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# Native bighead carp *Hypophthalmichthys nobilis* and silver carp *Hypophthalmichthys molitrix* populations in the Pearl River are threatened by Yangtze River introductions as revealed by mitochondrial DNA

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6 Chao Li<sup>1,2</sup>† Junjie Wang<sup>1</sup>† Jiaqi Chen<sup>1</sup> Kevin Schneider<sup>2</sup> Radhakrishnan K.
7 Veettil<sup>3</sup> Kathryn R. Elmer<sup>2</sup> Jun Zhao<sup>1</sup>

8

<sup>1</sup>Guangzhou Key Laboratory of Subtropical Biodiversity and Biomonitoring,
Guangdong Provincial Key Laboratory for Healthy and Safe Aquaculture, Guangdong
Provincial Engineering Technology Research Center for Environmentally Friendly
Aquaculture, School of Life Science, South China Normal University, Guangzhou
510631, China

<sup>2</sup>Institute of Biodiversity, Animal Health and Comparative Medicine, College of
Medical, Veterinary and Life Sciences, University of Glasgow, Glasgow G12 8QQ,
Scotland, UK

17 <sup>3</sup>College of Fisheries, Central Agricultural University, Agartala, India

18

19 Correspondence

Jun Zhao, Guangzhou Key Laboratory of Subtropical Biodiversity and
Biomonitoring, Guangdong Provincial Key Laboratory for Healthy and Safe
Aquaculture, Guangdong Provincial Engineering Technology Research Center for
Environmentally Friendly Aquaculture, School of Life Science, South China Normal

<sup>†</sup> Chao Li and Junjie Wang made an equal contribution to this work.

- 24 University. Guangzhou 510631, China
- 25 Email: zhaojun@scnu.edu.cn
- 26

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#### 35 Abstract

36 Bighead carp Hypophthalmichthys nobilis and silver carp Hypophthalmichthys *molitrix* have been two economically important aquaculture species in China for 37 centuries. In the past decades, bighead and silver carp have been introduced from 38 39 Yangtze River to many river systems in China, including Pearl River, in annual 40 large-scale stocking activities to enhance wild fisheries. However, few studies have assessed the ecological or genetic impacts of such introductions on native conspecific 41 42 fish populations. We obtained a mtDNA D-loop segment of 978 bp from 213 bighead carp samples from nine populations and a 975 bp segment from 204 silver carp 43 samples from ten populations to evaluate genetic diversity and population integrity. 44 45 Results from a haplotype network analysis found that most Pearl River haplotypes clustered with haplotypes of Yangtze River origin and only a small proportion were 46 distinct, suggesting that native Pearl River bighead and silver carp populations are 47 48 both currently dominated by genetic material from Yangtze River. Genetic diversity of Pearl River populations is high in both species because of this inter-population 49 gene flow, but native Pearl River diversity is low. We propose that, to preserve the 50 native genetic diversity, stocking of non-native fingerlings should cease immediately 51 52 and native Pearl River bighead and silver carp fish farms should be established. This research demonstrates the danger to native biodiversity across China of the substantial 53 54 ongoing stock enhancement activities without prior genetic assessment.

55

56 KEYWORDS Bighead carp, Silver carp, Pearl River, Genetic diversity, Domestic

57 introduction, Stock enhancement

#### 58 1 | INTRODUCTION

Bighead carp Hypophthalmichthys nobilis (Valenciennes 1844) and silver carp 59 Hypophthalmichthys molitrix (Richardson 1845), belonging to Cypriniformes, 60 Cyprinidae (Chen, 1998), are two major commercial species in China, which now 61 rank second and third, respectively, in Chinese freshwater aquaculture production 62 63 (MOA, 2018) and second and fifth, respectively, in global aquaculture production 64 (FAO, 2018). They are native to China, with a natural range extending from Pearl River in the south to Amur River in the north (Li et al., 1989). Since the 1950s, 65 bighead and silver carp have been introduced or spread into more than 88 countries 66 around the world, primarily for use in aquaculture, but also for plankton control and 67 fisheries enhancement (Kolar et al., 2007; Li et al., 2010; Li et al., 2011). In some of 68 69 these places, the carps have had serious negative ecological effects and are considered as invasive species (Irons et al., 2007). For example, in North America bighead and 70 71 silver carp have spread throughout the Mississippi River Basin and increased in 72 population size and biomass, resulting in negative environmental consequences and a 73 pressing issue for fisheries scientists (Sandra, 2016).

Conversely, since the 1960s alarming declines of both carp species have been observed in the Yangtze River and Pearl River due to the construction of dams, pollution, and overfishing (Li *et al.*, 2008; Mao *et al.*, 2010). For example, the Yangtze River yielded about 30 billions of fry for the four Chinese carps annually in the early 1960s, which subsequently declined to two billions in 1980 and one billion in 2010 (Mao *et al.*, 2010). Therefore, the two carp species included here are currently

classified as near threatened in China (Zhao, 2011; Huckstorf, 2012). Although both 80 species in these two rivers are decreasing quickly, the situation differs between 81 Yangtze River and Pearl River. Historically, the Yangtze River was the main 82 production area of the four major Chinese carp species in China, followed by Pearl 83 River and Amur River (Li, 1990). The fish fry used for aquaculture in these rivers 84 85 were mainly captured from wild populations until the success of artificial propagation approaches in 1958. Since then, only few wild individuals were collected for breeding 86 87 every year. More recently, the construction of the Three Gorges Dam caused concern 88 for its huge negative impact on the ecology and fisheries of the Yangtze River (Zhang et al., 2012), which led to the establishment of four national breeding farms in the part 89 of the river below the dam to restore wild fisheries. These subsequently served as the 90 91 main origin of carp fry across China, because Yangtze River populations have superior desirable traits for aquaculture, such as a bigger body size at maturity and a 92 faster growth rate (Li, 1990). 93

94 The Pearl River basin is considered the "lifeline" of terrestrial water resources in 95 China, which comprises three main tributaries: Xijiang River (usually represents the 96 Pearl River because it is the longest in the three tributaries), Beijiang River (the 97 second longest), and Dongjiang River. It has a different evolutionary history from Yangtze River (Li, 1981; Wang, 1994). For example, bighead and silver carp, which 98 originated from the Yangtze-Yellow eastern plain of China in the Pliocene about 3.5 99 100 million years ago, arrived at the Pearl River through the Yangtze River and the Qiantang River ca 110,000 years ago during the Pleistocene (Li & Fang, 1990), in the 101

102 course making the native carp populations evolutionarily distinct (Li et al., 2010; Li et 103 al., 2011). Nonetheless because of the perceived superiority of Yangtze stock, 104 bighead and silver carp stock enhancements have been carried out in Pearl River since 105 2001 (Mao et al., 2010). These activities were performed under unscientific and 106 varying rules in the basin, without knowing the genetic diversity and parental origin 107 of the released fingerlings, but which were believed to have originated from the 108 Yangtze River (Wu et al., 2016). Research on bighead carp collected from Yangtze River breeding and wild populations and a Pearl River fish farm population using 17 109 110 microsatellite loci suggested that the Pearl River-cultured population genetically 111 clustered with the Yangtze River wild population (Zhu et al., 2018), providing 112 evidence that introductions have become established. For these reasons, the native 113 Pearl River carp populations are likely in danger of being replaced by introduced carp populations from the Yangtze River. With the increasing introductions from Yangtze 114 115 River and large-scale, yearly stock enhancement of these introduced carps into Pearl 116 River, a detailed survey focusing on the genetic diversity and status of bighead and 117 silver carp are critically important for the conservation of native Pearl River 118 populations.

Although there are many published studies that addressed the genetic diversity of bighead and silver carp, most research in China has mainly focused on assessing fisheries and comparisons of genetic diversity among different populations of a species within the Yangtze River (Li, 1990; Li & Wang, 1990; Li *et al.*, 1998; Zhang *et al.*, 1999; Geng *et al.*, 2006; Shan *et al.*, 2006; Wang *et al.*, 2008; Li *et al.*, 2010;

124 Zhang et al., 2013), providing a good reference for comparison of genetic diversity of 125 potential introduced Yangtze River populations and native Pearl River populations. 126 On the other hand, studies on the genetic diversity of bighead and silver carp in Pearl 127 River (Li et al., 2010; Liu et al., 2010; Li et al., 2011; Wu et al., 2016) showed low 128 genetic diversity and proposed stock enhancement activities to recover wild fisheries. 129 However, the limitation of their sampling size constrained our understanding of the 130 real genetic diversity of carps in the Pearl River. In addition, the genetic diversity and 131 status of released carp individuals and their ecological impact on local populations 132 remain unknown. The fact that native populations can suffer from introgressive 133 hybridization with alien stocks of domestic origin has been well demonstrated, for 134 example in Atlantic brown trout (Caputo et al., 2004; Lorenzoni et al., 2006). In that 135 example, the Italian native trout showed various degrees of hybridization of two 136 mitochondrial lineages, with some populations totally replaced by non-native trout 137 (Splendiani et al., 2016), and a similar issue may occur in bighead and silver carp in 138 China.

In the present study, we aimed to (i) assess if colonization of bighead carp and silver carp from Yangtze River to Pearl River is evident and, (ii) if so, identify the extent to which the genetic material of Yangtze River bighead and silver carp have been established in the Pearl River populations, and (iii) its impact on the genetic diversity of these two species in Pearl River. The most important tributaries of Pearl River were sampled in order to evaluate the genetic diversity of native, non-stocked populations. The partial mitochondrial DNA control region (D-loop) was chosen rather than microsatellites because native populations between rivers are known to be
distinct at mtDNA marker (Li *et al.*, 2010; Li *et al.*, 2011) and this marker is suitable
for tracking the long evolutionary history of animal species (Moritz *et al.*, 1987), has
an estimated mutation rate and predictable evolution, and is comparable across studies
and species, unlike other markers such as for example microsatellites (Noor *et al.*,
2001). Based on our findings, possible conservation strategies of the remaining native
Pearl River populations of bighead and silver carp are proposed.

153

## 154 2 | MATERIALS AND METHODS

## 155 2.1 | Sampling and DNA extraction

The care and use of experimental animals complied with China animal welfare laws, guidelines and policies as approved by South China Normal University (permit reference number No.201303048). Fishes were collected for conservation purposes and to contribute to the fisheries knowledge of these species. All sampled fish were fatally anesthetized with MS-222 (Sigma).

We surveyed nine wild populations of co-existing bighead and silver carp in Pearl River basin between July 2013 and December 2015 by using gill nets. Individuals were selected according to their body size and total length. Populations were selected based on their natural distribution in the Pearl River (Chen, 1998; Pan *et al.*, 1990). The age of individuals was estimated according to length-age relationships. All individuals were juvenile (between one to three years old, standard length ranges from 26 to 44 cm; at this stage they can be easily identified). The specimens were identified

168	based on the current literature (Chen, 1998; Wang, 1994). In addition, fry of silver
169	carp were sampled from Xijiang River of Pearl River in July 2017 and raised for four
170	months in the laboratory and used for molecular analysis. A total of 213 individuals of
171	bighead carp were collected from Qingyuan (BJ, N=24), Longchuan (DJ, N=30),
172	Fengkai (XJYY, N=17) and Zhaoqing (XJ, N=23) in Guangdong Province, from
173	Shaoping (GJ, N=12), Laibin (HSH, N=30), Liuzhou (LJ, N=23), Pingguo (YJ, N=26)
174	and Guiping (YUJ, N=9) in Guangxi Province, and from Luoping (NPJ, N=36) in
175	Yunnan Province. A total of 187 individuals of adult silver carp were collected from
176	Qingyuan (BJ, N=20), Longchuan (DJ, N=30) and Zhaoqing (XJ, N=16) in
177	Guangdong Province, from Shaoping (GJ, N=5), Laibin (HSH, N=30), Liuzhou (LJ,
178	N=18), Pingguo (YJ, N=13) and Guiping (YUJ, N=21) in Guangxi Province, and
179	from Luoping (NPJ, N=34) in Yunnan Province. Seven sequences of bighead carp and
180	64 sequences of silver carp were downloaded from the NCBI database, which were
181	originally sampled from Jianli in Hubei province beside the Yangtze River (Hao et al.
182	2013; GenBank Accession Numbers for bighead carp: KC292939-KC292945;
183	GenBank Accession Numbers for silver carp: KC292923-KC292927,
184	KC292929-KC292934, KF384046-KF384088, KF384089-KF384096) (Table 1,
185	Figure 1).

For molecular analyses, a piece of muscle tissue was obtained from each individual and preserved in 95% ethanol or frozen. DNA was extracted using standard phenol-chloroform extraction protocols (Sambrook *et al.*, 1989), and the partial D-loop sequence was amplified using polymerase chain reaction (PCR) with the

190 DL1 (5'-ACCCCTGGCTCCCAAAGC-3') DH2 primers and 191 (5'-ATCTTAGCATCTTCAGTG-3') (Liu et al., 2002). Each 25 µL PCR reaction contained 1.0  $\mu$ L template DNA, 13  $\mu$ L 2×Tag PCR MasterMix (0.1U Tag DNA 192 193 Polymerase/ml, 500 mM per dNTP, 50 mM Tris-HCl (pH 8.7), 20 mM KCl, 4 mM 194 MgCl<sub>2</sub>), 10 ng of each primer, and 9 µL ddH<sub>2</sub>O. PCR amplification was conducted in 195 a thermal cycler (Eppendorf Master cycler) including a negative control using the 196 following conditions: one cycle of denaturation at 94°C for 2 min; 35 cycles of denaturation at 94°C for 45 s, annealing at 61°C for 45 s, and extension at 72°C for 197 198 1.2 min; followed by extension at 72°C for 7 min and storage at 4°C. PCR products 199 were purified by electrophoresis in 1.0% agarose gel using a 1×TAE buffer. The gel 200 was stained with ethidium bromide, and the DNA band was cut and eluted using the 201 Agarose Gel Purification Kit (QIAGEN, Valencia, CA, USA). The PCR products 202 were then subjected to cycle sequencing reactions prescribed by Sangon Biotech 203 (Shanghai) Co., Ltd using the DL1 primer. Sequences were obtained using an ABI 204 PRISM 3730XL sequencer with the BigDye Terminator kit (Applied Biosystems). All 205 sequences are deposited in GenBank under accessions MN641494-MN641562 for 206 bighead carp and MN641563-MN641680 for silver carp.

207

## 208 2.2 | Data analysis

Nucleotide sequences were aligned using Clustal X v2.1 (Thompson *et al.*, 1997).
The output files were checked by eye with MEGA v7.0 (Kumar *et al.*, 2016) and
haplotypes were identified with the default settings in DnaSP 5.10 software (Librado
& Rozas, 2009). Genetic distances between groups were calculated using MEGA v7.0

213 with a Kimura two-parameter (K2P) genetic distance model. Genetic diversity was 214 quantified based on haplotype diversity (h) and nucleotide diversity ( $\pi$ ) (Nei & Tajima, 215 1981) using DnaSP 5.10 software. The differences between populations were assessed 216 with pairwise genetic distance and genetic differentiation index (Fst) values 217 calculated in DnaSP 5.10 software. Haplotype networks were constructed using the 218 median-joining network method (Bandelt et al., 1999) in Network 4.6.1.0. software. 219 Demographic history was inferred using a mismatch analysis (Rogers & Harpending, 1992). The Tajima's D (Tajima, 1989), Fu's Fs (Fu, 1997), Harpending's raggedness 220 221 index, and the sum-of-squared deviations statistics (SSD) were calculated in Arlequin 222 3.5.2.2 (Excoffier & Lischer, 2010).

223

## 224 3 | **RESULTS**

#### 225 3.1 | Genetic diversity of bighead carp

226 The alignment of all D-loop sequences revealed 188 polymorphic positions, of 227 which 134 positions were parsimony informative. A total of 69 haplotypes were 228 identified from the 213 samples, with 68 haplotypes in Pearl River and six haplotypes 229 in the Yangtze River, five of which were shared with Pearl River (83.3%) (Table 2; 230 Supporting Information Figure S1). The overall haplotype diversity of Pearl River 231 populations combined was 0.936. The within-population haplotype diversity values 232 were high (>0.85) in all populations (Table 2). The overall nucleotide diversity of 233 Pearl River populations combined was 0.00565. The highest nucleotide diversity was 234 observed in Xijiang River population (XJ;  $\pi$ =0.00727), while the lowest was observed

in the Yujiang River population (YUJ;  $\pi$ =0.00282). Populations BJ, DJ, and LJ also had high nucleotide diversity (Table 2). Pearl River and Yangtze River populations had similar levels of genetic diversity (*h*=0.936,  $\pi$ =0.00569 vs *h*=0.952,  $\pi$ =0.00526).

#### 239 **3.2** | Genetic diversity of silver carp

240 222 polymorphic positions were detected in all D-loop sequences with 172 parsimony informative positions. 118 haplotypes were obtained of the 204 silver carp 241 242 samples. 98 haplotypes were from the Pearl River and 25 haplotypes were from the 243 Yangtze River. 20 of Yangtze River haplotypes were shared with the Pearl River (80.0%) (Table 3; Supporting Information Figure S2). The overall haplotype diversity 244 245 of Pearl River populations was 0.963 (range: 0.446-1). The within-population h values 246 were high in all populations except population DJ (Table 3). The nucleotide diversity of all Pearl River populations combined was 0.02174. The highest nucleotide 247 248 diversity was observed in Beijiang River population (BJ;  $\pi$ =0.03926), while the 249 lowest was observed in Dongjiang River population (DJ;  $\pi$ =0.00619) (Table 3). 250 Overall the genetic diversity of silver carp was higher in Pearl River (h=0.968,  $\pi$ =0.02474), than in Yangtze River (h=0.897,  $\pi$ =0.00864), though Pearl River also 251 252 had more individuals and more localities sampled.

253

#### 254 3.3 | Haplotype network analysis

255 **3.3.1** | Bighead carp

The haplotype network analysis identified two clades in bighead carp, with six mutation steps between them (Figure 2). Clade I contained 65 haplotypes found in

258 Yangtze and Pearl River and Clade II comprised only four haplotypes (5.8% of total 259 haplotypes), which are found exclusively in Pearl River (Table 2). Despite haplotype relationships between populations, the network analysis also showed that the 260 261 dominant haplotypes (e.g., H3, H11, H19) were shared between Yangtze River and 262 Pearl River samples (Figure 2). Besides, five out of six of Yangtze River haplotypes 263 were shared with Pearl River generally (Figure 2) and found in many individual 264 populations (e.g., population BJ, DJ and XJ) (Supporting Information Figure S1). For 265 example, population XJ had the H3 haplotype from Yangtze River and a H55 266 haplotype from Clade II, which is otherwise native Pearl River. Most individuals in 267 population DJ had the same haplotypes as Yangtze River samples, especially the H19 268 haplotype. Populations BJ, DJ, and LJ had one or two haplotypes grouped in Clade II 269 (Supporting Information Figure S1).

270

#### 271 **3.3.1** | Silver carp

272 In the haplotype network analyses, 118 mtDNA haplotypes fell into two 273 divergent lineages (I and II). The number of haplotypes clustered in Lineage II was 274 clearly lower than that in Lineage I (17 vs 101) (Figure 3, Table 3). Haplotypes from 275 lineage II formed two sublineages with ten mutation steps between them. The number 276 of mutations between two lineages in silver carp was considerable, at 57 steps. 277 Seventeen silver carp samples from Xijiang River (XJYY) contained both lineages of 278 this species (Table 2; Figure 3; Supporting Information Figure S2). The haplotype 279 H14 and H22 were the most frequent in sublineage A and B, respectively. All of these 280 dominant haplotypes in Lineage I were shared by Yangtze and Pearl River 281 populations (Figure 3). What is more, they were observed in most Pearl River 282 populations (e.g. populations BJ and DJ) (Supporting Information Figure S2). 283 However, population XJ did not share any frequent haplotypes with Lineage I, with 284 some unique haplotypes clustered with Lineage II. Interestingly, the most frequent 285 haplotype in population DJ was the H22 haplotype, which did not occur in other Pearl 286 River populations. Populations BJ, HSH, XJ, and XJYY had one or two haplotypes 287 grouped in Lineage II (Supporting Information Figure S2). Two populations (XJ and 288 XJYY) from the Xijiang River (downstream of Pearl River) had a similar proportion 289 of Lineage I and II haplotypes (Table 3).

290

## 291 3.4 | Genetic differentiation

We calculated the genetic distance and genetic differentiation index (*Fst*) between lineages or clades identified based on haplotype analysis. Results showed that the genetic distance between the two lineages in silver carp was larger (Table 5) than that between the two clades of bighead carp (Table 4).

The pairwise K2P genetic distances revealed low genetic differentiation among all populations of bighead carp, ranging from 0.004 to 0.007 (Supporting Information Table S1).

K2P genetic distances among most populations of silver carp were high, from
0.009 to 0.038 across sites (Supporting Information Table S2). Genetic distances
between silver carp populations with high nucleotide diversity within Pearl River (e.g.,

302 BJ and LJ) was larger than other pairs (e.g., GJ and DJ) (Supporting Information303 Table S2).

304

## 305 3.5 | Demographic history

Fu's *Fs* and Tajima's *D* statistics for bighead and silver carp were significantly negative except for Tajima's *D* for Clade II of bighead carp and Lineage I of silver carp (Supporting Information Table S3). Apart from Clade II of bighead carp, both species showed a multimodal distribution in the mismatch analyses (Supporting Information Figure S3). The Harpending's raggedness index was significant for Clade I of bighead carp and the SSD was significant for Lineage II of silver carp (Supporting Information Table S3).

313

#### 314 4 | DISCUSSION

## 315 4.1 | Evidence for colonization from Yangtze River

316 In this study, we aimed to assess if and to what extent colonization of bighead 317 and silver carp from Yangtze River has taken place. Similar to other fish species, such 318 as in the brown trout species complex (Splendiani et al., 2016), we clearly 319 demonstrated mitochondrial gene flow from the Yangtze River into the Pearl River 320 (Figure 2, 3). 'Gene flow' in this study could mean introgression, hybridization, or 321 replacement of one lineage by another by colonization or anthropogenic introduction. 322 We cannot exclude the possibility of migration but think it is unlikely because of the 323 biogeographic barriers between Pearl and Yangtze River.

324

For bighead carp, the haplotype network analysis identified two clades: clade I

325 contained 65 haplotypes from Yangtze and Pearl River and Clade II comprised only four Pearl River haplotypes (Table 2), suggestive of the close but distinct 326 327 phylogenetic relationship of Yangtze and Pearl River populations. Genetic distance 328 between these two clades was equivalent to the inter-population level (Table 4), 329 which suggests that Clade I consists of original Yangtze River haplotypes and Clade 330 II is comprised of native Pearl River fish. As shown, most haplotypes of Yangtze 331 samples were shared with Pearl River samples (Figure 2), indicating these haplotypes 332 were likely introduced from Yangtze River. Here, we argue that colonization from the 333 Yangtze River is the most likely cause for the observed pattern rather than shared 334 ancestral polymorphism for the following reasons: (1) according to Zhu et al. (2018), 335 their findings showed that samples from fish farms producing fish fry used for stock 336 enhancement activities were in fact introduced from the Yangtze River; (2) no private 337 haplotypes of Yangtze River were detected in Yangtze River samples while there are 338 some in Pearl River samples (Clade II haplotypes). Although the sample size of the 339 Yangtze River population of bighead carp was small, it is much larger than the 340 Yangtze River population of silver carp and no unique haplotypes of the Yangtze 341 River population were found either; (3) in nature, bighead carp spawns in rivers; their 342 offspring migrate to lakes and stays there until maturity (Chen, 1998; Pan et al., 1990). 343 Eggs of bighead carp can only hatch under sufficient accumulated temperature, which 344 means they cannot survive in fragmented rivers that resulted from dam construction 345 (Li & Fang, 1990; Mao et al., 2010). Populations present in these rivers (e.g., DJ and 346 upstream population, NPJ) shared most of their haplotypes with Yangtze River

samples (Supporting Information Figure S1, S2), providing a strong evidence of 347 348 introduction. However, we cannot reject the possibility that there is some shared 349 ancestral polymorphism between these two river systems. A more detailed survey of 350 all available populations in Yangtze and Pearl River using both nuclear and 351 mitochondrial markers would be valuable. Given these patterns of bighead carp, and 352 that three dominant haplotypes of Yangtze River are shared with all Pearl River 353 populations (Supporting Information Figure S1), our findings suggest that all the 354 present Pearl River populations have been invaded by fish of Yangtze River origin.

355 Similarly, for silver carp, the haplotype network analysis showed most 356 haplotypes of Pearl River clustered in Lineages I with Yangtze River haplotypes 357 (Figure 3). Given that the number of mutations (57 steps) and genetic differentiation 358 between two lineages in silver carp in Pearl River was considerable (Table 5; Figure 359 3), it is unlikely that the two lineages evolved in the Pearl River because there are no 360 geographic barriers in the river system and no similar results have been found in other 361 fishes at this genetic marker in the basin (Han et al., 2010; Yang et al., 2016; Li et al., 362 2018). Thus, a reasonable explanation for the results is that Lineage I consists of fish 363 of the original Yangtze River population while Lineage II is formed by pure Pearl 364 River fish.

According to Li & Fang (1990), the divergence time of the Yangtze and Pearl River populations of silver carp was about 110,000 years ago during the Pleistocene, which could be old enough for the evolution of the observed divergence. Two sublineages detected in Lineage I (native Yangtze River population) (Figure 3; 369 Supporting Information Figure S2) were consistent with a previous study on native Yangtze River populations (Sha et al., 2018). Apart from mtDNA evidence given by 370 371 this study, eight microsatellite markers used on Pearl River (sampled from 372 Nanpangjiang River, upper Pearl River) and Yangtze River (sampled from middle 373 location of Yangtze River) wild populations as well as Pearl River culture stocks 374 (sampled from a fish farm in lower Pearl River) showed similar genetic structure and 375 high gene flow among them (unpublished data), suggestive of the same pattern as 376 seen in bighead carp showing that these Pearl River fish farm populations originated 377 from Yangtze River and therefore facilitate the genetic pollution by Yangtze River 378 silver carp.

379 As in bighead carp, we cannot exclude the possibility that there is shared 380 ancestral polymorphism in silver carp between these two river systems, but we 381 consider it unlikely. Given that the 17 silver carp fry (population XJYY) sampled 382 from Xijiang River consisted of two divergent lineages (putatively native Yangtze and 383 Pearl River populations, respectively) (Figure 3), we can conclude that the Yangtze 384 River lineage of silver carp has founded a natural population in this main stream of 385 Pearl River. What is more, haplotypes in Lineage I were shared by Yangtze and all 386 Pearl River populations (Figure 3; Supporting Information Figure S2), demonstrating 387 that mitochondrial gene flow has affected sampled populations.

388 The Fu's Fs and Tajima's D statistics for bighead and silver carp were not 389 consistent with the mismatch distribution analysis. For example, for Clade II the 390 mismatch distribution of bighead carp was unimodal while the Fu's Fs and Tajima's D

statistics were not significant. However, this non-significant result could be because of the small sample size of Clade II. What is more, combined with the low nucleotide diversity and high haplotype diversity patterns of these two species, the significantly negative values for Fu's Fs and Tajima's D of all other clades indicate these two species likely experienced a bottleneck followed by a rapid population expansion event. These results could be related to sea level and climate change during the Pleistocene, and warrant further investigation.

In sum, we concluded that mitochondrial gene flow from the Yangtze River into the Pearl River has occurred extensively. This conclusion was also strengthened by knowledge of the authors that bighead and silver carp were introduced at the same time in aquaculture in China. Importantly, Clade I and II identified in bighead carp and Lineages I and II identified in silver carp corresponded with native Yangtze and Pearl River populations, respectively.

404

# 405 4.2 | Impacts of colonization on genetic diversity

As discussed above, mitochondrial gene flow between Yangtze River and Pearl River was observed across all sampled populations. Hence, it is reasonable that the high genetic diversity of bighead and silver carp (Table 2, 3; Supporting Information Figure S1) was found due to mitochondrial gene flow.

In bighead carp, the haplotype diversity was high (h=0.936,) while the
nucleotide diversity was relatively low (π=0.00569) (Table 2), indicating that
populations in the Pearl River may have experienced a population expansion (Grant
& Bowen, 1998), consistent with recent population growth after the introduction of

414 Yangtze River individuals. This agrees with previous published results (Li et al., 2010) 415 showing that the genetic diversity of the Pearl River population was slightly higher 416 than that of Yangtze River. The total genetic diversity of all sampled populations was 417 slightly higher than that of population CJ (Table 2) and introduced populations from 418 the Yangtze River (Wu *et al.*, 2016), supporting mitochondrial gene flow as the cause 419 for increased genetic diversity. Our results are also consistent with a previous study 420 about genetic diversity of bighead carp in Hongshuihe River (upstream of the Pearl 421 River) (Wu et al., 2016). For individual populations, impacts of mitochondrial gene 422 flow from Yangtze River into Pearl River were also found. For example, population 423 XJ has the highest nucleotide diversity and its haplotypes grouped in both Clade I and 424 II of bighead carp, indicating the contact of divergent clades in this population – and 425 other populations such as BJ, DJ, and LJ - resulted in the increased genetic diversity. In silver carp, the genetic diversity was higher in Pearl River than in Yangtze 426 427 River, also suggesting evidence of colonization from Yangtze River. When focusing 428 on the genetic diversity of populations, several populations (population XJ, XJYY, BJ, 429 etc.) have values exceeding the average. Based on the haplotype network analysis 430 (Supporting Information Figure S2), we can see that all these populations have 431 haplotypes of both Lineage I and Lineage II. Native Pearl River populations without 432 non-native mitochondrial haplotypes may still exist, and future research using both 433 mtDNA and nuclear molecular makers will improve our understanding of where the 434 native carp populations are in the Pearl River system.

435 Interestingly, the population DJ which was sampled from the Dongjiang River,

436 one main tributary of Pearl River, showed a low haplotype diversity (h=0.446). Since 437 the natural environments in the Dongjiang River could no longer meet the 438 requirements for the breeding of silver carp due to the cascade power station, it is very 439 likely that the current population was introduced from Yangtze River.

440 However, the genetic diversity of the native Pearl River clades in both bighead 441 and silver carp (i.e. Clade II (Table 2) or Lineage II (Table 3)) was extremely low. 442 Moreover, the genetic diversity of native bighead and silver carp in the Pearl River 443 was lower compared to Squaliobarbus curriculus, an important economic species 444 mainly distributed in the Pearl River, which was surveyed at the same time and 445 showed high haplotype diversity and nucleotide diversity based on D-loop sequences 446 (h=0.982,  $\pi$ =0.01353) (Li *et al.*, 2018). Other native economic freshwater fishes that 447 were not extensively cultured or introduced in the Pearl River, such as Schizothorax lissolabiatu (Han et al., 2010) and Culter alburnus (Yang et al., 2016), also had a 448 449 higher genetic diversity revealed by the same molecular marker. Overall, our results 450 suggest that human activities such as introductions of Yangtze River populations, dam 451 constructions (river fragments), and stock enhancement activities have already had a 452 negative impact on the genetic diversity of native bighead and silver carp.

453

# 454 **4.3** | Implications for conservation and population management

Knowledge on the amount of genetic diversity existing within and among populations, its distribution through the species range and its temporal stability over generations are key issues to design appropriate management and conservation strategies (Vera *et al.*, 2019). As two major domestic fishes in China and important 2 459 economic fishes in the Pearl River, bighead and silver carp have been used for 460 aquaculture and culture-based capture fisheries in natural systems for thousands of 461 years (Mao et al., 2010). However, due to the lack of proper resource assessments and 462 management plans, the released fry used for stock enhancement in Pearl River have 463 mainly been taken from hatchery stocks derived from the Yangtze River fish. It is 464 fortunate that we found a small number of native bighead and silver carp populations 465 in the Pearl River. However, Yangtze River populations have colonized all the 466 tributaries of the Pearl River system with the potential for hybridization with the 467 native Pearl River populations, which has also taken place in a similar manner in Mediterranean brown trout (Splendiani et al., 2019). Moreover, a large number of the 468 469 fish fry of these two species derived from Yangtze River parents were released in the 470 Pearl River every year without assessment of their genetic background.

Even though the genetic diversity of these two carps in the Pearl River was higher than in Yangtze River, this is likely due to the pronounced mitochondrial gene flow from Yangtze River. Thus, in the light of this new knowledge, we propose that stock enhancement should be stopped immediately until fish farms with native Pearl River bighead and silver carp are founded. At the same time, regulatory rules and actions on the fry sources used in releasing activities should be organized and strengthened.

478 Meanwhile, we propose a wide-range survey focusing on the genetic resources of 479 native Pearl River bighead and silver carp based on both mtDNA and nuclear markers 480 (e.g., microsatellites, SNPs, nuclear genes), as has been performed in Yangtze River

481 and Western Europe. This kind of survey is fundamental to the establishment of 482 national aquatic germplasm reserves for economically important species (Li et al., 483 1998; Geng et al., 2006; Wang et al., 2008; Berrebi et al., 2019; Splendiani et al., 484 2019; Vera et al., 2019) and will determine if admixture is occurring or if the 485 Yangtze-origin haplotypes are from released individuals. In the future, fish from the 486 Pearl River should be used to preserve the genetic diversity of native carp populations. 487 Moreover, based on the haplotype network analysis, population XJ had haplotypes 488 clustered in Clade I and II (bighead carp) (Supporting Information Figure S1) and 489 Lineages I and II (silver carp) (Supporting Information Figure S2). Therefore, Xijiang 490 River could be an appropriate site for setting up a state-level reserve for native 491 populations of various species in Pearl River, including two other carp species, black 492 carp (Mylopharyngodon piceus) and grass carp (Ctenopharyngodon idellus), to 493 provide high-quality germplasm not only for southern China but for the entire country and even the world. 494

495

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506

## 507 Author contributions

508 C. L., J. Z. and K. R. E. contributed to the study design; C. L. and J. Q. C.

509 contributed to the sample preparation and molecular experiments; C. L. and K. S.

510 performed the bioinformatic analyses; C. L., J. J. W., R. K. V. and K. S. wrote the

- 511 paper with significant input from K.R.E.. All authors read and approved the final
- 512 manuscript.
- 513

## 514 Conflicts of interest

515 The authors declare no conflicts of interest.

#### 516

#### 517 **REFERENCES**

- Bandelt, H.J., Forster, P., & Röhl, A. (1999). Median-joining networks for inferring
  intraspecific phylogenies. *Molecular Biology and Evolution*, 16(1), 37-48.
- 520 Berrebi, P., Barucchi, V.C., Splendiani, A., Muracciole, S., Sabatini, A., Palmas, F.,
- 521 Tougard, C., Arculeo, M., & Marić, S. (2019). Brown trout (*Salmo trutta* L.) 522 high genetic diversity around the Tyrrhenian Sea as revealed by nuclear and

523 mitochondrial markers. *Hydrobiologia*, **826**(1), 209-231.

- Grant, W.S., Bowen, B.W. (1998). Shallow population histories in deep evolutionary
  lineages of marine fishes: insights from sardines and anchovies and lessons for
  conservation. *Journal of Heredity*, **89**(5), 415-426.
- 527 Caputo, V., Giovannotti, M., Nisi, Cerioni, N., Caniglia, M.L., & Splendiani A.
  528 (2004). Genetic diversity of brown trout in central Italy. *Journal of Fish Biology*,
  529 65(2), 403-418.
- 530 Chen, Y.Y. (1998). Fuana Sinica Osteichthyes Cypriniformes II. Beijing: Science
  531 Press, 104-106. (in Chinese)

- Excoffier, L., & Lischer, H. E. L. (2010). Arlequin suite ver 3.5.2.2: A new series of
  programs to perform population genetics analyses under Linux and Windows. *Molecular Ecology Resources*, 10, 564-567.
- 535 FAO. (2018). Fishery and aquaculture statistics: 2016. Food and Agricultural
  536 Organization of the United Nations.
- Fu, Y. X. (1997). Statistical tests of neutrality of mutations against population growth,
  hitchhiking and background selection. *Genetics*, 147, 915-925.
- Geng, B., Sun, .XW., Liang, L.Q., OuYang, H.S., & Tong, J.G. (2006). Microsatellite
  Analysis of Genetic Diversity of *Aristichthys nobilis* in China. *Hereditas*, 28(6),
  683-688.
- 542 Grant, W.S., & Bowen, B.W. (1998). Shallow population histories in deep
  543 evolutionary lineages of marine fishes: insights from sardines and anchovies and
  544 lessons for conservation. *Journal of Heredity*, **89**(5), 415-426.
- 545 Han, H.F., & Dai, Y.G. (2010). Sequence polymorphism of mtDNA D-loop in the
  546 population of *Schizothorax lissolabiatus* from Kedu River in Pearl River system.
  547 *Journal of Fishery Sciences of China*, 17(1), 143-149. (in Chinese)
- Hao, J., Yang, Q., Bang, D., Liang, A.J., Zhang, X.H., & Dong, S. (2013). The
  sequence comparison of mt DNA D-loop and adjacent regions in six fish species. *Jounal of Dalian Ocean University*, 28(2), 160-165. (in Chinese)
- Huckstorf, V. (2012). *Hypophthalmichthys nobilis*. The IUCN Red List of Threatened
   Species. 2012:
- e.T166172A1116524. doi:10.2305/IUCN.UK.2012-1.RLTS.T166172A1116524.
  e
- Irons, K.S., Sass G.G., McClelland M.A., & Stafford J.D. (2007). Reduced condition
  factor of two native fish species coincident with invasion of non native Asian
  carps in the Illinois River, USA Is this evidence for competition and reduced
  fitness? *Journal of Fish Biology*, 71(sd), 258-273.
- Kolar, C.S., Chapman, D.C., Courtenay, J.W.R., Housel, C.M., Williams, J.D., &
  Jennings, D.P. (2007). Bigheaded carps: a biological synopsis and environmental
  risk assessment, vol Special Publication 33. American Fisheries Society,
  Bethesda.
- Kumar, S., Stecher, G., & Tamura, K. (2016). MEGA7: Molecular evolutionary
  genetics analysis version 7.0 for bigger datasets. *Molecular Biology and Evolution*, 33, 1870-1874.
- Li, C., Lan, Z.J., & Zhao, J. (2018). Genetic diversity and genetic differentiation of *Squaliobarbus curriculus* from six geographical populations of the Pearl River
  based on mitochondrial D-loop sequences. *Chinese Journal of Applied and Environmental Biology*, 24(3), 0615-0622. (in Chinese)
- 570 Li, S.F. (1990). Genetic resources of silver carp, bighead carp and grass carp in
  571 Changjiang river, Zhujiang river and Heilongjiang river. Shanghai: Shanghai
  572 Science and Technology Press, 25-50. (in Chinese)
- 573 Li, S.Z. (1981). Studies on zoogeographical divisions for freshwater fishes of China.
  574 Beijing, China: Science Press. (in Chinese).
- 575 Li, S.F., Lv, G.Q., & Louis, B. (1998). Diversity of mitochondrial DNA in the

- 576 population of silver carp, bighead carp, grass carp and black carp in the 577 middle-and lower reaches of the Yangtze river. *Acta Zoologica Sinica*, **44**(1), 578 82-93. (in Chinese)
- 579 Li, S.Z., & Fang, F. (1990). On the geographical distribution of the four kinds of
  580 pond-cultured carps in China. *Acta Zoologica Sinica*, 36, 244-248. (in Chinese)
- 581 Li, S.F., & Wang, R.X. (1990). Maturity speed and genetic analysis of silver carp and
  582 bighead carp from Changjiang and Zhujiang river systems. *Journal of fisheries of*583 *China*, 14(3), 189-197. (in Chinese)
- Li, S.F., Xu, J.W., Yang, Q.L., Wang, C.H., Chapman, D.C., & Lu, G.Q. (2011).
  Significant genetic differentiation between native and introduced silver carp
  (*Hypophthalmichthys molitrix*) inferred from mtDNA analysis. *Environmental Biology of Fishes*, 92, 503-511.
- Li, S.F., Yang, Q.L., Xu, J.W., Wang, C.H., Chapman, D.C., & Lu, G.Q. (2010).
  Genetic diversity and variation of mitochondrial DNA in native and introduced
  bighead carp. *Transactions of the American Fisheries Society*, 139(4), 937-946.
- Li, S.F., Zhou, B.Y., Ni, C.K., & Chen, Z.Q. (1989). Morphological variations of
  silver carp, bighead and grass carp from Changjiang, Zhujiang and Heilongjiang
  rivers. *Acta Zoologica Sinica*, 35(4), 390-398 (in Chinese)
- Li, Y.F., Li, X.H., Tan, X.C., Li, J., Wang, C., & Luo, J.R. (2008). The present situation and change of fishery resources in Zhaoqing of Xijiang river. *Reservoir Fisheries*, 28(2), 80-83 (in Chinese)
- Librado, P., & Rozas, J. (2009). DNASP v5: a software for comprehensive analysis of
  DNA polymorphism data. *Bioinformatics*, 25(11), 1451-1452.
- Liu, H., Tzeng, C., & Teng, H. (2002). Sequence variations in the mitochondrial DNA
  control region and their implications for the phylogeny of the Cypriniformes. *Canadian Journal of Zoology*, 80(3), 569-581.
- 602 Liu, L., Zeng, S.M., Tang, S.J., Chen, W.J., Li, Y.M., Wang, Y.M., & Fu, W.L. (2010)
- Study on genetic diversity of *Aristichthys nobilis* different geographical groups
  in the pearl river in Guangdong province. *Animal Husbandry and Veterinary Medicine*, 204. (in Chinese)
- Lorenzoni, M., Mearelli, M., & Ghetti, L. (2006). Native and exotic fish species fish
  in the tiber River watershed (Umbria–Italy) and their relationship to the
  longitudinal gradient. *Bull Fr Pêche Piscic*, 382,19-44.
- Mao, R.J., Zhang, Y.B., Zheng, W., Du, X.Y., & Sun, X.W. (2010). The Progress of
   Germplasm Resources of Four Major Chinese Farmed Carps. *Chinese Journal*
- 611 *Fisheries*, **23**(3), 52-59. (in Chinese)
- MOA. (2018). China Fishery Statistical Yearbook: 2017. *Ministry of Agriculture and Rural Affairs of People's Republic of China*. (in Chinese)
- Moritz, C., Dowling, T., & Brown, W. (1987). Evolution of animal mitochondrial
  DNA: Relevance for population biology and systematics. *Annual Review of Ecology and Systematics*, 18(1), 269-292.
- 617 Nei, M., Tajima, F. (1981). DNA polymorphism detectable by restriction 618 endonucleases. *Genetics*, **97**, 145-163.
- 619 Noor M.A.F., Kliman R.M., Machado C.A. (2001). Evolutionary history of

- 620 microsatellites in the obscura group of *Drosophila*. *Molecular Biology and* 621 *Evolution*, **18**, 551-556.
- Pan, J.H., Zhong, L., Zheng, C.Y., Wu, H.L., & Liu, J.Z. (1991). The freshwater
  fishes of Guangdong Province. Guangdong Science and Technology Press,
  Guangzhou. (in Chinese)
- Rogers, A., & Harpending, H. (1992). Population growth makes waves in the
  distribution of pairwise genetic differences. *Molecular Biology and Evolution*, 9,
  552-569.
- Sambrook, J., Fritsch, E.F., & Maniatis, T. (1989). Molecular cloning: a laboratory
  manual. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York.
- 630 Sandra, L.C. (2016). Anticipating the spread and ecological effects of invasive
  631 bigheaded carps (*Hypophthalmichthys* spp.) in North America: a review of
  632 modeling and other predictive studies. *Biological invasions*, 18, 315-344.
- Sha, H., Luo, X.Z., Li, Z., Zou, G.W., & Liang, H.W. (2018). Genetic diversity of six
  silver carp (*Hypophthalmichthys molitrix*) geographical populations based on
  mitochondrial COI sequences. *Journal of Fishery Sciences of China*, 25(4),
  783-792. (in Chinese).
- 637 Shan, Q., Dong, S., Wu, H.F., & Taniguchi, N. (2006). Diversity analysis on mtDNA
  638 D-loop region of three populations of *Aristichthys nobilis*. *Journal of Fishery*639 *Sciences of China*, 13(2), 174-180. (in Chinese)
- Splendiani, A., Giovannotti, M., Righi, T., Fioravanti, T., Cerioni, P.C., Lorenzoni,
  M., Carosi, A., Porta, G.L., & Barucchi, V.C. (2019). Introgression despite
  protection: the case of native brown trout in Natura 2000 network in Italy. *Conservation Genetics*. https://doi.org/10.1007/s10592-018-1135-y.
- Splendiani, A., Ruggeri, P., Giovannotti, M., Pesaresi, S., Occhipinti, G., Fioravanti,
  T., Lorenzoni, M., Cerioni, P.C., & Barucchi, V.C. (2016). Alien brown trout
  invasion of the Italian peninsula: the role of geological, climate and
  anthropogenic factors. *Biological Invasions*, 18(7), 2029-2044.
- Tajima, F. (1989). Statistical method for testing the neutral mutation hypothesis by
  DNA polymorphism. *Genetics*, **123**, 585-595.
- Thompson, J.D., Gibson, T.J., Plewniak, F., François, J., & Desmond, G.H. (1997).
  The Clustal X win-dows interface: Flexible strategies for multiple sequences
  align-ment aided by quality analysis tools. *Nucleic Acids Research*, 25(24),
  4876-4882.
- Vera, M., Bouza, C., Casanova, A., Heras, S., Martínez, P., & García-Marín, J.L.
  (2019). Identification of an endemic Mediterranean brown trout mtDNA group
  within a highly perturbed aquatic system, the Llobregat River (NE Spain).
  Hydrobiologia, 827(1), 277-291.
- Wang, Z.Y. (1994). The Pearl River annals. Guangzhou: Guangdong Science Press.(in Chinese)
- Wang, C.Z., Liang, H.W., Zou, G.W., Luo, X.Z., Li, Z., Tian, H., & Hu, G.F. (2008)
  Genetic variation analysis of two silver carp populations in the middle and upper
  Yangtze River by microsatellite. *Hereditas*, **30**(10), 1341-1348. (in Chinese)
- 663 Wu, W.J., Peng, M., Han, Y.Q., Shi, J., Lei, J.J., Wang, D.P., & He, A.Y. (2016).

- 664 Comparison of mitochondrial D-loop and Cyt *b* sequences of *Aristichthys nobilis*.
  665 *Popular Science & Technology*, **18**(197), 26-28. (in Chinese)
- Yang, Z.T., Sun, J.J., Li, G.F., Xiao, S.B., Zhang, H.Q., Yang, H.R., Zhao, H.H., &
  Liu, L. (2016). Genetic diversity and genetic differentiation of *Culter alburnus*from eleven geographical populations of the Pearl River based on mitochondrial
  D-loop gene. *Acta Scientiarum Naturalium Universitatis Sunyatseni*, 55(2),
  89-95. (in Chinese)
- Kang, D.C., Zhang, X.Y., Yang, D.S., Yu, L.N., Fang, Y.L., Deng, F.J., & Liu, S.Y.
  (1999). Studies on Genetic Diversity of Bighead Carp (*Aristichthys nobilis*) in
  the Yangtze River. *Journal of Wuhan University (Nature Science Edition)*,
  45(6), 857-886. (in Chinese)
- Zhang, M.Q., Liu, K., Xu, D.P., Duan, J.R., Zhou, Y.F., Fang, D.A., & Shi, W.G.
  (2013). Analysis of Genetic Diversity in Populations Released for Stock
  Enhancement and Population Caught in Natural Water of Bighead Carp in the
  Lower Reaches of the Yangtze River Using Microsatellite Markers. *Acta Agriculturae Universitatis Jiangxiensis*, 35(3), 579-586. (in Chinese)
- Zhang, G., Wu, L., Li, H.T., Liu, M., Cheng, F., Murphy, B.R., & Xie, S.G. (2012).
  Preliminary evidence of delayed spawning and suppressed larval growth and
  condition of the major carps in the Yangtze River below the Three Gorges Dam. *Environmental Biology of Fishes*, 93, 439-447.
- 684 Zhao, H. (2011). *Hypophthalmichthys molitrix*. IUCN Red List of Threatened
  685 Species. IUCN. 2011:
- 686 e.T166081A6168056. doi:10.2305/IUCN.UK.2011-2.RLTS.T166081A6168056.
  687 en.
- Zhu W.B., Fu J.J., Fang M., Wang L.M., Dong Z.J., & Liu Q.G. (2018). Genetic
  analysis for nine populations of bighead carp (*Hypophthalmichthys nobilis*) using
- 690 microsatellites. The Annual Conference of the Asian Society of Ichthyologists,
- 691 2018, Nanning, Guangxi.

- 692 Tables
- 693 TABLE 1 Sampling locations, location codes, and sample sizes of bighead
- 694 Hypophthalmichthys nobilis and silver carp Hypophthalmichthys molitrix. XJYY: Fry
- 695 of silver carp sampled from the Xijiang River and raised for four months in laboratory.
- 696 Yangtze River\*: Sequences downloaded from NCBI database.
- 697 TABLE 2 Genetic diversity of bighead carp Hypophthalmichthys nobilis based on
- 698 D-loop sequences. Refer to Table 1 for the abbreviations of populations. Clade I and
- 699 Clade II are classified by the haplotype network analysis.
- 700 TABLE 3 Genetic diversity of silver carp Hypophthalmichthys molitrix based on
- 701 D-loop sequences. Refer to Table 1 for the abbreviations of populations. Lineage I
- and Lineage II are classified by the haplotype network analysis.
- 703 TABLE 4 Matrix of genetic distance of bighead carp Hypophthalmichthys nobilis
- 704 based on D-loop sequences. Two sequences of silver carp sampled from the Pearl
- 705 River (HM01, HM02) were used as an out-group.
- 706 TABLE 5 Matrix of genetic distance of silver carp Hypophthalmichthys molitrix
- 707 based on D-loop sequences. Two sequences of bighead carp sampled from the Pearl
- 708 River (HN01, HN02) were used as an out-group.
- 709
- 710

- 711 Figures
- 712 **FIGURE 1** Map showing the sampling sites of bighead carp *Hypophthalmichthys*
- 713 nobilis and silver carp Hypophthalmichthys molitrix in the Pearl River. Refer to Table
- 1 for the abbreviations of populations. The map at the top left corner shows the
- 715 relative location of the research area in China.





FIGURE 2 The haplotype network of bighead carp *Hypophthalmichthys nobilis*based on D-loop sequences. The arabic numerals near the dash line mean number of
mutations. Clade II is the native Pearl River genetic signature. Samples from the
Yangtze River are coloured blue, samples from the Pearl River are coloured red.



FIGURE 3 The haplotype network of silver carp *Hypophthalmichthys molitrix* based on D-loop sequences. The arabic numerals near the dash line mean number of mutations. Lineage II is the native Pearl River genetic signature. A, B are two sublineages of lineage I. Samples from the Yangtze River are coloured blue, samples from the Pearl River are coloured red.



#### 730 Supporting Information

- 731 TABLE S1 Matrix of genetic distance (below diagonal) and
- 732 genetic differentiation index Fst (above diagonal) of bighead Hypophthalmichthys
- 733 nobilis based on D-loop sequences. BJ: Beijiang River; CJ: Yangtze River; DJ:
- 734 Dongjiang River; GJ: Guijiang River; HSH: Hongshuihe River; LJ: Liujiang River;
- 735 NPJ: Nanpanjiang River; XJ: Xijiang River; YJ: Youjiang River; YUJ: Yujiang River.
- 736 TABLE S2 Matrix of genetic distance (below diagonal) and
- 737 genetic differentiation index Fst (above diagonal) of silver carp Hypophthalmichthys
- 738 molitrix populations based on D-loop sequences. BJ: Beijiang River; CJ: Yangtze
- 739 River; DJ: Dongjiang River; GJ: Guijiang River; HSH: Hongshuihe River; LJ:
- 740 Liujiang River; NPJ: Nanpanjiang River; XJ: Xijiang River; XJYY: fry sampled from
- 741 Xijiang River; YJ: Youjiang River; YUJ: Yujiang River.
- 742 **TABLE S3** Demographic indices of bighead carp *Hypophthalmichthys nobilis* and
- railver carp *Hypophthalmichthys molitrix* based on D-loop sequences.
- 744 FIGURE S1 The haplotype network of bighead carp Hypophthalmichthys nobilis
- 745 based on D-loop sequences. The arabic numerals near the dash line mean number of
- 746 mutations. BJ: Beijiang River; CJ: Yangtze River, its haplotypes were indicated by
- 747 the arrows; DJ: Dongjiang River; GJ: Guijiang River; HSH: Hongshuihe River; LJ:
- 748 Liujiang River; NPJ: Nanpanjiang River; XJ: Xijiang River; YJ: Youjiang River; YUJ:
- 749 Yujiang River.
- 750 FIGURE S2 The haplotype network of silver carp Hypophthalmichthys molitrix
- 751 based on D-loop sequences. The arabic numerals near the dash line mean number of

752	mutations. BJ: Beijiang River; CJ: Yangtze River; DJ: Dongjiang River; GJ: Guijiang
753	River; HSH: Hongshuihe River; LJ: Liujiang River; NPJ: Nanpanjiang River; XJ:
754	Xijiang River; XJYY: fry sampled from Xijiang River; YJ: Youjiang River; YUJ:
755	Yujiang River.
756	FIGURE S3 Mismatch distributions of bighead carp Hypophthalmichthys nobilis and
757	silver carp Hypophthalmichthys molitrix based on D-loop sequences. Light green

curves show the expected distribution of mutations according to the null hypothesis of

759 constant population size. The number of pairwise differences and their frequencies is

760 shown on the horizontal and vertical axes, respectively.

- 761
- 762 Supporting Information
- 763 TABLE S1 Matrix of genetic distance (below diagonal) and genetic differentiation index Fst (above diagonal) of bighead carp
- 764 Hypophthalmichthys nobilis populations based on D-loop sequences. BJ: Beijiang River; CJ: Yangtze River; DJ: Dongjiang River; GJ: Guijiang

765	River; HSH: Hongshuihe Ri	ver; LJ: Liujiang	River; NPJ: Nanpar	njiang River; XJ: Xiji	ang River; YJ: Youji	ang River; YUJ: Yujiang River
		ý 1 C		J U / J		

Population	BJ	CJ	DJ	GJ	HSH	LJ	NPJ	XJ	YJ	YUJ
BJ	-	-0.045	0.044	-0.008	-0.001	-0.024	0.037	0.013	-0.004	0.087
CJ	0.005	-	0.032	-0.061	-0.066	-0.046	-0.025	-0.031	-0.065	0.083
DJ	0.007	0.006	-	0.049	0.039	0.030	0.001	0.003	0.064	0.182
GJ	0.006	0.005	0.006	-	-0.038	-0.030	-0.003	-0.025	-0.043	0.109
HSH	0.005	0.004	0.006	0.004	-	-0.017	-0.008	-0.017	-0.019	0.124
LJ	0.006	0.006	0.007	0.006	0.005	-	0.003	-0.016	-0.013	0.060
NPJ	0.006	0.005	0.006	0.005	0.005	0.006	-	-0.016	0.015	0.165
XJ	0.006	0.006	0.007	0.006	0.005	0.007	0.006	-	-0.010	0.118

YJ	0.005	0.004	0.006	0.005	0.004	0.005	0.005	0.006	-	0.150
YUJ	0.005	0.004	0.006	0.004	0.004	0.005	0.005	0.005	0.004	-

769 TABLE S2 Matrix of genetic distance (below diagonal) and genetic differentiation index Fst (above diagonal) of silver carp

770 Hypophthalmichthys molitrix populations based on D-loop sequences. BJ: Beijiang River; CJ: Yangtze River; DJ: Dongjiang River; GJ: Guijiang

771 River; HSH: Hongshuihe River; LJ: Liujiang River; NPJ: Nanpanjiang River; XJ: Xijiang River; XJYY: fry sampled from Xijiang River; YJ:

Population	BJ	CJ	DJ	GJ	HSH	LJ	NPJ	XJ	XJYY	YJ	YUJ
BJ	-	0.248	0.277	0.231	0.029	0.195	0.066	0.012	-0.037	0.192	0.099
СЈ	0.034	-	0.402	-0.061	0.105	0.194	0.091	0.106	0.219	0.063	0.019
DJ	0.033	0.013	-	0.385	0.191	0.150	0.150	0.186	0.239	0.225	0.223
GJ	0.035	0.009	0.014	-	0.068	0.132	0.062	0.069	0.193	-0.027	-0.019
HSH	0.037	0.022	0.022	0.022	-	0.060	-0.020	-0.035	-0.006	0.038	-0.007
LJ	0.034	0.014	0.012	0.014	0.023	-	0.031	0.073	0.135	-0.007	0.076
NPJ	0.036	0.019	0.019	0.020	0.027	0.020	-	-0.016	0.027	0.011	-0.007
XJ	0.038	0.024	0.024	0.024	0.030	0.025	0.029	-	-0.010	0.054	-0.007

772 Youjiang River; YUJ: Yujiang River.

XJYY	0.038	0.030	0.029	0.030	0.033	0.029	0.033	0.035	-	0.134	0.070
YJ	0.034	0.012	0.013	0.012	0.022	0.013	0.020	0.025	0.029	-	0.011
YUJ	0.036	0.016	0.018	0.017	0.026	0.019	0.024	0.028	0.032	0.018	-

						Harpending's
	Species	Group	Tajima's D	Fu's Fs	SSD	Raggedness
_						index
	Bighead carp					
		Clade I	-2.06816*	-24.36304*	0.00778	0.01815*
		Clade II	-0.66823	-0.33158	0.03639	0.11000
	Silver carp					
		Lineage I	-0.77187	-5.70386*	0.51856	0.00513
_		Lineage II	-1.58286*	-23.94610*	0.01309*	0.02973

773	TABLE S3 Demographic indices of bighead carp Hypophthalmichthys nobilis and
774	silver carp <i>Hypophthalmichthys molitrix</i> based on D-loop sequences.

775 SSD: the sum of squared deviation. \*Statistically significant results (P < 0.05).



2 FIGURE S1 The haplotype network of bighead carp Hypophthalmichthys nobilis

3 based on D-loop sequences. The arabic numerals near the dash line mean number of

4 mutations. BJ: Beijiang River; CJ: Yangtze River, its haplotypes were indicated by

5 the arrows; DJ: Dongjiang River; GJ: Guijiang River; HSH: Hongshuihe River; LJ:

6 Liujiang River; NPJ: Nanpanjiang River; XJ: Xijiang River; YJ: Youjiang River; YUJ:

7 Yujiang River.



1

2 **FIGURE S2** The haplotype network of silver carp *Hypophthalmichthys molitrix* 

3 based on D-loop sequences. The arabic numerals near the dash line mean number of

4 mutations. BJ: Beijiang River; CJ: Yangtze River; DJ: Dongjiang River; GJ: Guijiang

5 River; HSH: Hongshuihe River; LJ: Liujiang River; NPJ: Nanpanjiang River; XJ:

6 Xijiang River; XJYY: fry sampled from Xijiang River; YJ: Youjiang River; YUJ:

7 Yujiang River.