnancy. *Clin. Endocrinol. (Oxf).* **57**, 609–613 (2002).
- Ricke-Hoch, M. *et al.* Opposing roles of Akt and STAT3 in the protection of the maternal heart from peripartum stress. *Cardiovasc. Res.* **101**, 587–596 (2014).
- Hoes, M. F. *et al.* The role of cathepsin D in the pathophysiology of heart failure and its potentially beneficial properties: a translational approach. *Eur. J. Heart Fail.* ejhf.1674 (2019).
- 145. Yadati, T., Houben, T., Bitorina, A. & Shiri-Sverdlov, R. The Ins and Outs of Cathepsins:

 Physiological Function and Role in Disease Management. *Cells* **9**, 1679 (2020).
- 146. Cruz-Soto, M. E. *et al.* Cathepsin D Is the Primary Protease for the Generation of

 Adenohypophyseal Vasoinhibins: Cleavage Occurs within the Prolactin Secretory Granules.

- 884 Endocrinology **150**, 5446–5454 (2009).
- Piwnica, D. *et al.* Cathepsin D processes human prolactin into multiple 16K-like N-terminal fragments: study of their antiangiogenic properties and physiological relevance. *Mol. Endocrinol.* **18**, 2522–42 (2004).
- Triebel, J., Bertsch, T., MartÃ-nez de la Escalera, G. & Clapp, C. On the Path toward Classifying

 Hormones of the Vasoinhibin-Family. *Front. Endocrinol. (Lausanne).* **6**, (2015).
- Bajou, K. *et al.* PAI-1 mediates the antiangiogenic and profibrinolytic effects of 16K prolactin.
 Nat. Med. 20, 741–747 (2014).
- 150. Tabruyn, S. P., Nguyen, N.-Q.-N., Cornet, A. M., Martial, J. A. & Struman, I. The Antiangiogenic Factor, 16-kDa Human Prolactin, Induces Endothelial Cell Cycle Arrest by Acting at Both the GO–G1 and the G2–M Phases. *Mol. Endocrinol.* **19**, 1932–1942 (2005).
- Tabruyn, S. P. S. P. *et al.* The antiangiogenic factor 16K human prolactin induces caspasedependent apoptosis by a mechanism that requires activation of nuclear factor-kappaB. *Mol. Endocrinol.* **17**, 1815–23 (2003).
- Lee, S.-H., Kunz, J., Lin, S.-H. & Yu-Lee, L. 16-kDa Prolactin Inhibits Endothelial Cell Migration
 by Down-Regulating the Ras-Tiam1-Rac1-Pak1 Signaling Pathway. *Cancer Res.* **67**, 11045–
 11053 (2007).
- 901 153. Feyen, E. *et al.* ERBB4 and Multiple MicroRNAs That Target ERBB4 Participate in Pregnancy-902 Related Cardiomyopathy. *Circ. Hear. Fail.* (2021).
- 903 154. Odiete, O., Hill, M. F. & Sawyer, D. B. Neuregulin in Cardiovascular Development and Disease.
 904 *Circ. Res.* **111**, 1376–1385 (2012).
- 905 155. Gassmann, M. *et al.* Aberrant neural and cardiac development in mice lacking the ErbB4
 906 neuregulin receptor. *Nature* **378**, 390–394 (1995).
- 156. Koga, K. *et al.* Elevated Serum Soluble Vascular Endothelial Growth Factor Receptor 1

 (sVEGFR-1) Levels in Women with Preeclampsia. *J. Clin. Endocrinol. Metab.* **88**, 2348–2351

 (2003).

- 910 157. Sliwa, K. *et al.* Evaluation of Bromocriptine in the Treatment of Acute Severe Peripartum
 911 Cardiomyopathy. *Circulation* **121**, 1465–1473 (2010).
- 912 158. Hilfiker-Kleiner, D. *et al.* Bromocriptine for the treatment of peripartum cardiomyopathy: a
 913 multicentre randomized study. *Eur. Heart J.* **38**, 2671–2679 (2017).
- 914 159. Podewski, E. K. *et al.* Alterations in Janus kinase (JAK)-signal transducers and activators of 915 transcription (STAT) signaling in patients with end-stage dilated cardiomyopathy. *Circulation* 916 **107**, 798–802 (2003).
- 917 160. Triebel, J., Clapp, C., Martínez de la Escalera, G. & Bertsch, T. Remarks on the Prolactin
 918 Hypothesis of Peripartum Cardiomyopathy. *Front. Endocrinol. (Lausanne).* **8**, (2017).
- 161. Khurana, S., Liby, K., Buckley, A. R. & Ben-Jonathan, N. Proteolysis of Human Prolactin:

 Resistance to Cathepsin D and Formation of a Nonangiostatic, C-Terminal 16K Fragment by

 Thrombin1. *Endocrinology* **140**, 4127–4132 (1999).
- 922 162. Triebel, J. *et al.* Matrix Metalloproteases and Cathepsin D in Human Serum do not Cleave 923 Prolactin to Generate Vasoinhibin. *Clin. Lab.* **66**, (2020).
- 924 163. Bello, N. A. & Arany, Z. Molecular mechanisms of peripartum cardiomyopathy: A
 925 vascular/hormonal hypothesis. *Trends Cardiovasc. Med.* **25**, 499–504 (2015).
- 164. Li, Z. *et al.* Recombinant Vascular Endothelial Growth Factor 121 Attenuates Hypertension and Improves Kidney Damage in a Rat Model of Preeclampsia. *Hypertension* **50**, 686–692 (2007).
- 929 165. van der Pol, A. *et al.* Accumulation of 5-oxoproline in myocardial dysfunction and the 930 protective effects of OPLAH. *Sci. Transl. Med.* **9**, (2017).
- 931 166. van Spaendonck-Zwarts, K. Y. *et al.* Titin gene mutations are common in families with both 932 peripartum cardiomyopathy and dilated cardiomyopathy. *Eur. Heart J.* **35**, 2165–73 (2014).
- 933 167. Schafer, S. *et al.* Titin-truncating variants affect heart function in disease cohorts and the 934 general population. *Nat. Genet.* **49**, 46–53 (2017).
- 168. Verdonschot, J. A. J. et al. Titin cardiomyopathy leads to altered mitochondrial energetics,

936	increased fibrosis and long-term life-threatening arrhythmias. Eur. Heart J. 39, 864–873
937	(2018).

- 938 169. Stevens, C. M. *et al.* Changes in the dynamics of the cardiac troponin C molecule explain the effects of Ca2+-sensitizing mutations. *J. Biol. Chem.* **292**, 11915–11926 (2017).
- He, H., Javadpour, M. M., Latif, F., Tardiff, J. C. & Ingwall, J. S. R-92L and R-92W Mutations in

 Cardiac Troponin T Lead to Distinct Energetic Phenotypes in Intact Mouse Hearts. *Biophys. J.*942

 93, 1834–1844 (2007).
- Toepfer, C. N. *et al.* Myosin Sequestration Regulates Sarcomere Function, Cardiomyocyte
 Energetics, and Metabolism, Informing the Pathogenesis of Hypertrophic Cardiomyopathy.
 Circulation 141, 828–842 (2020).
- 946 172. Gamperl, A. K. & Driedzic, W. R. Chapter 7 Cardiovascular Function and Cardiac Metabolism.

 947 in 301–360 (2009).

Figure legends

Figure 1 – Pregnancy hormone levels correlated to the cardiac bioenergetic profile. Fluctuating hormone levels correlate with changes in cardiac metabolic substrate during the progression of pregnancy. Late gestation is associated with increased utilization of free fatty acids and ketones, while pyruvate and lactate usage is reduced. The yellow line denotes childbirth and red gradients highlight general time of PPCM onset.

Figure 2 – Pregnancy hormones elicit tissue-specific effects. The pregnancy hormones prolactin (and its cleavage product 16 kDa prolactin), oestrogens, FGF21, and progesterone have pleiotropic effects in the vasculature and in cardiomyocytes related to apoptosis, angiogenesis, metabolism, vascular function, cardiac hypertrophy, and oxidative stress.

Figure 3 – hypothesized pathogenic pathways of PPCM. During early gestation, the placenta secretes high levels of vascular endothelial growth factor (VEGF) and placental growth factor (PIGF) into the maternal circulation, which stimulate vascularization and organ perfusion. As pregnancy progresses, circulating levels of VEGF and PIGF rise in concert with the increased secretion of the angiostatic soluble VEGF receptor-1 (sFIt-1). Alternatively, cardiomyocyte stress induces the exocytosis of the proteolytic enzyme cathepsin D (CTSD), which cleaves prolactin into 16 kDa fragment that is cytotoxic to endothelial cells. Consequently, endothelial cells secrete exosomes loaded with microRNA-146a (miR-146a) and inhibits various cardiomyocyte processes, including ERBB4-mediated metabolism. Both mechanisms have a central role for the vasculature in the heart and may lead to the development of PPCM.

Key points

- Physiological cardiovascular changes during pregnancy appear to uniquely boost PPCM development in predisposed women.
- 2. PPCM onset typically overlaps with the most profound changes in hormone levels.
- Models for PPCM indicate disruption of metabolic flexibility and angiogenic balance, possibly due to aberrant hormonal signaling.
 - 4. Most mechanisms derived from animal models remain to be confirmed in humans but form the basis for current clinical guidelines and future experiments.
 - 5. Registries should be based on consecutive screening and help to determine actual geographic variation of incidence rates.

Competing interests

J.B. received honoraria for lectures/consulting from Novartis, Vifor, Bayer, Servier, Abiomed, Pfizer, Boehringer Ingelheim, AstraZeneca, Cardior, Daichii Sankyo, CVRx, BMS, MSD, Amgen, Corvia, not

related to this article; and research support for the department from Zoll, CVRx, Vifor, Abiomed, not related to this article. P.v.d.M. received consultancy fees and/or grants from Novartis, Corvidia, Singulex, Servier, Vifor Pharma, Astra Zeneca, Pfizer, Pharmacosmos, PharmaNord and Ionis.

989

990

Author contributions

- 991 M.F.H. wrote the manuscript. All authors contributed substantially to the discussion of content,
- researching data for the article, and editing before submission.





