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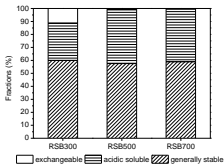
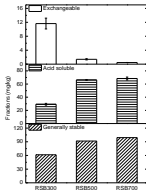
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*Graphical Abstract

**Lead on high temperature rice straw biochars
is generally more stable**



- Higher temperature biochars remove Pb faster and have higher removal capacity
- Precipitation was a mechanism by which all three biochars immobilized Pb
- RSB300 has 11.34% of total immobilized Pb attributed to the exchangeable fraction
- Exchangeable Pb was less than 1% on RSB500 and RSB700
- High temperature rice straw biochars may be more suitable for soil remediation

1 **Effect of production temperature on lead removal mechanisms by rice straw biochars**

2 Zhengtao Shen ^{1,2,3}, Deyi Hou ^{1,2,*}, Fei Jin ⁴, Junxian Shi ⁵, Xiaoliang Fan ⁵, Daniel

3 C.W. Tsang ⁶, Daniel S. Alessi ³

4 1 National Engineering Laboratory for Site Remediation Technologies, Beijing 100015,

5 China

6 2 School of Environment, Tsinghua University, Beijing 100084, China

7 3 Department of Earth and Atmospheric Sciences, University of Alberta, Edmonton

8 T6G 2E3, Canada

9 4 School of Engineering, University of Glasgow, Glasgow G12 8QQ, UK

10 5 School of Ecology and Environment, Inner Mongolia University, Hohhot 010021,

11 China

12 6 Department of Civil and Environmental Engineering, Hong Kong Polytechnic

13 University, Hung Hom, Kowloon, Hong Kong, China

14 *Corresponding author: Deyi Hou, E-mail: houdeyi@tsinghua.edu.cn

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Abstract: Production temperature significantly affects biochar properties and consequently the removal mechanisms of heavy metals. In this study, rice straw biochars were produced at 300, 500 and 700 °C (RSB300, RSB500 and RSB700). The influence of production temperature on the adsorption characteristics and removal mechanisms of lead on this set of rice straw biochars were investigated by batch adsorption tests, micro-structural analyses and sequential metal extractions. Biochars produced at higher temperatures had significantly higher pH values and surface areas, resulting in higher metal removal capacities and faster uptake kinetics. Precipitation was a key mechanism for lead removal from solution for all biochars: lead oxalate was precipitated on RSB300, and hydrocerussite was precipitated on RSB500 and RSB700. The immobilized lead fraction on the biochars could be divided into exchangeable, acid soluble and non-available fractions. RSB300 had 11.34% of the total immobilized Pb attributed to the exchangeable fraction, whereas for RSB500 and RSB700, it was less than 1%. Immobilized Pb on RSB500 and RSB700 was almost exclusively attributable to the acid soluble and non-available fractions (>99%). Based on our results, RSB500 and RSB700 are likely much more appropriate for soil remediation of Pb as compared with RSB300.

Keywords: soil remediation, biochar, lead immobilization, adsorption, precipitation, production temperature

40 1 Introduction

41 Soil contamination is becoming an increasing threat to human health in China.
42 Polluted agricultural land (19% of total sampling points in China's national soil survey)
43 may produce crops with elevated heavy metals concentrations, which may enter into
44 the human body through ingestion and ultimately damage organ systems (Zhao et al.,
45 2015; Hou and Li, 2017). For these reasons, there is an urgent need to seek effective
46 and sustainable technologies to remediate contaminated agricultural land (Hou and
47 Al-Tabbaa, 2014). Biochar is regarded as a promising material that can reduce the
48 bioavailability/mobility of heavy metals after its addition into contaminated soils (Sohi,
49 2012). In addition, biochar amendment can promote soil fertility and enhance crop
50 yields, which promotes its use as a green and sustainable remediation material (Hou
51 et al., 2017a; Zhao et al., 2017).

52 Extensive studies have been carried out recently, to investigate the potential
53 application of biochar in remediation of heavy metal contaminated soils (O'Connor et
54 al., 2018b). Biochar reduces the bioavailability/mobility of heavy metals in soils
55 through various mechanisms, including: physical adsorption, cation exchange,
56 electrostatic interaction, precipitation and complexation (Inyang et al., 2015). Heavy
57 metals immobilized on biochar in soils via these various mechanisms result in
58 different mobility and environmental risks. Physically adsorbed and exchangeable
59 heavy metals on biochar are readily bioavailable and pose immediate risks to the
60 environment (Shen et al., 2017). The stability of the metals immobilized on biochar
61 through electrostatic interaction and precipitation depends on environmental

conditions. For instance, they may be stable in the short term; however, if the environmental conditions change (e.g., significant reduction of soil pH), they may be desorbed/dissolved from the biochar surface and thereby pose risks to the environment (Filgueiras et al., 2002). Complexed metals on biochar are generally regarded as stable and environmentally benign (Filgueiras et al., 2002).

For the reasons discussed above, immobilization mechanisms are very important for biochar application in soil remediation/stabilization. Although some biochars show high adsorption capacities of heavy metals, if the adsorption mechanisms are mainly physical adsorption or cation exchange, these biochars may not be suitable for soil remediation as these weakly bound metals may pose elevated environmental risks in soil. On the other hand, if the immobilization is via precipitation and/or complexation, relatively higher long-term stability is expected compared to physical adsorption and cation exchange.

The mechanisms for biochar removal of heavy metals depend on biochar chemical properties, which is significantly affected by the biochar production process (e.g., production temperature). Although a range of studies have investigated the characteristics and mechanisms of heavy metal adsorption to biochar (Shen et al., 2015; Shen et al., 2017; Shen et al., 2018b), the influence of biochar production temperature on the removal mechanisms remains unclear. It is important to link biochar production temperature and removal mechanisms of heavy metals, for optimal production and application of biochar in soil remediation.

To address this knowledge gap, rice straw biochars were produced at differing temperatures and the effect of production temperature on adsorption characteristics and immobilization mechanisms of lead on the biochars was investigated. Rice straw was selected as a feedstock due to its high availability in China, as one of the most abundant agricultural wastes (Wang et al., 2017), and lead was selected as a representative heavy metal due to its high concentrations in contaminated soils in China (Zhao et al., 2015) and high health risks (especially to children) (Shen et al., 2018a) (O'Connor et al., 2018c). The hypothesis to be verified in this study is that lead retention on the rice straw biochars produced at various temperatures will have different mechanisms, and therefore understanding these differences is critical in producing appropriate biochar for soil remediation.

2 Materials and methods

2.1 Biochar production and characterization

The rice straws used in this study were obtained from Taicang, Suzhou, China. After reception, they were oven dried at 60 °C to reach constant weight. The resulting dried feedstock was then used to produce biochars at 300, 500 and 700 °C (RSB300, RSB500 and RSB700). During production, the rice straw was pyrolyzed in a furnace (under limited air) with a heating rate of 10 °C/min and held at the target terminal temperature for 1 h. After preparation, the biochars were sieved through a #100 mesh sieve (< 0.15 mm) and stored in sealed sample bags.

The pH of the biochars was determined based on the works of Alam et al. (2018 a and

b) and von Gunten et al. (2017). Briefly, a certain amount (0.1 g) of biochar was mixed with 10 mL of deionized water and shaken at 250 rpm for 24 h. After centrifugation, the pH of the supernatant was measured. The Brunauer–Emmett–Teller (BET) surface area of the biochar was determined by a Tristar II 3020 (Micromeritics) instrument. The C, H, O, N contents of biochar were tested using a CE-440 Elemental Analyzer (Perkin Elmer).

The fundamental vibrations and associated rotational-vibrational structure of the biochars were tested using a VERTEX 70v Fourier-transform infrared spectroscopy (FT-IR) spectrometer (BRUKER) between wavenumbers 4000 to 600 cm^{-1} . The crystalline phases in the biochar were examined using a D8 ADVANCE X-ray diffractometer (XRD) (Bruker). To do so, dry samples were mounted on a flat holder and examined with a Cu K α source operating at 40 kV and 40 mA, emitting radiation at a wavelength of 1.5406 Å. The scanning regions were between 2 θ values of 10-60° at a step rate of 0.1 s/step and a resolution of 0.01°/step. The surface morphology of the biochars (coated with Pt) was tested using a SU8010 Ultra-High Resolution (1.0 nm) Scanning Electron Microscope (SEM) (Hitachi, Japan).

2.2 Adsorption tests

Batch adsorption tests were carried out to reveal the characteristics of Pb adsorption on the rice straw biochars. In order to assess the adsorption kinetics, a certain amount of biochar (0.1 g) was added to 20 mL solution of 5 mM Pb(NO₃)₂ containing 0.01 M NaNO₃ (for a relatively stable solution ionic strength) in 50-mL polyethylene tubes.

The mixtures were shaken at 250 rpm for 5 min, 10 min, 20 min, 30 min, 1 h, 2 h, 3 h, 6 h, 12 h, 18 h or 24 h in a temperature-controlled shaker (SHA-CA, Kexi Instrument CO., Ltd.). After the designated shaking time, a mixture was filtered through a 0.45 µm membrane. The Pb concentration in the collected filtrate was measured by inductively coupled plasma/optical emission spectrometry (ICP-OES) (Perkin-Elmer, Optima 8300) after dilution (if necessary) and acidification. The kinetics tests confirmed that the equilibrium adsorption time for all biochars was less than 24 hours.

In order to construct a sorption isotherm for each biochar, 0.1 g of biochar was added to 20 mL solutions containing different Pb concentrations (0.1, 0.2, 0.3, 0.5, 1, 2, 3, 5, 10 or 20 mM) and 0.01 M NaNO₃. The mixture was shaken at 200 rpm for 24 h to reach equilibrium before filtration and ICP-OES testing as described above.

Pseudo first order, pseudo second order and intraparticle diffusion models are three of the most commonly used kinetics models to reveal the time-dependence of adsorption (Cui et al., 2016; Inyang et al., 2011), and were used to simulate the kinetics data:

$$q_t = q_e(1 - e^{-k_1 t}) \quad (1)$$

$$q_t = \frac{k_2 q_e^2 t}{1 + k_2 q_e t} \quad (2)$$

$$q_t = k_i t^{0.5} + C \quad (3)$$

where k_1 (h⁻¹) is the rate constant of pseudo first order adsorption, k_2 (g/mg h) is the pseudo second order rate constant, and k_i (mg/g h^{-0.5}) is the coefficient of intraparticle diffusion. In equation (2), q_e (mg/g) is the equilibrium adsorption capacity, and q_t (mg/g)

146 in each equation is the adsorbed Pb at time t.

147 Langmuir and Freundlich models are two of the most commonly used isotherm
148 models and were used to describe the equilibrium sorption data (Foo and Hameed,
149 2010):

$$150 \quad q_e = \frac{Q_{max} b C_e}{1 + b C_e} \quad (4)$$

$$151 \quad q_e = K_f C_e^{1/n} \quad (5)$$

152 where Q_{max} (mg/g) is the maximum monolayer adsorption capacity, b is the Langmuir
153 isotherm constant (L/mg), K_f (mg/g) is the Freundlich isotherm constant, n represents
154 the adsorption intensity, q_e is the adsorbed Pb (mg/g) at equilibrium and C_e (mg/L) is
155 the equilibrium Pb concentration in solution.

156 2.3 Mechanistic analysis

157 The biochar samples after Pb adsorption were tested by FT-IR and XRD to reveal the
158 mineralogical and molecular changes, using the same testing procedure described in
159 section 2.1. In order to investigate the fractions of immobilized Pb on the biochars, a
160 simplified sequential extraction test was carried out (Shen et al., 2017): (1)
161 approximately 0.1 g solid sample was extracted with 8 mL of 0.5 M $MgCl_2$ (adjust to
162 pH 7.0 using NaOH or HCl) and shaken for 20 min at room temperature; (2) another
163 0.1 g solid sample was extracted with 8 mL of 1 M NaOAc (adjusted to pH 5.0 with
164 HOAc) and shaken for 5 h at room temperature. After shaking, the mixtures were
165 centrifuged and the supernatants collected and filtered through 0.45 μm membranes,
166 and then tested for Pb concentrations by ICP-OES test after dilution (if necessary)

and acidification. The exchangeable Pb on biochar can be calculated from extraction (1); the exchangeable + acid soluble Pb on biochar can be calculated from extraction (2), and therefore the acid soluble Pb can be obtained by subtracting (1) from (2) (Shen et al., 2017). The stable Pb on biochar can be calculated by subtracting the exchangeable and acid soluble Pb from the total adsorbed Pb (obtained from the adsorption studies).

2.4 Statistical analysis

The pH, adsorption and sequential extraction tests were conducted in duplicates, and the mean and standard deviation was reported for each experiment. The micro-structural and other tests were conducted once, and so no statistical analysis appears for these measurements.

3 Results and discussion

3.1 Biochar characterization

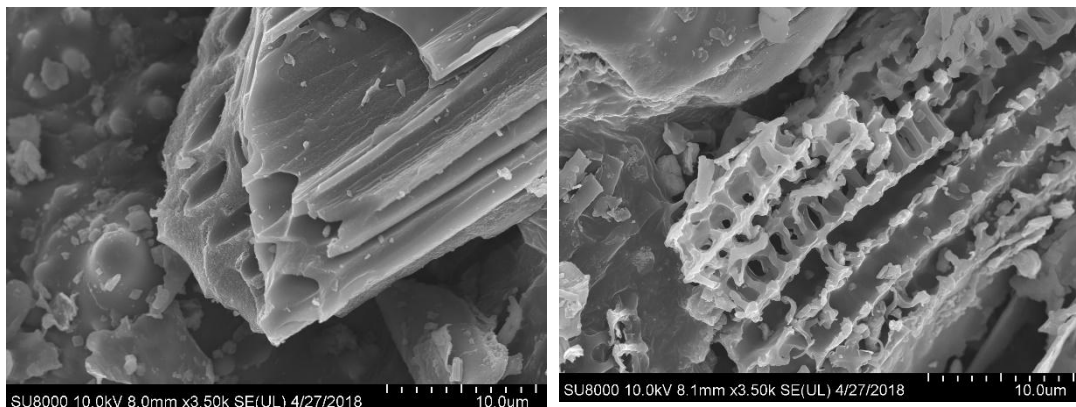
Table 1 Characteristics of the rice straw biochars

	RSB300	RSB500	RSB700
Yield	38%	31%	30%
pH	7.89 ± 0.01	10.40 ± 0.01	10.68 ± 0.01
BET surface area (m ² /g)	6.766	22.383	115.465
Average pore size (nm)	14.273	9.576	3.585

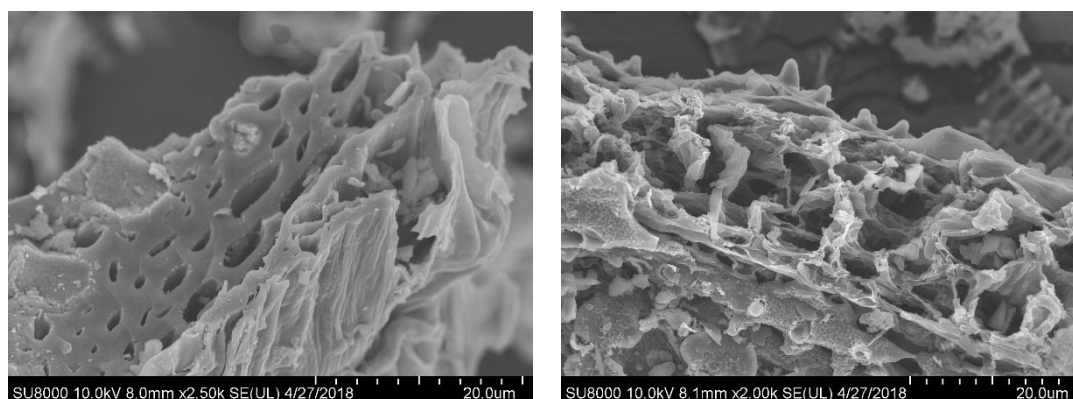
C (%)	44.66 ± 0.14	44.53 ± 0.36	44.91 ± 0.75
H (%)	3.06 ± 0.01	1.88 ± 0.01	1.24 ± 0.01
N (%)	1.62 ± 0.02	1.35 ± 0.02	1.03 ± 0.02
O (%)	19.29 ± 0.53	9.75 ± 0.25	5.91 ± 0.24
H:C	0.07	0.04	0.03
O:C	0.43	0.22	0.13

181 Biochar properties are shown in Table 1. Biochar yield decreases as production
 182 temperature increases, however the decreases diminished at high temperatures (from
 183 500 to 700 °C), because most of the volatile components had been removed at lower
 184 temperatures (Zhao et al., 2017). RSB300 is slightly alkaline with a pH of 7.89. In
 185 comparison, higher production temperature for RSB500 and RSB700 results in much
 186 higher pH values (10.40 and 10.68), because higher production temperature leads to
 187 the decomposition of acidic functional groups (e.g., carboxyl and phenol) of biochar
 188 and the formation of alkaline minerals (e.g., K₂O) (Dodson, 2011). The carbon content
 189 of the three biochars were all near 45%. In contrast, H, N, and O contents in the
 190 biochars all decreased with increasing production temperature due to the loss of the
 191 volatile component and dehydration of organic compounds (Zhao et al., 2017).
 192 Correspondingly, H:C and O:C of the biochars decreased with increasing production
 193 temperature, suggesting the surface of the biochars were more aromatic and less
 194 hydrophilic (Zhao et al., 2017).

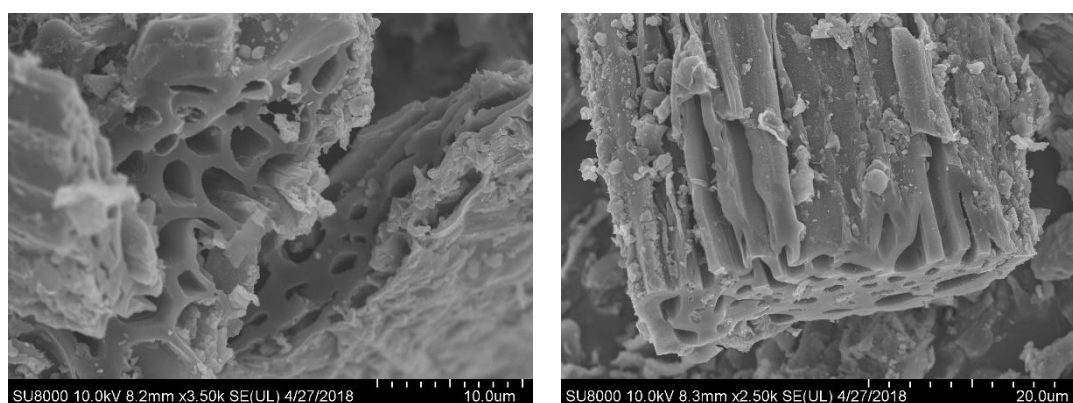
195 The BET surface area of the rice straw biochars was also significantly affected by
196 production temperature. The surface area of RSB300 ($6.766 \text{ m}^2/\text{g}$) was relatively low,
197 however, it climbs to 22.383 for RSB500 and dramatically increases to $115.465 \text{ m}^2/\text{g}$
198 for RSB700. The increased surface area suggests that pores in biochar gradually
199 developed with increased production temperature in $300\text{-}700^\circ\text{C}$, which was also
200 observed in previous studies (Keiluweit et al., 2010; Yang et al., 2015). The increase
201 in surface area from 300 to 500°C was due to the degradation and destruction of
202 cellulose, which results in the formation of amorphous carbon structure and
203 micropores (Chen et al., 2012; Zhao et al., 2017). The dramatic increase in surface
204 area from 500°C to 700°C was due to the decomposition of lignin and quick release of
205 H_2 and CH_4 , which generates significant densities of micropores (Chen et al., 2012;
206 Zhao et al., 2017). This can also be revealed by the decrease of the average pore size
207 with increased production temperature, whereby more and more macropores
208 transferred to meso and micro pores (Table 1). SEM results (Figure 1) show the
209 porous structure of the biochars at the micron scale, however, a quantitative
210 comparison of the porous structure is best confirmed by the BET results discussed
211 above.



RSB300



RSB500

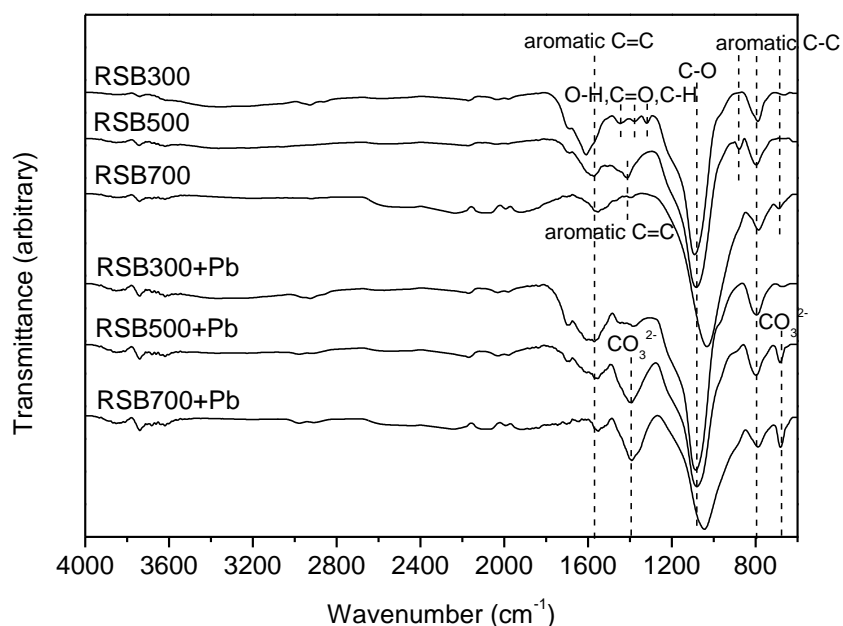


RSB700

Figure 1 Surface morphology of the rice straw biochars at different magnifications and cross sections under SEM (a)RSB300; (b)RSB500 and (c)RSB700

The FT-IR spectra of the biochars are shown in Figure 2. The peaks at ~ 1575 and $\sim 800\text{ cm}^{-1}$ that appear in each spectrum are typically found for biochars, representing aromatic carbon (Keiluweit et al., 2010). The peaks at $\sim 1080\text{ cm}^{-1}$ are attributable to the stretching of C-O bonds (Wang et al., 2017). A range of peaks (O-H, C=O and C-H) between $1440\text{--}1300\text{ cm}^{-1}$ were observed for RSB300, which originate from the cellulose, hemicellulose and lignin (Keiluweit et al., 2010). With higher production

220 temperature, these raw materials decomposed and a more active aromatic structure
 221 formed, resulting in the disappearance of the peaks between 1440-1300 cm^{-1} and the
 222 appearance of peaks representing aromatic C=C stretching (1414 cm^{-1}) for RSB500
 223 (Keiluweit et al., 2010; Wu et al., 2012). At a production temperature of 700 $^{\circ}\text{C}$, the
 224 C=C stretching peak declined due to the progressive condensation of the aromatic
 225 structure at high temperatures (Keiluweit et al., 2010; Wu et al., 2012). The XRD
 226 patterns are shown in Figure 3, and indicate that the biochars are highly amorphous
 227 and contain only KCl as the crystalline phase, which is in line with previous findings on
 228 rice straw biochars (Qian et al., 2016; Tan et al., 2017).



229
 230 Figure 2 FT-IR spectra of the rice straw biochars before and after lead adsorption

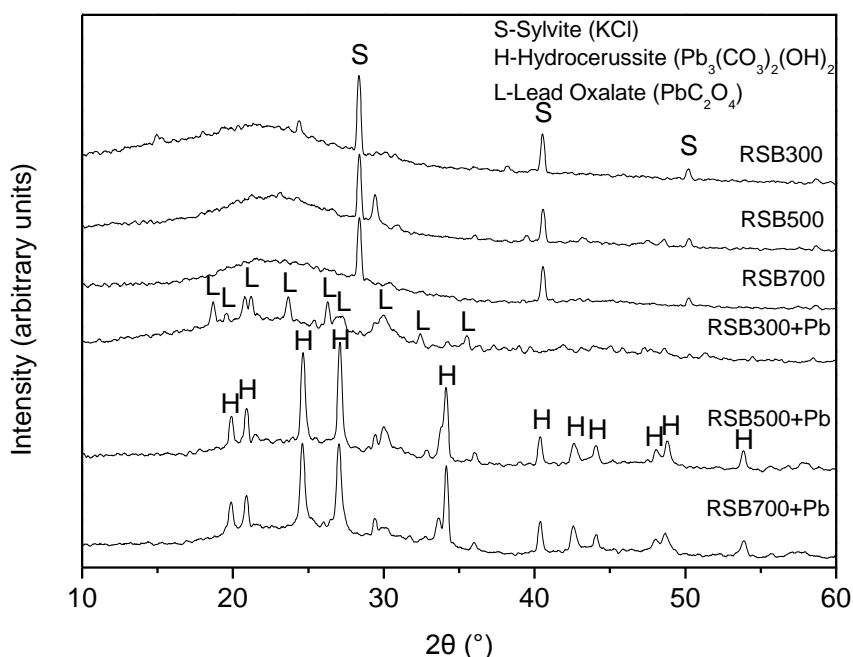


Figure 3 XRD patterns of the rice straw biochars before and after lead adsorption

3.2 Adsorption kinetics and isotherms

The adsorption kinetics of lead on the rice straw biochars are shown in Figure 4 and Table 2. The kinetics data were better described using a pseudo second order model (R^2 of 0.88-0.97), which was originally created to describe the kinetics of heavy metal removal by zeolites (Blanchard et al., 1984), as compared with a pseudo first order model (R^2 of 0.71-0.95) for all three biochars. The adsorption capacity (q_e) and k_2 values obtained from the pseudo second order model were both in the order of $RSB700 > RSB500 > RSB300$. This suggests that higher production temperature can both aid in higher Pb adsorption capacity and faster adsorption kinetics (Plazinski et al., 2009). Nearly all of the Pb (94%) was removed by RSB700 within 1 h. In

comparison, RSB500 and RSB300 only removed 73% and 65% of the total Pb at 1 h,
 respectively, and required 6 h to reach >90% total Pb removal from solution.

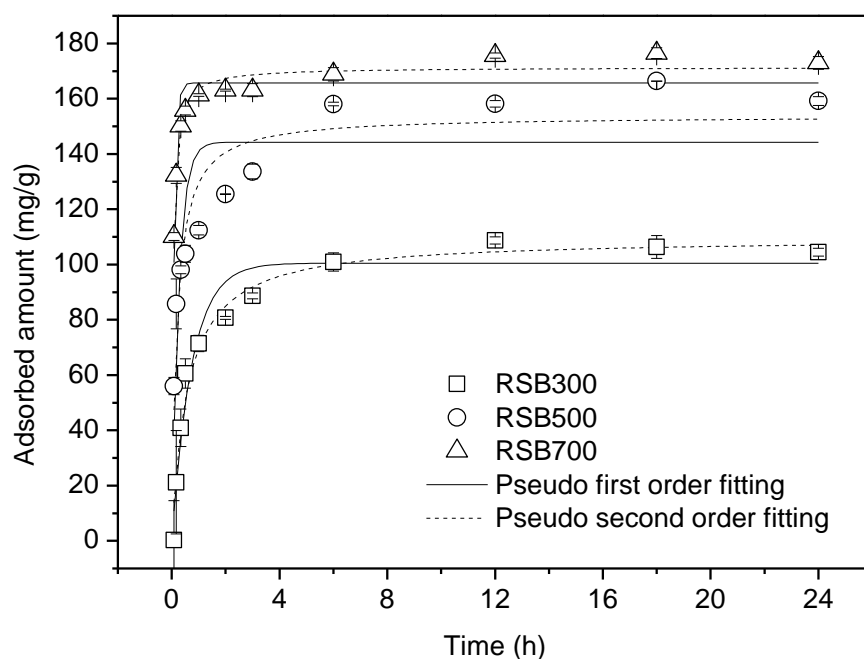


Figure 4 Adsorption kinetics of lead on rice straw biochars (0.1 g biochar in 20 mL of 5mM Pb solution)

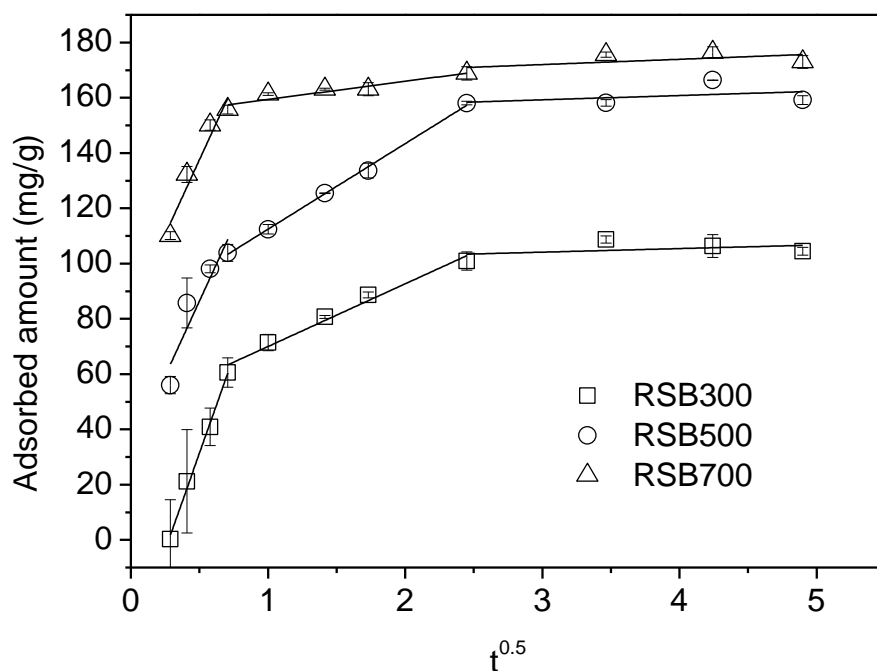
Table 2 Kinetics parameters for lead adsorption to the rice straw biochars (0.1 g biochar in 20 mL of 5mM Pb solution).

Biochar	Pseudo first order			Pseudo second order		
	q_e (mg/g)	k_1 (h^{-1})	R^2	q_e (mg/g)	k_2 (g/mg h)	R^2
RSB300	100.50 ± 3.76^a	1.36 ± 0.21	0.95	109.59 ± 3.42	0.02 ± 0.003	0.97
RSB500	144.25 ± 7.42	3.82 ± 0.93	0.71	153.81 ± 5.46	0.03 ± 0.008	0.88

RSB700	165.68±2.98	11.22±1.35	0.81	171.34±1.41	0.12±0.01	0.97
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250 (a – mean ± standard error)

251 The kinetics were further analyzed by intraparticle plotting (Figure 5 and Table S1).
252 The three biochars exhibit the same three stages of adsorption. The first fast stage is
253 due to the movement of Pb across the external liquid film boundary layer to the
254 external surface sites of biochars (Choy et al., 2004). The second, slower stage is
255 attributed to the migration of Pb within the pores of the biochars by intraparticle
256 diffusion (Choy et al., 2004). The third stage, which appears nearly flat, occurs
257 because the final stage of Pb adsorption was at internal surface sites, which is rapid
258 (Choy et al., 2004). Equilibrium sorption was complete by approximately 6 h, and
259 therefore remained unchanged from 6 to 24 h. The majority of total adsorbed Pb had
260 been adsorbed on RSB700 after the first stage. This was because RSB700 has the
261 largest surface area (Table 1) amongst the three biochars, and therefore has the most
262 accessible surface sites to seat the Pb ions. The data additionally indicate that
263 intraparticle diffusion is the main rate-limiting step for lead adsorption on all the three
264 biochars.



265

266 Figure 5 Plots of intraparticle diffusion for lead adsorption on the rice straw biochars

267 The adsorption isotherms are shown in Figure 6 (equilibrium pH shown in Table S2).

268 The Langmuir model generally shows better goodness of fitness for the equilibrium

269 data, except RSB700, for which Freundlich resulted in a slightly better fit. The

270 calculated maximum monolayer adsorption capacities for RSB300, RSB500 and

271 RSB700 were 100.27, 164.62 and 198.18 mg/g respectively. This finding was in line

272 with the kinetics findings that the rice straw biochars with higher production

273 temperatures tend to have higher Pb adsorption capacities. The observed maximum

274 monolayer adsorption capacities of the rice straw biochars in this study were higher

275 than most of the reported adsorption capacities of biochars reported in existing

276 literature (Inyang et al., 2015; Li et al., 2017), suggesting that rice straw is a promising

277 biochar feedstock for heavy metal adsorption.

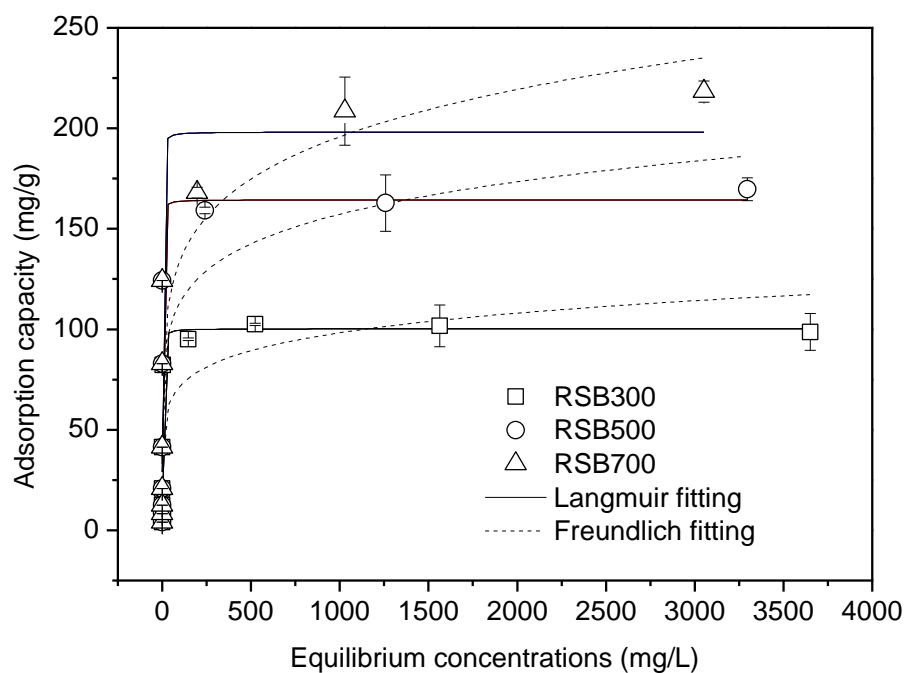


Figure 6 Adsorption isotherms of lead on rice straw biochars (0.1 g biochar in 20 mL solution)

Table 3 Parameters for Langmuir and Freundlich modelling (0.1 g biochar in 20 mL solution)

Biochar	Langmuir			Freundlich		
	Q_{\max} (mg/g)	b (L/mg)	R^2	K_f (mg/g)	$1/n$	R^2
RSB300	100.27 ± 2.88^a	1.30 ± 0.22	0.98	38.18 ± 8.65	0.14	0.74
RSB500	164.62 ± 21.81	2.26 ± 1.10	0.71	59.15 ± 15.66	0.14	0.65
RSB700	198.18 ± 25.40	1.96 ± 1.08	0.73	63.13 ± 6.10	0.16	0.77

(a – mean \pm standard error)

284 3.3 Adsorption mechanisms

285 After batch adsorption experiments, the rice straw biochars were tested by XRD,
286 FT-IR and sequential extractions to investigate the immobilization mechanisms. The
287 XRD results (Figure 3) show that lead oxalate (PbC_2O_4) was formed on RSB300,
288 whereas hydrocerussite ($\text{Pb}_3(\text{CO}_3)_2(\text{OH})_2$) was formed on RSB500 and RSB700.
289 Low-temperature rice straw biochars (100-300 °C) favor the formation of calcium
290 oxalate (CaC_2O_4) (Qian et al., 2016). With the increase of production temperature (to
291 500 and 700 °C), the calcium oxalate decomposes and forms calcite (CaCO_3) (Qian et
292 al., 2016). This explains the formation of lead oxalate for RSB300 and hydrocerussite
293 for RSB500 and RSB700 after Pb adsorption. The XRD results generally suggest that
294 precipitation was a mechanism for Pb removal by the rice straw biochars. The FT-IR
295 results (Figure 2) also show the presence of CO_3^{2-} for RSB500 and RSB700 after Pb
296 adsorption, which is likely due to the newly-formed hydrocerussite.

297 The fractions of Pb immobilized by the biochars are shown in Figure 7 and Figure S1.
298 The stable Pb concentrations on RSB300, RSB500 and RSB700 were 61.50, 91.59
299 and 99.11 mg/g, accounting for 60.01%, 57.55% and 59.00% of their total Pb loadings,
300 respectively. The acid soluble Pb on RSB300, RSB500 and RSB700 was 29.33, 66.18
301 and 68.40 mg/g, accounting for 28.62%, 41.58% and 40.72% of their total Pb loadings
302 respectively. These results clearly show that the increased Pb removal capacity by
303 increased production temperature for the rice straw biochars (Table 3) comes from the
304 increased acid soluble and stable Pb. In contrast, the exchangeable Pb significantly
305 decreased for the high-temperature rice straw biochars. RSB300 has 11.64 mg/g

exchangeable Pb, accounting for 11.34% of its total immobilized amount, however, RSB500 and RSB700 only have 1.39 and 0.47 mg/g exchangeable Pb, which are 0.87% and 0.28% of their total immobilized amount respectively.

The higher exchangeable Pb concentration for RSB300 was due to its having more exchangeable cations (H^+ , K^+) that exchanged with Pb^{2+} , as compared with RSB500 and RSB700. These cations come from carboxylic and phenolic groups on RSB300 (Dodson, 2011). With the increase of production temperature, the carboxylic and phenolic groups decompose, and H^+ and K^+ form H_2O and K_2O (Dodson, 2011). Therefore, RSB500 and RSB700 have less exchangeable cations for Pb exchange, but higher pH due to the loss of H^+ and formation of alkaline minerals (e.g., K_2O) (Dodson, 2011). The higher pH of RSB500 and RSB700 thereby aids the formation of hydrocerussite.

3.4 Implications for soil remediation

Soil contamination by heavy metals is a great environmental concern in China and globally (Hou et al., 2017a). It is important to seek greener and more sustainable technological solutions to address contaminated land (Hou et al., 2017b; O'Connor et al., 2018a). The rationale for biochar application in soil remediation is to immobilize the heavy metals within the soil and thereby reduce environmental risks that result from their presence. As mentioned in the introduction section, biochar can immobilize/adsorb heavy metals through a range of mechanisms, and each mechanism results in different binding strengths and environmental risks in the

327 context of the soil environment.

328 In this study, the majority of Pb immobilized by the biochars are generally stable
329 according to sequential leaching results (Filgueiras et al., 2002), suggesting that the
330 majority of Pb on rice straw biochars are of low risk in the context of the soil
331 environment. RSB300 has 11.34% of total immobilized Pb attributing to exchangeable
332 fraction, which is highly mobile and of high risks. Therefore, if applied in soil
333 remediation, a certain amount of immobilized Pb on RSB300 may easily leach to the
334 environment, suggesting that RSB300 may not be as appropriate for Pb
335 decontamination as compared with RSB500 and RSB700, which have very low
336 exchangeable fractions. RSB500 and RSB700 have ~40% of the total immobilized Pb
337 attributed to the acid soluble fraction. This Pb may be stable in the short-term,
338 however, when applied in soil, may pose risks after field ageing (e.g., acidic rain
339 exposure) in the long term.

340 The XRD results show that a certain amount of lead was precipitated to lead oxalate
341 on RSB300 and hydrocerussite on RSB500 and RSB700. Lead oxalate is slightly
342 soluble (e.g., K_{sp} of 10^{-9} , and 4.5 mg/L Pb^{2+} for saturated solution of lead oxalate in
343 water at 26 °C (Kolthoff et al., 1942)). In addition, higher oxalate concentrations (e.g.,
344 ≥ 0.05 M) can significantly dissolve lead oxalate due to complexation (Kolthoff et al.,
345 1942). Lower solution pH (e.g., < 4) can also dramatically dissolve lead oxalate (Sayer
346 et al., 1999). Therefore, lead oxalate is unstable in the soil environmental under
347 aggressive environmental conditions and may pose risks in the long term.

348 In comparison, hydrocerussite has a much lower solubility (e.g., K_{sp} of 10^{-47} , and ~
349 0.08 mg/L Pb^{2+} for saturated solution at 20 °C (Mohammadzadeh et al., 2015)).
350 RSB500 and RSB700 have significantly higher pH values (Table 1) as compared to
351 RSB300, and consequently have much stronger buffering capacities against
352 conditions such as acidic rain in the context of the soil environment. Therefore,
353 RSB500 and RSB700 are more applicable for soil remediation of divalent metals such
354 as Pb.

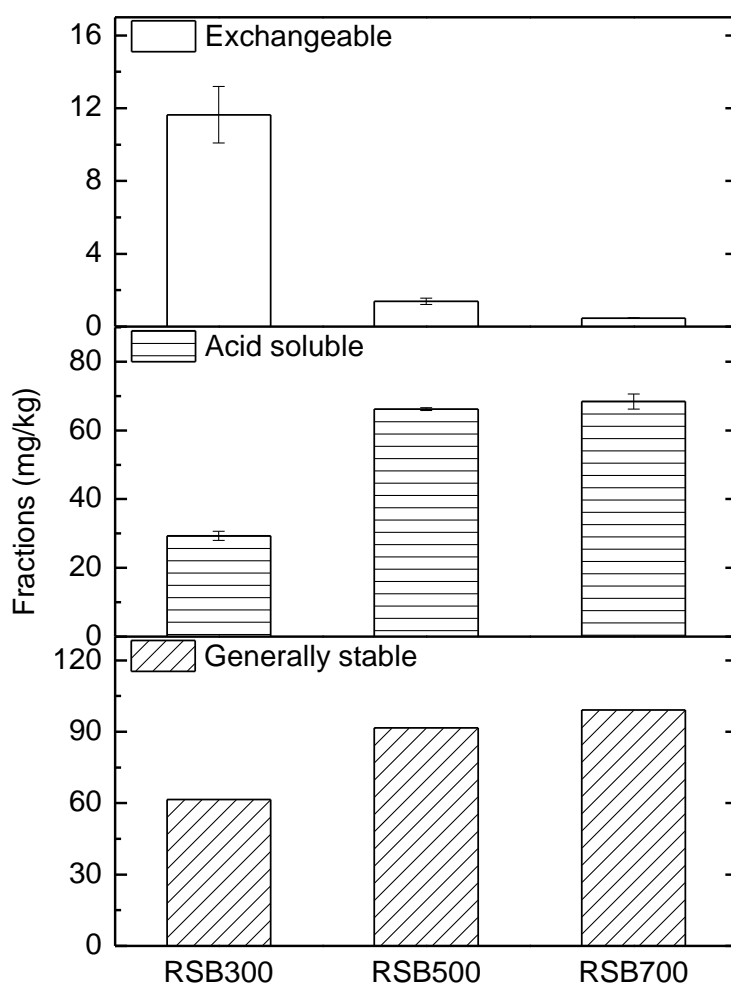


Figure 7 Fractions (mg/mg) of lead adsorbed on the biochars

4 Conclusions

Rice straw biochars were produced at 300, 500 and 700 °C. The influence of production temperature on the adsorption characteristics and mechanisms of lead on rice straw biochars were investigated. Biochars produced at higher temperatures have significant higher pH values and surface areas, resulting in higher Pb removal capacities and faster adsorption kinetics. Precipitation was a mechanism by which all three biochars immobilized Pb: lead oxalate was precipitated on RSB300 and hydrocerussite was precipitated on RSB500 and RSB700. The immobilized lead on the biochars can be divided into exchangeable, acid soluble and non-available fractions. RSB300 has 11.34% of total immobilized Pb attributed to the exchangeable fraction, whereas for RSB500 and RSB700, this fraction was less than 1%. The immobilized Pb on RSB500 and RSB700 is attributable to acid soluble and non-available fractions (>99%). Based on the mechanistic analyses discussed above, RSB500 and RSB700 are much more appropriate for soil remediation as compared with RSB300. Field-scale applications of rice straw biochars for heavy metal stabilization in soil are suggested for future research, to verify the laboratory findings of the present study.

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