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Abstract

Microplastics (MPs, <5 mm in size) are classified as emerging contaminants but treatment
processes are not designed to remove these small particles. Wastewater treatment systems
have been proposed as pathways for MPs pollution to receiving waters but quantitative and
qualitative data on MP occurrence and transport remains limited, hindering risk assessment
and regulation. Here, for the first time, the stepwise abundance and loading of MPs (60-2800
μm) in a tertiary wastewater treatment plant in the UK was assessed by sampling from May
2017 to February 2018. Microplastics were found in all sampling campaigns, with an
average inflow of 8.1 x 10 ⁸ (95% CI, 3.8 x 10 ⁸ to 1.2 x 10 ⁹) items day ⁻¹ . Their prevalence
decreased from influent to final effluent. Overall abundances decreased on average by 6%,
68%, 92%, and 96% after the pre-treatment, primary, secondary, and tertiary treatment
stages respectively, although considerable variability occurred throughout the year.
Sufficient particles remained in the treated effluent to generate an average discharge of 2.2
\times 10 ⁷ (95% CI, 1.2 \times 10 ⁷ to 3.2 \times 10 ⁷]) particles day ⁻¹ to the recipient river. Secondary MPs
were predominant, while primary MP abundances were minimal. Fibres comprised 67% of
all items, followed by films (18%) and fragments (15%). Chemical characterisation
confirmed the presence of different types of polymers, with polypropylene fibres and
fragments most abundant (23%). This research informs understanding of how wastewater
effluent may channel MPs to the natural environment and their composition, and helps
understand control points for optimising advanced treatment processes.

Keywords: microplastic pollution, WWTP, sewage, effluent discharge, FTIR-ATR

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1 Introduction

Microplastics (MPs; <5 mm) are ubiquitous in the environment and may pose a threat to biota and humans (Anbumani & Kakkar 2018), thus are classed as emerging contaminants but remain unregulated by water quality standards. This may be largely because they have not been fully assessed due to their heterogeneous nature and high spatio-temporal variations, even within localized environmental compartments. Furthermore, a lack of standardized protocols leads to limited comparability across available surveys and a lack of guidelines to monitor MPs in aquatic systems. Current empirical data is still too limited to fully understand the extent of their pollution and the severity of their threat, making it difficult for regulators to determine what types of MPs need to be prioritised in monitoring programmes and where controls should be implemented. Nevertheless, similar to other anthropogenic contaminants, 80% of MPs are considered to originate from land-based sources (Rochman et al. 2015). Therefore the role of wastewater treatment plants (WWTPs) as potential barriers of MP pollution should be considered, as they are important links between the anthropogenic and natural environments (Ou & Zeng 2018).

Wastewater treatment systems are designed to remove contaminants from household and trade effluent, so their role in MPs removal has been generating increasing attention, yet they remain largely unexplored (**Table 1**). The majority of available studies quantify MPs in secondary effluent, with fewer studies considering tertiary treatment plants (**Table 1**). Here, secondary treatment refers to biological wastewater treatment (e.g. activated sludge) resulting in the separation of decanted effluent and sludge containing microbial biomass (European Environment Agency 2019). Tertiary or advanced treatment refers to post-secondary polishing steps (e.g. chemical removal, advanced filtration) to eliminate pollutants not removed by secondary treatment (European Environment Agency 2019).

Current understanding suggests that a mixture of primary and secondary MPs may be entering the treatment facilities daily, at varying levels of pollution (Sun et al. 2019). Microplastic concentrations in raw wastewater are reported so far to range from <1 particle L-1 as observed by multiple studies (**Table 1**), to 18,285 particles L-1 reported in a secondary treatment site in Denmark (Simon et al. 2018). Conversely, effluent concentrations between 8 x 10-4 (Magnusson and Noren. 2014) and 447 (Simon et al. 2018) particles L-1 have been observed in secondary WWTPs, and between 0 (Carr et al. 2016) and 51 particles L-1 (membrane bioreactor, MBR; Leslie et al. 2017) after advanced treatment (Sun et al. 2019), with larger facilities likely discharging higher loads (Mason et al. 2018). While the WWTP literature has grown over the past two years, each study differs in methodologies (e.g. sampling volumes, detection limits), plant capacity, and type of treatment technologies and stages examined. Therefore, it is difficult to determine what variation across studies is due to site differences or analytical bias, limiting comparability of findings and comprehensive understanding of the occurrence and fate of MPs in these systems.

Comparison of influent vs effluent concentrations is a common approach to estimate removal efficiencies, which range between 40% and 99.9% (**Table 1**). While absolute values may be difficult to compare, reporting of removal percentages may improve intra-study comparisons, but not all studies report this. Despite high retention efficiencies, low concentrations in final or treated effluent may represent daily releases of millions of MPs when scaled for the discharge volumes (Mason et al. 2016; Murphy et al. 2016). For instance, concentrations of 2.5×10^{-1} and 4×10^{-3} particles L⁻¹ in final effluent, equated to discharges of 6.5×10^{7} and 5×10^{4} MPs day⁻¹, respectively in secondary treatment plants in Scotland, UK (Murphy et al. 2016) and San Francisco, USA (Mason et al. 2016). Microplastic discharges from WWTPs appear highly variable, and treatment procedure employed at the facility is presumed to be crucial in their retention.

The role of different treatment processes in removing contaminants from these systems can be assessed by a stage-wise inspection of MPs abundances during their passage through a single facility. Owing to challenges of sample collection and processing times, only a few studies have done this (**Table 1**), and stages sampled vary across studies. It appears that between ~63 and 98% of the removal can occur by the primary stage (Sun et al. 2019). Secondary treatment may reduce an additional 7 to 20% of MPs not captured by preliminary and primary treatment (Talvitie et al. 2017b; Ziahjaromi et al. 2017; Gies et al. 2018). The observation of MPs in different types of biosolids suggest that their removal during earlier stages is through their capture in various sludge fractions including grit and grease skimmings (Murphy et al. 2016), sewage sludge (Bayo et al. 2016; Murphy et al. 2016; Leslie et al. 2017; Mintenig et al. 2017; Li et al. 2018), and returned activated or excess sludge (Carr et al., 2016; Talvitie et al. 2017a; Lares et al. 2018).

While the nature of primary and secondary treatment is mostly consistent across studies, there is an array of advanced treatment techniques. Studies comparing MPs in tertiary vs. secondary effluent found that different advanced treatment technologies can further decrease MPs before discharge (Michielssen et al. 2016; Mintenig et al. 2017; Talvitie et al. 2017a,b; Ziahjaromi et al. 2017; Lares et al. 2018; Magni et al. 2019). Overall, MBR (Lares et al. 2018; Talvitie et al. 2017a) and advanced filtration technologies (Michielssen et al. 2016; Mintenig et al. 2017; Talvitie et al. 2017 a,b; Ziahjaromi et al. 2017; Magni et al. 2019) have been reported as effective means in reducing MPs from final effluent. Dissolved air flotation in Finland (Talvitie et al. 2017a) and reverse osmosis and decarbonation in Australia (Ziahjaromi et al. 2017) also showed high performance. However, in other studies, advanced treatment by gravity sand filtration (Carr et al. 2016) and MBR (Leslie et al. 2017) did not promote further reduction in particle concentrations. These different findings in advanced

WWTP studies support the need for further research on a range of treatment technologies to produce a representative assessment of their role in removing MPs from wastewater. This information could help identify control points within these systems, and what development or modification of operational procedures may decrease MPs discharge to the recipient waters.

Further research of WWTPs is crucial in MPs research because wastewater is a complex and heterogeneous matrix, and pollution levels and removal efficiencies appear to exhibit high inter- and intra-site variability (Mason et al. 2017). Especially, empirical data are needed for multiple stages other than final effluent and to explore factors driving spatio-temporal variabilities. Here, a study was conducted in a WWTP in the UK (Scotland) to: (1) understand the inflow and outflow loading of MPs (quantity and composition) in a tertiary treatment plant, accommodating temporal variability, and (2) assess the stepwise effect of treatment stage on the distribution and fate of MPs sized between 60-2800 µm. To our knowledge, this is the first study to evaluate MPs in advanced treatment systems in the UK by long-term (i.e. 10 months) spatial sampling in a single facility.

2 Materials and methods

2.1 Study site and sampling

The study site was a tertiary wastewater treatment plant in Scotland, UK, with 184,500 population equivalents (p.e.) and receiving a mix of trade and domestic sewage. The plant consists of preliminary treatment of wastewater by coarse screening (12 mm) and grit removal, primary settling tanks (phases 1 and 2), activated sludge treatment and clarification in final settling tanks (phases 1 and 2), and nitrification on plastic media trickling filters (**Fig**

1), with final discharge of treated effluent into a freshwater river. Phases 1 and 2 were created due to an expansion of the treatment plant. This splits the stream into parallel channels for primary and secondary stages but there is no difference in treatment between the two.

Sampling was conducted five times between May 2017 and February 2018: 19 May 2017 (sampling event, SE1), 13 July 2017 (SE2), 20 October 2017 (SE3), 11 January 2018 (SE4), and 16 February 2018 (SE5). The flow range covered by the sampling events was 111,496 to 184,703 m³ day¹, representing low to medium flow (Qmean = 166,422 m³ day¹; Fig S1). During each sampling event, a 5-L wastewater sample was collected from each of eight sample collection points (P): influent before screens (P1), preliminary effluent after coarse screening and grit removal (P2), primary effluent phase 1 (P3a) and phase 2 (P3b), secondary effluent after tertiary treatment (P6) (Fig 1). Samples were collected in the morning, with two additional afternoon samples on the same day during SE5 from the influent (P1, pm) and effluent (P6, pm), to explore daily fluctuations. A bulk sample, taken by lowering a metal bucket into the stream, was filtered through a 2.8 mm metal sieve, and collected in plastic bottles for transport to the laboratory. Bottles were kept in black plastic bags at 3°C until processing within a maximum of 8 weeks after collection.

2.2 MP extraction

The methodology for extraction and characterisation is broadly adapted from wet peroxide oxidation (WPO) protocols (Nuelle et al. 2014). As sewage can contain pathogens, all samples were processed in a Category 2 biological safety cabinet (Cat 2 BSC) and room, which also helped minimise potential background contamination of samples. Samples were transferred to glass Erlenmeyer flasks and spiked with 50 standard polyethylene (PE) beads

each (0.71-0.85 mm diameter, ρ =0.96 g cm⁻³; Cospheric LLC, Santa Barbara, California), to determine recovery rates. The spiked samples were treated with 30% hydrogen peroxide (H₂O₂; 1:1, v/v) for digestion of labile organics, heated in a water bath to 75°C for 30 minutes to accelerate the reaction, stirred using a magnetic stirrer for 10 minutes, and digested at room temperature for three days. After the digestion period, samples were treated with UV light for 30 minutes to ensure they were sufficiently sterile to be removed from the Cat 2 BSC room for filtration under vacuum through Whatman 1.2- μ m glass fibre filters (47 mm diameter). This processing stage was very time-consuming, indeed samples still contained some level of suspended solids and therefore filtration of 5-L samples was slow and required several filters. It was the step that limited the volume of samples that could be processed between sampling events. However, the entire sample was processed and filtered in this fashion to minimise the potential loss of smaller MPs by on-site filtration.

2.3 MP characterisation

Particle characterisation followed a two-step process starting with visual sorting of suspected MPs into four categories based on morphology: pellets, fibres, fragments, and films. Each entire filter area was examined using a Leica MX₇₅ stereo microscope with magnification between 10x and 32x to identify and quantify particles of size range between 60 and 2800 μm (Blair et al. 2019).

A subsample of 70 pieces, equivalent to 5% of total particles identified during visual inspection, was selected for chemical confirmation of plastics by Fourier-transform infrared-attenuated total reflectance spectroscopy (FTIR-ATR), using a Shimadzu IRAffinity-1S FTIR with diamond crystal and 20 scans. Manipulation of small particles was difficult, thus chemical analysis was only possible for fibres (n=19), fragments (n=10) and films (n=41)

larger than 300 µm. Pellets could not be analysed as they were lost during transfer due to their small sizes and smooth surfaces. Materials were identified by comparing the unknown spectra to those in the Shimadzu LabSolutions IR libraries, which contain approximately 12,000 reference spectra. For each particle, the top three automated matches were compared visually to assess closeness of match, and except for four pieces, the highest score was considered acceptable and reported (**Table S3**). The counts for confirmed plastics were used to estimate percentages for each category, subsequently extrapolated to correct all visual counts, including the 60-300 µm fraction. Further details of the FTIR-ATR characterisation process are in the **Supplementary Material**.

2.4 Quality control

A procedural blank was created for each SE by running 5 L of DI water through the same sample equipment used to collect samples, and processed the same way as wastewater. The purpose of the procedural blanks was to evaluate possible cross-contamination from generation of particles from plastic equipment used during sampling – these include plastic bottles, synthetic ropes, and a plastic funnel. Laboratory blanks were created in triplicates by placing 1 L of DI water in the same glass containers used for sample processing and leaving uncovered on lab benches during the extraction process, and filtering in parallel with each run of field samples. The purpose of the lab blanks was to capture cross-contamination from deposition of airborne particles in the general environment. Procedural and lab blanks, respectively, contained 4-14 and 0-3 coloured fibres by count (**Supplementary Material**), while no other type of particles were observed. It was not possible to analyse fibres in the blanks chemically, but their presence is considered evidence of cross-contamination from the environment and the use of synthetic sampling ropes.

Fragmentation tests using MP-spiked DI water were carried out to assess if the extraction process could generate secondary MPs at various stages. This is reported in the **Supplementary Material**. It was found fragmentation could occur, but the MPs used to assess this (microbeads) were rare in the samples, and so this understanding could not be used to refine MP estimates.

2.5 MP estimation

For each category, visual counts were corrected by subtracting the corresponding procedural blank. To ensure MPs were quantified correctly, blank-correct data were multiplied by the percentage of FTIR-confirmed plastics in each category. Such FTIR correction was employed for conservative estimates of daily discharge from a secondary WWTP in Vancouver, although blank correction was not incorporated in their calculation (Gies et al. 2018). The FTIR-corrected counts were summed to estimate total MP abundance (items L⁻¹), for each stage and each sampling campaign. Daily flow data for the WWTP were used to estimate incoming and outgoing MP loads in items day⁻¹ and stage-wise removal efficiencies.

3 Results and discussion

3.1 Chemical confirmation of MPs

During visual characterisation, a total of 1308 items across all samples were considered potential MPs: 871 fibres, 191 fragments, 239 films, and 7 pellets (n=7) (**Fig 2**). Chemical characterisation confirmed that MPs were present and comprised 39% of the total pieces measured by FTIR-ATR (**Fig 3**). Within each category of suspected MPs, plastics comprised

63%, 80%, and 17% of fibres, fragments, and films respectively. In absence of chemical confirmation and thus based on appearance, all micropellets (the lowest abundance of particle) recovered from wastewater samples were counted as primary MPs. Thus, based on FTIR-corrected data, a total of 749 MPs were observed across all wastewater samples, consisting of 549 fibres, 153 fragments, 41 films, and 7 pellets.

Different types of polymers identified (**Fig 3**) included commonly-used plastics like polypropylene (PP, 23%) and PE (4%), and some less common, such as polyvinyl stearate (PVS, 7%) and polyoxymethylene (POM, 1%). The remaining MPs identified here were grouped as copolymers and included an ethylene-ethyl acrylate film and a PE-PP fragment. Polypropylene and PE are often reported in relatively high abundances across available surveys (Sun et al. 2019), as they are used in a wide number of applications including personal care and packaging products. The second-most detected polymer was PVS, a material not yet reported in other studies to date, and of limited use in the plastics industry (Gooch 2011). Polyvinyl stearate can be co-polymerised with polyvinyl chloride, PVC (Gooch 2011) so may indicate construction applications. The POM particles also may not be common, only reported to date from a Danish secondary WWTP. The same study found PE-PP copolymers in raw and treated wastewater (Simon et al. 2018), but in higher abundance than this study.

Non-plastic materials were also present in the subsample (**Fig 3**): cellulose (36%), lecithin (13%), and protein (1%). While these are not the focus of this paper, their presence should still be noted as depending on sample purification process, they may not be entirely removed from samples and thus mistaken as MPs. The remaining pieces classed as "Other" included 5 fibres, 2 fragments, and 1 film. These particles could not be identified as they showed no

distinguishable peaks to allow for manual annotation or to produce any hits during the library search (Fig S3).

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3.2 MP morphology

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Secondary MPs were predominant in the wastewater samples, comprising 99.5% of total pieces. Fibres were the most common type of MPs, followed by fragments and films. The predominance of fibres here is consistent with previous wastewater surveys (e.g. Sutton et al. 2016; Gies et al. 2018; Lares et al. 2018; Conley et al. 2019). Fibre abundance is expected to be higher in densely-populated areas as they can be carried by washing machine effluent. For example, clothes washing can release between 1.9 x 10³ (Browne et al 2011) and 6 x 10⁶ fibres per wash (De Falco et al. 2018). The highest releases have been observed from polyester (Pest) and polyamide (PA) garments, but these materials were not identified by FTIR-ATR here. This may be as Pest and PA fibres were settling out of suspension due to higher densities. Therefore, their concentrations in the liquid fractions would be lower than the detection limit allowed by a 5-L sample. Alternatively, they may have been smaller than 300 µm and thus were not subsampled for chemical identification. However, PP fibres may highlight the importance of other sources like sanitary products, thermal clothing, medical applications, and construction materials (Mandal 2019), but the discussion on these alternative sources of fibres to WWTPs is limited in the literature. Fibre count was highlyvariable across sampling events, and while generally decreased after each treatment stage (Fig 4), some fibres persisted through the process and were observed in final effluent.

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Fragments were present throughout all treatment stages and at least one particle was observed in final effluent (**Fig 4**). Most fragment removal seemed to occur after the primary stage (when settling of solids takes place) and again after tertiary treatment. Films were

mostly removed during pre-treatment, which may indicate they are more likely to be captured in the grit and grease biosolids as observed in a similar study in a Scottish secondary WWTP (Murphy et al. 2016). Different types of fragmented pieces have also been observed across multiple WWTPs (Sun et al. 2019) and generally refer to uneven or irregular pieces. As observed here, fragments were the second most-abundant MPs after fibres in a Swedish secondary WWTP (Magnusson and Noren 2014), in secondary and tertiary WWTPs in the USA (Mason et al. 2016; Sutton et al. 2016), and in an Italian tertiary treatment plant (Magni et al. 2019). Here, fragmented pieces were categorised as either films or fragments to distinguish between two-dimensional thin particles and three-dimensional pieces with broken edges, respectively. However, the terms used to categorise these particles may vary across surveys (Hidalgo-Ruz et al. 2012), thus it is necessary to unify classifications for adequate consideration.

Fragments can be produced from a wide variety of sources and enter the wastewater stream via household and industrial effluent, but fragments generated during the treatment process cannot be excluded, supported by evidence of fragmentation of larger MPs beads (>700 µm) in controlled tests here. This needs to be validated for other particle types and sizes. Furthermore, the WWTP may have plastic equipment that if degrades over time could release MPs, but to our knowledge this has not been explored. The mechanical generation of MP fragments, particularly in sizes that may be evading detection, presents an important research gap in these systems that warrants further investigation as without it WWTP loading and MP redistribution cannot be fully understood.

Lastly, microbeads were only observed before secondary treatment (**Fig 4**). This is consistent with previous observations in Swedish secondary WWTPs where 95-99% of microbeads were considered to settle out in sludge (Magnusson and Noren, 2014), and in the UK where

microbeads were only found in grease fractions removed during pre-treatment (Murphy et al. 2016). These observations are for particles >65 µm. Therefore, entrapment in sludge may explain why these particles were only observed in the early treatment stages in this study also. Primary MPs (i.e. microbeads) can be introduced to WWTPs via household sewage, but primary MPs represent only a small portion of the plastic load in this catchment. This discussion is relevant to current considerations on MP control measures of MPs, especially as current actions such as regulatory bans are mainly aimed at reducing primary MPs inputs, and few focus on secondary sources.

3.3 MP abundances

Microplastics were present throughout the system. Concentrations ranged from ~1 to 13 MPs L⁻¹, with highest abundances in pre-treatment effluent during SE1 (**Fig 5**). Total concentrations of MPs were highly-variable across sampling dates and time, consistent with other reports of high variability (Sun et al. 2019). Influent concentrations were between 3 and 10 MPs L⁻¹, with maximum abundances observed in January and minimum in February and July. In effluent, concentrations were between <1 and 3 MPs L⁻¹. The lowest concentrations were mostly observed after tertiary treatment (final effluent), except during SE2, when concentrations reached their minimum after the mixed secondary liquor. Both influent and effluent abundances observed here are comparable to those in a secondary WWTP in Glasgow, Scotland (Murphy et al. 2016) but considerably lower than in three secondary WWTPs in South Carolina, USA (Conley et al. 2019). Nevertheless, current methods may not be suited to detect small MPs (e.g. <300 μm) so it is probable that MP concentrations are underestimated, especially as small MPs have been observed in greater abundances than larger pieces (Carr et al. 2016; Mintenig et al. 2017). Moreover, small MPs (e.g. 20-190 μm) may be more common in final effluent as they are more likely to pass

through filtration barriers if not retained in biosolid fractions and smaller than the pore size (Ziajahromi et al. 2017; Sun et al. 2019).

Abundances were highly variable across sampling events and between the morning and afternoon samples collected on the same day, despite similar flow conditions. A survey of three USA WWTPs observed concentrations to vary by a factor of 2.5 and 4.8 in influent and effluent respectively, and long-term variations were greater than in short-term (Conley et al. 2019). However, the absence of replicates in the present study limited this assessment of short-term variation, and future work should explore this to support considerations of regulating inflow concentrations of different types of MPs to the system.

3.4 MP removal and loadings

Average MP inflow to the treatment plant over one year was 8.1×10^8 , 95% CI [3.8×10^8 , 1.2×10^9] particles day⁻¹. Influent loads based on incoming concentrations and plant flows are only reported by a few studies (Magnusson and Noren 2014; Murphy et al. 2016; Lares et al. 2018; Conley et al. 2019), but their findings suggest these loads may be partially dependent on the size of population served. For example, among three WWTPs in South Carolina, a WWTP serving 1.8×10^5 p.e. received considerably higher MP loading than a treatment plant serving a smaller population (Conley et al. 2019). In an earlier survey in the same catchment of this study in Scotland, a larger secondary treatment plant serving 6.5×10^5 p.e. received an average daily load of 4×10^9 MPs >65 μ m. Incoming loads in the present study were mostly comparable to those of a Finnish secondary treatment plant (p.e. not specified) in Finland with a reported daily inflow of 6.2×10^8 MPs >0.25 μ m (Lares et al. 2018).

Particles concentration decreased between influent and final outflow with each treatment stage removing different proportions of MPs (Fig 6). Mean concentrations decreased by 6% (standard error 16) after pre-treatment. Preliminary treatment has only been assessed by two studies, and removal efficiencies in this research are lower than those reported, ~35-58% (Michielssen et al. 2016; Murphy et al. 2016). Primary treatment removed between 60 (P3a, standard error 10) and 76% (P3b, standard error 6) of overall MP counts and is consistent with other surveys (63-81%, Dris et al. 2016; 84-88%, Michielssen et al. 2016; 78%, Murphy et al. 2016; 97.4-98.4%, Talvitie et al. 2017b; ~68%, Ziahjaromi et al. 2017). There was indication of further removal after secondary treatment, but this was only evident at the secondary mixed liquor stage after the channels are joined back together (P5). As there is no remediation between P4 and P5 stages, this reduction suggests that engineering parameters and infrastructure may play a role in MP retention, especially if a large portion of removal is attributed to settling. After secondary treatment (P5), removal reached 92% (standard error 3), comparable to a Finnish secondary treatment plant where 7-20% of MPs were removed by activated sludge treatment (Talvitie et al. 2017b). A similar study in a larger UK secondary treatment plant had a retention efficiency of 98% and discharged 6.5 x 10⁷ particles day⁻¹ (Murphy et al. 2016). Although the data come from different WWTPs, both studies are located in the same catchment, serve a similar population demographic, and observed a similar profile of MPs. Therefore, the differences between the two plants emphasise that removal of MPs will depend on site-specific engineering parameters besides loading and general treatment process.

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Tertiary treatment produced an average 4% (standard error 1) decrease in MPs in secondary effluent, bringing the total retention efficiency to ~96% (**Fig 6**). The plant discharges on average 2.2 x 10⁷, 95% CI [1.2 x 10⁷, 3.2 x 10⁷] MPs day⁻¹ under low- to medium-flow conditions. The removal ranges and discharges here are within those observed elsewhere

(Table 1), noting cross study comparisons are difficult as different sampling volumes and size ranges can introduce uncertainty to MP measurements reported across sites. No other sites of the same type of treatment considered here (i.e. use of plastic media in nitrifying trickling filters) have been documented, but removal percentages in this WWTP were higher than those observed by advanced sand filters (Magni et al. 2019) and lower than MBR (Michielssen et al. 2016; Talvitie et al. 2017b). The differences among these treatment technologies may be expected because of differences in the porosity of the filters they use, and so may indicate a way in which performance of tertiary treatment may be predicted. Nevertheless, the diversity of advanced systems and the contrasting results reported for different facilities, mean more research in WWTPs is needed to help identify which technologies optimise removal of MPs pollution in and from these systems.

4 Conclusions

Here, the occurrence, distribution, and fate of MPs in an advanced WWTP were assessed. A continuous input of MPs and other microdebris to the treatment site was observed over the course of ten months. The presence of MPs was confirmed by FTIR-ATR analysis, with PP identified as the most abundant type and present as fibres and fragments. Microplastics were mainly observed as secondary types, and while a few pellets were present, their chemical composition could not be determined due to size limitations of the FTIR-ATR approach employed here. Fibres were dominant. Their high abundance is expected as they are often associated with washing machine effluent, but their presence in blanks suggests that some may be entering the system via atmospheric, possible as the wastewater is treated in open channels. The system investigated here had apparent removal efficiencies at the higher end of that observed elsewhere, but MPs were not entirely removed and at least 1.2 x 10⁷ particles may be discharged daily from this site even during low flow. These estimates are limited to

particles sized 60-2800 µm but there will be smaller MPs in the system that need to be investigated further. As observed by other studies, the largest concentration reduction was observed in early treatment stages. Generally, this is linked to retention of microplastics in the sludge and so the concentration and fate of MPs in sludge needs further attention because rather than providing a solution, it may be displacing delivery of MPs to the environment. This research generates new understanding of MPs in WWTPs by its consideration of multiple stages, including tertiary treatment, not yet considered elsewhere and by employing a longer sampling period in a single facility to generate spatio-temporal understanding. Further research could use larger sample volumes to reduce the blank sensitivity and incorporate greater sampling frequency to assess short-term variation and thus contextualise seasonal observations. As wastewater treatment plants are expected to play an increasingly important role in regulating the delivery of MPs coming from land-based sources, this and similar studies can help to inform regulators about what needs to be prioritized in monitoring programmes and where controls should be implemented, thus guiding fundamental action.

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References

408	Andumani S, Kakkai P. 2018. Ecoloxicological effects of inicropiastics on biota. a review.
469	Environ Sci Pollut Res 25: 14373-14396
470	
471	Bayo J, Olmos S, López-Castellanos J, Alcolea A (2016) Microplastics and microfibers in
472	the sludge of a municipal wastewater treatment plant. Int J Sus Dev Plann 11: 812-
473	821
474	
475	
476	Blair RM, Waldron S, Phoenix V, Gauchotte-Lindsay (2019) Microscopy and elemental
477	analysis characterisation of microplastics in sediment of a freshwater urban river in
478	Scotland, UK. Environ Sci Pollut Res 26: 12491-12504
479	
480	Browne MA, Crump P, Niven SJ, Teuten E, Tonkin A, Galloway T, Thompson R. 2011.
481	Accumulation of microplastic on shorelines worldwide: sources and sinks. Environ
482	Sci Technol 45: 9175-9179
483	
484	Carr SA, Liu J, Tesoro AG (2016) Transport and fate of microplastic particles in wastewater
485	treatment plants. Water Res 91: 174-182
486	
487	Conley K, Clum A, Deepe J, Lane H, Beckingham B. 2019 Wastewater treatment plants as
488	a source of microplastics to an urban estuary: Removal efficiencies and loading per
489	capita over one year. Water Res X 3: 100030
490	
491	De Falco F, Gullo MP, Gentile G, Di Pace E, Cocca M, Gelabert L, Brouta-Agn'esa M,
492	Rovira A, Escudero R, Villalba R, Mossotti R, Montarsolo A, Gavignano S, Tonin

493	C, Avella M. 2018 Evaluation of microplastic release caused by textile washing
494	processes of synthetic fabrics. Environ Pollut 236: 916-925
495	
496	Dris R, Gasperi J, Rocher V, Saad M, Renault N, Tassin B (2015) Microplastic
497	contamination in an urban area: a case study in Greater Paris. Environ Chem 12: 592-
498	599
499	
500	Dyachenko A, Mitchell J, Arsem N (2017) Extraction and identification of microplastic
501	particles from secondary wastewater treatment plant (WWTP) effluent. Analysis
502	Methods 9: 1412-1481
503	
504	European Environment Agency (2019) EEA Glossary. Available at
505	http://glossary.eea.europa.eu/EEAGlossary (accessed 18 July 2019)
506	
507	Gies EA, LeNoble JL, Noel M, Etemadifar A, Bishay F, Hall ER, Ross PS (2018) Retention
508	of microplastics in a major secondary wastewater treatment plant in Vancouver,
509	Canada. Mar Pollut Bull 133: 553-561
510	
511	Gooch JW (2011) Polyvinyl Stearate. In: Gooch J.W. (eds) Encyclopedic Dictionary of
512	Polymers. Springer, New York, NY
513	
514	Hidalgo-Ruz V, Gutow L, Thompson RC, Thiel M (2012) Microplastics in the marine
515	environment: a review of the methods used for identification and quantification.
516	Environ Sci Technol 46: 3060-3075
517	

518	Lares M, Ncibi MC, Sillanpaa M, Sillanpaa M (2018) Occurrence, identification and
519	removal of microplastic particles and fibers in conventional activated sludge process
520	and advanced MBR technology. Water Res 133: 236-246
521	
522	Leslie HA, Brandsma SH, van Velzen MJM, Vethaak AD (2017) Microplastics en route:
523	Field measurements in the Dutch river delta and Amsterdam canals, wastewater
524	treatment plants, North Sea sediments and biota. Environ Int 101: 133-142
525	
526	Li X, Chen L, Mei Q, Dong B, Dai X, Ding G (2018) Microplastics in sewage sludge from
527	the wastewater treatment plants in China. Water Res 142: 75-85
528	
529	Magni S, Binelli A, Pittura L, Avio CG, della Torre C, Parenti CC, Gorbi S, Regoli F (2019)
530	The fate of microplastics in an italian wastewater treatment plant. Sci Total Environ
531	652: 602-610
532	
533	Magnusson K, Norén F (2014) Screening of Microplastic Particles in and down-Stream a
534	Wastewater Treatment Plant, Report C55. Swedish Environmental Research
535	Institute: Stockholm
536	
537	Mandal J (2019) Polypropylene fiber and its manufacturing process, properties, advantages,
538	disadvantages and applications of polypropylene fiber. Textile Learner. Available at
539	https://textilelearner.blogspot.com/2013/01/polypropylene-fiber-and-its.html
540	(accessed 18 June 2019)
541	

542	Mason SA, Garneau D, Sutton R, Chu Y, Ehmann K, Barnes J, Fink P, Papazissimos D,
543	Rogers DL (2016) Microplastic pollution is widely detected in US municipal
544	wastewater treatment plant effluent. Environ Pollut 218: 1045-1054
545	
546	Michielssen MR, Michielssen ER, Ni J, Duhaime MB (2016) Fate of microplastics and other
547	small anthropogenic litter (SAL) in wastewater treatment plants depends on unit
548	processes employed. Environ Sci Water Res Technol 2: 1064-1073
549	
550	Mintenig SM, Int-Veen I, Loder MGJ, Primpke S, Gerdts G (2017) Identification of
551	micriokastic in effluents of waste water treatment plants using focal plane array-
552	based micro-Fourier-transform infrared imaging. Water Res 108: 365-372
553	
554	Murphy F, Ewins C, Carbonnier F, Quinn B (2016) Wastewater treatment works (WwTW)
555	as a source of microplastics in the aquatic environment. Environ Sci Technol 50:
556	5800-5808
557	
558	Nuelle MT, Dekiff JH, Remy D, Fries E (2014) A new analytical approach for monitoring
559	microplastics in marine sediments. Environ Pollut 184:161-169
560	
561	Ou H, Zeng EY (2018) Occurrence and fate of microplastics in wastewater treatment plants.
562	In: Zeng EY (eds) Microplastic Contamination in Aquatic Environments: An
563	Emerging Matter of Environmental Urgency. Elsevier, pp 317-338
564	
565	Rochman CM, Kross SM, Armstrong JB, Bogan MT, Darling ES, Green SJ, Smyth AR,
566	Veríssimo D (2015) Scientific evidence supports a ban on microbeads. Env Sci
567	Technol 49: 10759-10761

568	
569	Simon M, van Alst N, Vollertsen J (2018) Quantification of microplastic mass and removal
570	rates at wastewater treatment plants applying Focal Plane Array (FPA)-based Fourier
571	Transform Infrared (FT-IR) imaging. Water Res 142: 1-9
572	
573	Sun J, Dai X, Wang Q, van Loosdrecht MCM, Ni B-J (2019) Microplastics in wastewater
574	treatment plants: Detection, occurrence and removal. Water Res 152: 21-37
575	
576	Sutton R, Mason SA, Stanek SK, Willis-Norton E, Wren IF, Box C (2016) Microplastic
577	contamination in the San Francisco Bay, California, USA. Mar Pollut Bull 109: 230-
578	235
579	
580	Talvitie, J, Mikola A, Koistinen A, Setala O (2017a) Solutions to microplastic pollution -
581	removal of microplastics from wastewater effluent with advanced wastewater
582	treatment technologies. Water Res 123: 401-407
583	
584	Talvitie J, Mikola A, Setala O, Heinonen M, Koistinen A (2017b) How well is microliter
585	purified from wastewater? - A detailed study on the stepwise removal of microliter
586	in a tertiary level wastewater treatment plant. Water Res 109: 164-172
587	
588	Ziajahromi S, Neale PA, Rintoul L, Leusch FDL (2017) Wastewater treatment plants as
589	pathways for microplastics: development of a new approach to sample wastewater-
590	based microplastics. Water Research: 112: 93-99
591	