

Review

TGF β , smooth muscle cells and coronary artery disease: a reviewEmma L. Low^a, Andrew H. Baker^b, Angela C. Bradshaw^{a,*}^a Institute for Cardiovascular and Medical Sciences, University of Glasgow, 126 University Place, Glasgow G12 8TA, UK^b Queen's Medical Research Institute, University of Edinburgh, 47 Little Crescent, Edinburgh EH16 4TJ, UK

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ABSTRACT

Excessive vascular smooth muscle cell (SMC) proliferation, migration and extracellular matrix (ECM) synthesis are key events in the development of intimal hyperplasia, a pathophysiological response to acute or chronic sources of vascular damage that can lead to occlusive narrowing of the vessel lumen. Atherosclerosis, the primary cause of coronary artery disease, is characterised by chronic vascular inflammation and dyslipidemia, while revascularisation surgeries such as coronary stenting and bypass grafting represent acute forms of vascular injury. Gene knockouts of transforming growth factor-beta (TGF β), its receptors and downstream signalling proteins have demonstrated the importance of this pleiotropic cytokine during vasculogenesis and in the maintenance of vascular homeostasis. Dysregulated TGF β signalling is a hallmark of many vascular diseases, and has been associated with the induction of pathological vascular cell phenotypes, fibrosis and ECM remodelling. Here we present an overview of TGF β signalling in SMCs, highlighting the ways in which this multifaceted cytokine regulates SMC behaviour and phenotype in cardiovascular diseases driven by intimal hyperplasia.

1. Introduction

Classic ultrastructural studies by Schwartz *et al* were the first to show the presence of morphologically identifiable vascular smooth muscle cells (SMCs) migrating through the internal elastic lamina following acute vascular injury in a rat model of balloon angioplasty [1]. Later, seminal work by Clowes *et al* using [³H]-thymidine labelling showed that over 40% of medial SMCs were actively proliferating 48 hours post-injury, indicating that a large proportion of SMCs within the vascular wall retain the capacity to re-enter the cell cycle and contribute to vascular remodelling and repair in adult animals [2]. This phenotypic plasticity of SMCs is now understood to play a significant role in the development of intimal hyperplasia, a pathological vascular remodelling process that occurs during the development of coronary artery disease following prolonged exposure to dyslipidaemia, hypertension and inflammation [3–5] or as a consequence of revascularisation surgery, such as coronary artery bypass grafting (CABG) or percutaneous coronary intervention (PCI) [6]. In the latter, a combination of ischemic-reperfusion injury, acute physical damage and increased longitudinal and circumferential shear stress results in endothelial cell (EC) activation, triggering the release of cytokines and growth factors, including transforming growth factor-beta (TGF β) [7]. In concert, these growth factors and cytokines drive the de-differentiation of quiescent ‘contractile’ SMCs into an active ‘synthetic’ state,

in which they display enhanced proliferation, migration and secretory capacity [6].

2. The TGF β signalling pathway

TGF β is the prototype of the highly-conserved TGF β superfamily, members of which are potent regulators of SMC phenotype and function in vascular homeostasis and disease [8]. TGF β superfamily share the same overall structure, consisting of two extended monomers held together by an intermolecular disulphide bond [9]. All TGF β monomers incorporate a characteristic ‘cysteine knot’ structure, composed of three intramolecular disulphide bonds linking six conserved cysteine residues [10]. Three TGF β isoforms are expressed in mammals (TGF β 1–3) and are differentially localised in major blood vessels during development, with TGF β 1 highly localised to the tunica intima, TGF β 2 restricted to the tunica media and TGF β 3 expressed throughout the whole vessel wall [11,12]. In adults, TGF β 1 and TGF β 3 proteins are mainly localised to the arterial intima, with TGF β 1 present in around 50% of the intimal stellate-shaped SMC population [13]. TGF β is secreted as part of a large latent complex (LLC), consisting of the C-terminal mature TGF β peptide and N-terminal latency associated peptide (LAP) covalently bound to large latent TGF β binding proteins (LTBP) [14]. LTBPs stabilise latent TGF β complexes and facilitate their retention at the cell surface through direct interactions with fibrillin and other ECM proteins [15], while

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RGD sequences in the LAP target latent TGF β to integrin receptors [16]. Activation of latent TGF β at the cell surface is induced primarily by proteases such as furin and plasmin, which cleave the covalently-bound LAP-LTBP pair from the mature TGF β molecule [17]. Proteolytic cleavage of LAP-LTBP yields short-lived, biologically active TGF β homodimers which are able to interact with transmembrane TGF β type III receptors such as betaglycan (also known as T β RIII) and endoglin [18]. Betaglycan is expressed in the majority of cell types, whereas endoglin is most abundantly expressed in vascular ECs, although recent studies have also shown localisation to SMCs in diseased vessels [19–22]. Both betaglycan and endoglin are now thought to have important cellular functions beyond their actions as TGF β co-receptors, which are reviewed at length elsewhere [23,24].

Binding to type III accessory receptors facilitates TGF β signalling through presentation of ligand to signal transduction receptors at the cell surface. Active TGF β homodimers signal via specific transmembrane heteromeric complexes comprised of two type I and two type II serine/threonine kinase receptors [25]. Five TGF β superfamily type II receptors and seven type I receptors exist in mammals [26]. The type I and type II receptors are structurally similar with small cysteine-rich ECDs (100–140 amino acids), single TMDs (30–35 amino acids) and highly conserved intracellular serine/threonine kinase domains (S/TKD; 350–400 amino acids) [9]. Each member of the TGF β superfamily binds to a characteristic combination of type I and type II receptors (Table 1). Analysis of the crystal structures of TGF β ligand:receptor ternary complexes has revealed that the length and conformation of the ligand fingertips and receptor ligand binding loops are important determinants of ligand: receptor specificity [27]. These studies have illustrated that TGF β ligands use their conserved Site IIa in their fingertip region to bind the β 1 and β 2 strands within the ECD of the TGF β type II receptor (T β RII) [28]. Importantly, the β 4- β 5 region within the ECD of the T β RII contains a 5–8 amino acid insertion which ensures type II receptor specificity by blocking binding of T β RII to bone morphogenetic protein (BMP) ligands. Of the five mammalian type II receptors, TGF β binds specifically to T β RII (also known as TGFBR2), which is highly expressed throughout the intima and media of adult vessels [13].

Table 1
- Ligands, receptors and R-Smads in the TGF β superfamily

Ligand	Type I receptor	Type II receptor	Type III receptor	R-Smad
TGF β 1	ALK1/5	T β RII	Betaglycan	Smad1/5/8
TGF β 2			Endoglin	Smad2/3
TGF β 3				
BMP2	ALK3/6	BMPRII	RGM	Smad1/5/8
BMP4			Betaglycan/Endoglin	
BMP5	ALK2/3/6	BMPRII	Betaglycan	Smad1/5/8
BMP6		ActRIIA	Endoglin	
BMP7		ActRIIB		
BMP8A	ALK3/5	BMPRII/ActRIIA	Not known	Smad1/5/8
BMP8B		ActRIIB/T β RII		Smad2/3
BMP9	ALK1/3/6	BMPRII/ActRIIA	Endoglin	Smad1/5/8
BMP10		ActRIIA		
GDF7	ALK2/3/6	BMPRII/ActRIIA/ActRIIB	Not known	Smad1/5/8
GDF6		ActRIIA/ActRIIB		
GDF5		ActRIIB		
AMH	ALK2/3/6	AMHRII	Not known	Smad1/5/8
Activin A/AB/B	ALK4	BMPRII	Betaglycan	Smad2/3
GDF8		ActRIIA	Endoglin	
GDF11		ActRIIB		
BMP16/Nodal	ALK7	BMPRII/ActRIIA/ActRIIB	Not known	Smad2/3

TGF β = transforming growth factor beta, BMP = bone morphogenetic protein, GDF = growth/differentiation factor, AMH = anti-Müllerian hormone, RGM = repulsive guidance molecule

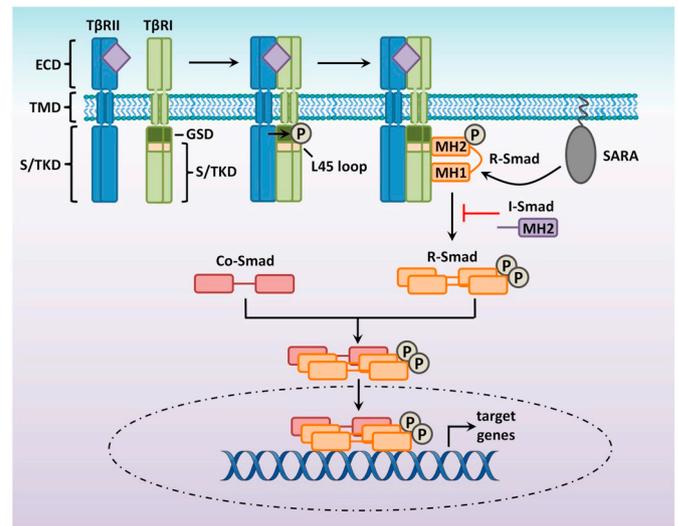


Fig. 1. - Canonical TGF β signalling pathway. Active TGF β homodimers signal via binding to specific transmembrane receptor complexes comprised of two type I (T β RI) and two type II (T β RII) serine/threonine kinase receptors. T β RI and T β RII are structurally similar with small cysteine-rich extracellular domains (ECD), single transmembrane domains (TMD) and highly conserved intracellular serine/threonine domains (S/TKD). TGF β binding to T β RII induces the assembly of T β RII and T β RI receptors into a heteromeric complex, within which constitutively active T β RII phosphorylates T β RI at several serine and threonine residues within its conserved glycine-serine domain (GSD). R-Smads become phosphorylated by the activated T β RI at their C-terminal SSXS motif. The L45 loop of T β RI and the L3 loop of the R-Smad MH2 domain determine R-Smad receptor specificity, with ALK5 specifically phosphorylating Smads 2 and 3. The adaptor protein Smad anchor for receptor activation (SARA) can also facilitate recognition of R-Smads by the receptors. I-Smads contain MH2 domains and can act to turn off Smad TGF β signalling by interfering with Smad-receptor or Smad-Smad interactions. Phosphorylated R-Smads form a heteromeric complex with Co-Smad, accumulate in the nucleus and directly regulate the transcription of specific target genes.

Early membrane crosslinking studies confirmed the expression of T β RII in SMCs, also showing binding of I¹²⁵TGF β 1 to receptor complexes composed of type I, II and III TGF β receptors [29]. T β RII ligand binding induces the assembly of type I and type II receptors into a heteromeric complex, within which constitutively active T β RII phosphorylates type I receptors at several serine and threonine residues within their conserved glycine-serine (GS) domains [8,30]. TGF β ligands principally signal via activin receptor-like kinase 5 (ALK5, a type I receptor also known as T β RI) [14]. In addition to ALK5, TGF β can also signal via another type I receptor called activin receptor-like kinase 1 (ALK1), via a distinct Smad-mediated signalling pathway to ALK5 [31–35]. While ALK5 is predominantly expressed in medial SMCs in vessels from healthy adult animals, ALK1 is chiefly localised to the endothelium, although it is upregulated in SMCs following acute vascular injury or during atherogenesis [36–38]. Following activation, type I TGF β receptors propagate the signal inside the cell through activation of the canonical Smad signalling pathway, as well as other Smad-independent kinase pathways (Fig. 1; [25]). Readers are directed to a series of excellent reviews on TGF β signalling via non-canonical kinase pathways [26,39,40].

3. Canonical Smad TGF β signalling

Smad proteins are the principal intracellular mediators of TGF β superfamily signalling. Of the eight Smad proteins expressed in mammals (Smads 1–8), Smads 2 and 3 are the primary receptor-regulated Smads (or R-Smads) activated by receptors for the three TGF β ligands [25,41]. Smad4, also known as Co-Smad, serves as a common partner

for all R-Smads. Smad6 and Smad7 are inhibitory Smads (I-Smads) which act to turn off Smad TGF β signalling by interfering with Smad-receptor or Smad-Smad interactions [25]. In general, all Smads are widely expressed throughout development and in adult animals [42]. The R-Smads and Co-Smad share homologous N- and C-terminal regions, called the Mad-homology 1 (MH1) and MH2 domains respectively, separated by a divergent proline-rich linker region [43]. I-Smads contain conserved MH2 domains but do not possess MH1 domains [8]. With the exception of Smad2, the MH1 domains of Smads exhibit sequence specific DNA binding activity, whereas MH2 domains mediate Smad oligomerisation and Smad-receptor interactions [25,44]. The linker region of R-Smad contains multiple phosphorylation sites which allow specific crosstalk with other signalling pathways including mitogen-activated protein kinases (MAPKs) and cyclin-dependent kinases, and a PY motif which mediates specific interactions with the Smurf ubiquitin ligases [25].

In non-stimulated cells, Smads undergo a constant process of nucleocytoplasmic shuttling, with the rate of nuclear export being higher than the rate of import, such that the R-Smads are predominantly localised to the cytoplasm [45]. In contrast, I-Smads tend to be localised within the nucleus in non-stimulated cells and Smad4 is distributed equally between both compartments [46]. Upon ligand stimulation, R-Smads become phosphorylated by the activated type I receptor at their C-terminal SXS motif, which increases their affinity for Smad4 [25]. The L45 loop of the type I receptor (located adjacent to its GS region) and the L3 loop of the R-Smad C-terminal domain determine R-Smad receptor specificity. The primary TGF β type I receptor in SMCs, ALK5, specifically phosphorylates Smads 2 and 3 [47,48]. Receptor recognition of R-Smads can be facilitated by auxiliary proteins, such as the adaptor protein, Smad anchor for receptor activation (SARA). SARA contains a phospholipid binding FYVE domain which targets Smads 2 and 3 to the plasma membrane and early endosomes, where it facilitates their interaction with the activated T β RI [49]. Phosphorylated R-Smads form a heteromeric complex with Smad4 and accumulate in the nucleus following importin-mediated nuclear translocation [25].

Nuclear R-Smad/Smad4 complexes bind directly to Smad-binding elements (SBE) in the promoters of TGF β target genes via a highly conserved β -hairpin loop within their MH1 domain [50]. Although many Smad-responsive promoter regions contain one or more SBEs [50], oligonucleotide binding assays have shown that Smad complexes can also recognise and bind GC-rich promoter sequences, demonstrating a relaxed DNA-binding specificity of the Smad MH1 domain [41]. As the affinity of Smad binding to a single SBE is insufficient to support sustained binding to DNA in the absence of co-operating transcriptional partners [50,51], they exert the majority of their effects on gene expression in co-operation with DNA binding co-factors, co-activators and co-repressors [41]. For example, the transcription factor δ EF1 (also known as ZEB-1) is selectively expressed in SMCs and transactivates the promoters of SMC differentiation markers following TGF β 1 stimulation of SMCs, by directly binding Smad3 and serum response factor (SRF) [52]. Similarly, the transcriptional coactivator myocardin physically associates with Smad3 in SMCs, co-ordinately transactivating the promoters of the *SM22a*, smooth muscle myosin heavy chain (*SMMHC*) and smooth muscle α -actin genes (*ACTA2*; [53]). Thus, while Smad proteins are ubiquitously expressed, the expression of Smad transcriptional partners is generally restricted to certain cell types, thereby providing a mechanism for cell lineage-specific gene responses [41]. Readers are directed to two excellent recent reviews on the contextual control of gene transcription elicited by Smad proteins [54,55].

4. TGF β in coronary artery disease

Coronary artery disease (CAD) is primarily caused by atherosclerosis, which leads to the formation of occlusive, lipid-rich plaques in affected vessels (Fig. 2A) [56]. Prolonged exposure to cardiovascular

risk factors such as dyslipidemia, hypertension and inflammation promotes endothelial dysfunction, which precedes atherosclerotic lesion formation [3–5]. The increased vascular permeability of dysfunctional, activated endothelial cells (ECs) promotes the entry of low density lipoproteins (LDLs) from the circulation into the vascular intima. Proteoglycans in the arterial wall (such as versican, biglycan and decorin) bind and retain LDLs, which become oxidised (oxLDL; [57–59]). OxLDL induces the secretion of chemokines and the expression of leukocyte adhesion molecules, which together promote monocyte infiltration into the sub-endothelial space [60]. Within the intima, SMC- and EC-derived cytokines induce monocytes to differentiate into macrophages that engulf oxLDL, forming foam cells. In turn, inflammatory cells within the early lesion secrete cytokines and growth factors which promote the development of intimal hyperplasia. Resident SMCs are key drivers of intimal hyperplasia in the initiation and early progression of atherosclerosis, which is characterised by SMC dedifferentiation, proliferation and migration [61]. Secretory SMCs synthesise an abundant array of ECM components, which form a fibrous cap over the plaque, further encroaching on the vessel lumen [62]. Increased synthesis of proteoglycans by secretory SMCs also promotes lipoprotein retention in the growing lesions, while dedifferentiated SMCs acquire phenotypic characteristics of the osteoblast, adipocyte and macrophage lineages [63]. Advanced, rupture-prone plaques are characterised by lipid-rich necrotic cores (composed of apoptotic foam cells and cellular debris) thin fibrotic caps (a consequence of matrix metalloproteinase secretion), vascular calcification and neoangiogenesis.

Several genome-wide association studies (GWAS) have identified an association between CAD and single nucleotide polymorphisms (SNPs) in genes encoding TGF β signalling pathway components. For example, functional polymorphisms in the promoter, signal peptide sequence and coding sequence of the *TGF β 1* gene are associated with increased risk of myocardial infarction [64,65] and stroke [66]; meta-analyses have also shown an association between these polymorphisms and CAD [67–70]. In addition, a joint analysis of two GWAS on CAD patients identified an association with an intronic SNP in the *SMAD3* gene [71] which was later shown to reduce enhancer activity and attenuate Smad3 expression [72,73]. Clinical studies have illustrated that plasma levels of active TGF β 1 are markedly reduced in patients with advanced atherosclerosis compared with healthy controls [74,75]. In contrast, other groups have reported an increase in active TGF β 1 levels in the plasma of CAD patients, where patients with triple vessel disease had twice the level of circulating active TGF β 1 compared with those with no or mild CAD [76]. These differences may be due to discrepancies between sample preparation methods, which can affect the level of TGF β 1 protein detected in plasma [77]. Nevertheless, immunolocalisation studies of human atherosclerotic lesions strongly support a role for TGF β in the pathogenesis of CAD, showing high levels of TGF β 1 and TGF β 3 in SMCs and macrophage-derived foam cells in early fatty streak lesions, co-localising with T β RII and ALK5 [13]. > 50% of SMCs in early lesions also stain positive for phospho-Smad2, indicative of TGF β signalling pathway activation in these cells [78]. Advanced fibrous lesions also express significant amounts of TGF β 1, whereas TGF β 3, T β RII and ALK5 are expressed at more reduced levels in the fibrous plaque and the associated media [13].

Studies in experimental models of atherosclerosis indicate that TGF β can be both atheroprotective and atherogenic. Early animal studies using global TGF β inhibition or genetic knockout approaches demonstrated that reduced availability of TGF β was pro-atherogenic and associated with the development of macrophage-rich, pro-inflammatory plaques which were prone to rupture [79]. Mice heterozygous for the deletion of *tgfb1* on a cholesterol-enriched diet had reduced SMC differentiation (determined by levels of α SMA and SMMHC, two mature SMC marker proteins), accelerated lipid lesion formation and increased vascular inflammation compared with wild-type littermate controls [80]. Similarly, administration of a neutralising anti-TGF β 1 antibody [81], or systemic infusion of a dominant negative T β RII in

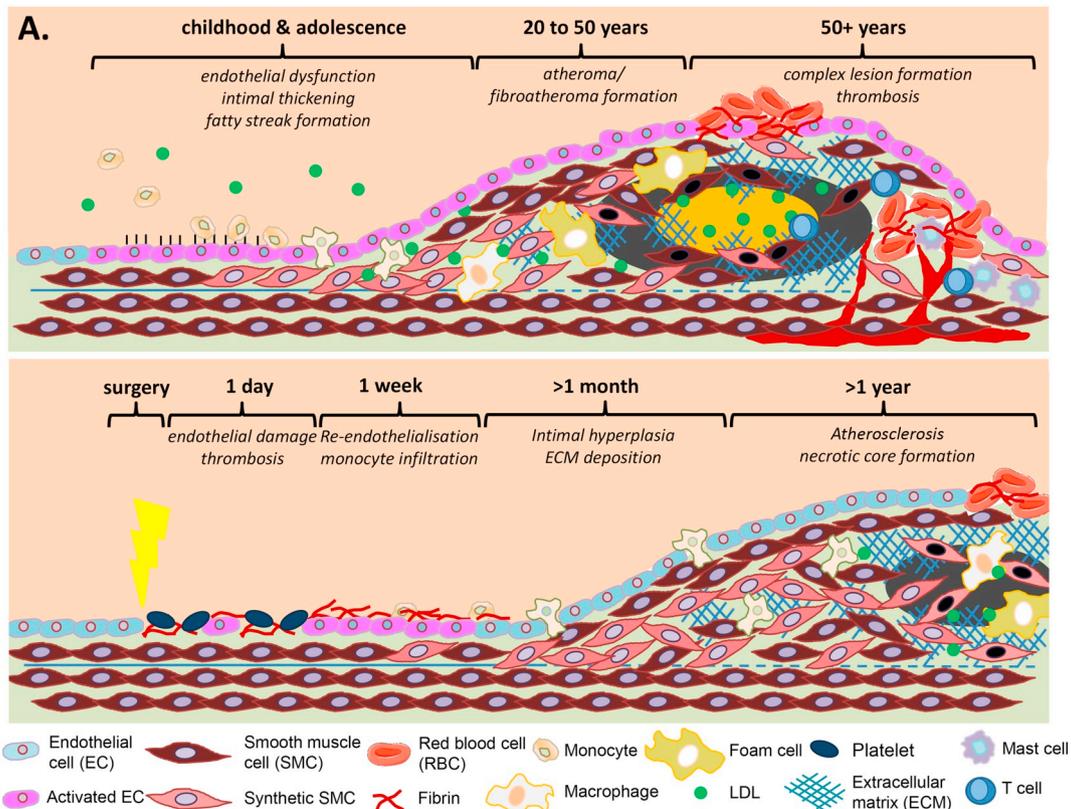


Fig. 2. – Vascular remodelling during atherosclerosis (A) and after revascularisation surgery (B) (A) Atherosclerosis is initiated by the activation of the endothelium in response to oxidative, haemodynamic or biochemical stimuli. Activated endothelial cells (ECs) upregulate surface adhesion molecules and secrete growth factors and cytokines, promoting rolling adhesion of circulating leukocytes as well as activation of the underlying smooth muscle cells (SMCs). Activated SMCs dedifferentiate and start proliferating and migrating, contributing to the growing neointima. Leukocytes adhering to the endothelium migrate into the intima through diapedesis, maturing into macrophages and phagocytosing low density lipoproteins to become foam cells, characteristic of the ‘fatty streak’ lesions that can be observed from adolescence onwards. Fibroatheromas form from areas of intimal thickening, which consist of foam cells, remnants of apoptotic SMC and a lipid rich ECM pool. Early fibroatheromas are characterised by an acellular necrotic core and a thick fibrous cap, composed of collagen fibrils interspersed with SMCs. Advancing fibroatheromas contain cholesterol crystals, neovessels and lymphocytes, and have thin fibrous caps due to proteolytic ECM degradation, making these lesions particularly susceptible to rupture and thrombosis. Rupture and thrombosis frequently occurs at the shoulder regions of plaques, where mast cells accumulate and secrete pro-angiogenic factors and enzymes to further promote microvessel formation. (B) Vein graft implantation or coronary stent deployment induces endothelial damage and denudation. Within hours, platelets and red blood cells adhere to the endothelial layer, initiating a coagulation cascade that results in the deposition of fibrin-rich layers. In the weeks following surgery, circulating leukocytes attach and infiltrate the vascular endothelium, while SMCs in the media are activated and start migrating into the growing neointima. Growth factors and cytokines released by cells in the vessel wall induce SMC proliferation and ECM deposition, resulting in further intimal thickening and inward vascular remodelling. Intimal thickening can act as a substrate for superimposed atherosclerosis or neoatherosclerosis, which is frequently observed between 2–5 years following revascularisation surgery. The pathogenesis of superimposed atherosclerosis/neoatherosclerosis bears many similarities with native coronary artery atherosclerosis (A), albeit within a much shorter timeframe.

apolipoprotein E (ApoE)-deficient mice [82] significantly enhanced lipid infiltration in the vascular wall, decreased collagen type I and III secretion by SMCs and was associated with frequent intraplaque haemorrhages. Corroborating these TGF β knockdown studies, overexpression of an activated TGF β 1 expression construct via viral gene transfer markedly reduced atherosclerotic lesion formation in fat-fed LDL receptor knockout mice [83]. In these animals, medial and intimal SMCs showed reduced expression of the oxidative stress marker nitrotyrosine, with CD68+ macrophage infiltration also substantially attenuated as a result of diminished SMC-derived M-CSF secretion [83]. Similarly, overexpression of active TGF β 1 in the hearts of ApoE $^{-/-}$ mice reduced aortic root plaque formation by decreasing inflammatory cell infiltration and increasing SMC collagen secretion to form more stable atherosclerotic lesions [84]. Interestingly, pre-incubation of rat SMCs with atorvastatin enhanced the TGF β 1-mediated activation of Smad2/3; similar results were observed in ApoE $^{-/-}$ mice treated with a moderate dose of statin, accompanied by increased collagen and α SMA staining in plaques [85]. Together, these studies support the ‘protective cytokine’ theory of atherosclerosis [86], indicating that TGF β can protect against the development of unstable plaque lesions by

promoting the expression of contractile SMC proteins, suppressing leukocyte recruitment, and reinforcing the fibrous cap by enhancing ECM production by resident SMCs.

There is, however, an important caveat to these observations; while the induction of contractile marker proteins (such as α SMA and SM22 α) by TGF β can be viewed as atheroprotective in SMCs, very recent studies investigating the origin of α SMA+ cells within atherosclerotic lesions have demonstrated that activation of this transcriptional programme by TGF β in endothelial cells (ECs) can instead promote the induction of atherosclerosis. Using endothelial lineage tracing mice on an ApoE $^{-/-}$ background (*ScfCreER^{T2}; R26RstopYFP; ApoE $^{-/-}$*), Evrard *et al* found that TGF β could induce endothelial-to-mesenchymal transition (EndMT) during atherogenesis, enhancing expression of α SMA and fibrotic markers in ECs without affecting collagen expression (87). Of note, immunohistological evaluation of human atherosclerotic lesions revealed a higher proportion of cells co-expressing endothelial and fibroblast markers in type VI plaques (complicated lesions with unstable features) compared to type V plaques (stable fibrocalcific lesions/fibroatheromas) supporting a role for TGF β -induced EndMT in the clinical context [87]. There is also accumulating evidence that TGF β can

elicit atherogenic effects through its actions on SMCs in early plaque lesions. For example, while the promotion of contractile protein expression in SMCs is an important part of TGF β 's anti-atherogenic repertoire during the later stages of plaque development, increased vascular resistance and SMC hypercontractility is also associated with the induction of atherosclerosis [88]. Additionally, TGF β is now known to be a potent inducer of proteoglycan (PG) synthesis by SMCs, enhancing the gene expression and glycosaminoglycan (GAG) sidechain elongation of PGs such as biglycan [89,90] and versican [91]. PGs directly contribute to the initiation of atherosclerosis through their electrostatic interactions with lipoproteins, promoting the retention of lipoproteins in the sub-endothelial space (reviewed in [92]). Accordingly, treatment of atheroprone LDLr^{-/-} mice with the TGF β neutralising antibody 1D11 substantially repressed biglycan expression, reducing biglycan colocalisation with apoB lipoproteins and attenuating atherosclerotic lesion formation [93]. Finally, it has recently been recognised that TGF β can drive the transdifferentiation of SMCs into proliferative, α SMA-positive migratory myofibroblasts, thereby contributing to the early development of atherosclerotic plaques, whilst on the other hand promoting stability of more advanced lesions through fibrotic cap formation [94,95]. Thus, while TGF β generally acts as a potent pro-fibrotic and anti-inflammatory mediator in CAD, the pathophysiological outcome of these actions is highly context-dependent, varying according to the specific cell type, stage of atherosclerosis (early/advanced) and type of lesion (stable/unstable).

5. TGF β in acute vascular injury: vein graft failure and restenosis

Revascularisation surgeries such as percutaneous coronary intervention (PCI) or coronary artery bypass grafting (CABG) are frequently prescribed for advanced or acute presentations of CAD, aiming to widen occluded coronary arteries. However, the long-term patency of such procedures is hampered by the development of intimal hyperplasia within the vessel, resulting in re-occlusion and the need for repeat intervention. Similar to the intimal hyperplasia (IH) that forms a fertile 'soil' for atherosclerosis in CAD, IH following revascularisation surgery is initiated by activation of the endothelium. Stent deployment (PCI) or exposure of venous bypass conduits to increased arterial shear stress (CABG) induces acute endothelial injury, leading to adhesion of circulating platelets and monocytes to the endothelium. Pro-inflammatory growth factors and cytokines released by activated endothelial cells, platelets and leukocytes drive SMC dedifferentiation, proliferation and migration, cellular processes that are critical to the development of IH. Medial SMC proliferation is rapidly induced following vascular injury and peaks around 7 days post-injury (10 - 20 % medial SMC proliferation; [96] [97]). Proliferating medial SMCs also migrate and accumulate in the intima, contributing to the overall lesion cell mass [98,99]. The resulting hyperplastic neointima can act as a substrate for accelerated atherosclerotic plaque formation, which further contributes to the occlusion of the vessel. Compared to native CAD, which takes decades to develop, vein graft atherosclerosis develops over a period of months to a few years. Lesions are also more concentric and diffuse than native atherosclerotic lesions and are more susceptible to thrombosis and rupture ([100,101]; reviewed in [102]). There is growing awareness that in-stent atherosclerosis (most frequently termed 'neoatherosclerosis') is one of the primary causes of late stent failure, with several studies showing that the development of neoatherosclerosis is accelerated in drug-eluting stents (DES) compared to first-generation bare-metal stents (BMS) [103–106]. Although the mechanisms causing neoatherosclerosis have yet to be fully characterised, histopathological and intravascular imaging studies indicate that stent-induced shear stress, chronic inflammation and endothelial dysfunction may be key contributing factors (Fig. 2B) [102].

Clinical studies of restenotic arteries following balloon angioplasty were among the first to suggest that TGF β plays a prominent role in the development of IH following revascularisation surgery. These studies

showed that the mRNA expression of TGF β 1 was significantly elevated in restenotic lesions compared with both primary atherosclerotic lesions and control non-atherosclerotic tissues [107]. IHC co-staining for TGF β 1 and α SMA within sections taken adjacent to those studied by *in situ* hybridisation demonstrated that TGF β 1 was localised to the intimal and medial SMC layers in restenotic lesions [107]. Two further groups demonstrated that the majority of human restenotic lesions showed positive staining for TGF β 1 throughout the vascular media and intima, identifying SMCs as a key source of TGF β 1 in restenotic vessels [108,109]. Pre-clinical studies support this observation, showing that TGF β 1 is upregulated in SMCs at sites of acute vascular injury in rodents. TGF β 1 mRNA levels were significantly increased in rat carotid arteries 6 hours post-injury and remained significantly elevated for at least 2 weeks [110,111]. Increased TGF β 1 expression (both at the mRNA and protein level) 2 weeks after wounding was associated with a parallel increase in fibronectin, collagen I and collagen III mRNA expression, three pro-fibrotic genes known to be regulated by TGF β [111]. In a porcine coronary angioplasty model, levels of active TGF β 1 were significantly elevated between 2 hours and 7 days following angioplasty, with immunohistochemical studies showing strong localisation to both SMCs and ECs [112]. TGF β 1 mRNA and protein levels have also been shown to be chronically upregulated 6 months post-grafting in a rabbit CABG model, accompanied by increases in connective tissue growth factor (CTGF), a well-defined TGF β 1 responsive gene [113].

Subsequently, numerous *in vivo* interventional studies have convincingly shown that TGF β promotes IH in vein grafts and models of PCI (Table 2). Overexpression of TGF β 1 in porcine arteries using an expression plasmid resulted in increased procollagen, collagen and proteoglycan synthesis by neointimal SMCs and was accompanied by marked intimal and medial hyperplasia [114]. Furthermore, adenoviral overexpression of active TGF β 1 in uninjured rat arteries resulted in a hyperplastic neointima [96] or a larger collagen-rich neointima in grafted rat arteries compared with control [115]. Similarly, infusion of purified recombinant TGF β 1 into rats 2 weeks after carotid artery injury increased [3H]-thymidine-labelled SMC nuclei within the neointima, indicating that TGF β 1 stimulates SMC proliferation as well as collagen synthesis in this model of vascular injury [111]. Substantiating these TGF β 1 overexpression studies, inhibition of TGF β by antisense treatment or by intravenous infusion of a soluble T β R2 reduced IH and adventitial fibrosis in balloon-injured rat carotid arteries [116,117]. Interestingly, adenovirus-mediated overexpression of TGF β 3 (but not TGF β 1) in pig coronary arteries inhibited constrictive remodelling and reduced lumen loss after coronary angioplasty [118]. Similarly, direct infusion of TGF β 3 to goat carotid arteries after anastomosis reduced vessel wall thickness by 30%, in part by reducing collagen type VII content 3 months post-surgery [119]. In contrast to the increased intimal hyperplasia observed in interventional studies using TGF β 1, these groups also showed reduced SMC proliferation in TGF β 3-treated animals, which suggests that there may be differences in how SMCs respond to specific TGF β isoforms *in vivo*. The intracellular signalling mechanisms that drive IH in response to TGF β have yet to be fully characterised, however the majority of studies to date have identified an important role for the canonical Smad2/3 pathway. Kundi *et al* showed that carotid artery injury in rats leads to significant induction of Smad3 in medial SMCs, while overexpression of Smad3 via gene transfer resulted in increased collagen accumulation [120] and SMC proliferation via a p27-dependent mechanism [97]. Furthermore, adenoviral overexpression of Smad7 in rat balloon-injured arteries reduced intimal thickening, lumen area loss and collagen synthesis 14 days post-injury [121], with *in vitro* studies indicating that these effects were due to direct effects on resident SMCs [122,123]. Interestingly, oral dosing of an ALK5/ALK4 small molecule inhibitor following balloon injury decreased intimal collagen production but had no effect on intimal SMC proliferation [124], suggesting that other TGF β receptor signalling pathways may be responsible for TGF β -induced SMC proliferation.

Table 2
In vivo studies employing different approaches to target TGF β activity after vascular injury

Therapy	Animal model	Outcome compared to control	Reference
Soluble T β RII	Rat carotid artery balloon injury	Reduced intimal thickening, constrictive remodelling, lumen area loss and collagen type I/III mRNA expression	Smith et al, 1999 [116]
ALK4/5/7 inhibitor (SB431542)	Rat carotid artery balloon injury	Reduced intimal thickening, neointimal SMC proliferation, reduced recruitment of MSCs	Zhao et al, 2016 [182]
ALK4/5 kinase inhibitor (SM16)	Rat carotid artery balloon injury	Reduced intimal thickening, inhibition of adventitial myofibroblast formation, collagen deposition	Fu et al, 2008 [124]
Anti-TGF β 1 ribozyme oligonucleotides	Rat carotid artery balloon injury	Reduced intimal thickening, TGF β 1 mRNA expression, collagen type I/III expression and synthesis	Yamamoto et al, 2000 [117]
Anti-TGF β 1 phosphorothioate oligonucleotides	Rabbit carotid artery balloon injury	Reduced intimal thickening, proteoglycan synthesis and TGF β 1 mRNA expression	Merrilees et al, 2000 [183]
Tranilast	Rat carotid artery balloon injury	Reduced SMC migration, TGF β 1 mRNA expression, T β RI/T β RII mRNA expression and α V β 3 mRNA expression	Ward et al, 1998 [184]
TGF β 1 antisense mRNA (adenoviral overexpression)	Rat femoral artery vein grafting	Reduced intimal thickening, reduced collagen and TIMP mRNA expression	Wolff et al, 2006 [115]
Recombinant TGF β 3	Pig coronary artery balloon injury	Reduced constrictive remodelling, lumen area loss and increased collagen synthesis	Kingston et al, 2003 [118]
Smad7 (adenoviral overexpression)	Rat carotid artery balloon injury	Reduced intimal thickening, lumen area loss, collagen synthesis and adventitial fibroblast migration	Maallawaarachchi et al, 2005
p38 MAPK inhibitors	Rat carotid artery balloon injury	Reduced intimal thickening and SMC proliferation	Ohashi et al, 2000 [185]
Pyrrrole-imidazole polyamide targeting the TGF β 1 promoter	Rat carotid artery balloon injury	Reduced intimal thickening, TGF β 1, collagen and fibronectin mRNA expression and accelerated re-endothelialisation	Yao et al, 2009 [186]

TGF β = transforming growth factor beta, BMP = bone morphogenetic protein, GDF = growth/differentiation factor, AMH = anti-Müllerian hormone, RGM = repulsive guidance molecule

6. TGF β signalling and SMC function

As stated in the introduction, studies have conclusively shown that TGF β is a potent regulator of SMC phenotype and function. The atheroprotective effects of TGF β are in part attributed to its capacity for stimulating SMC differentiation by inducing the expression of a large set of mature SMC genes (including α SMA, SM22 α and SMMHC [125]) via Smad2 and/or Smad3, which interact with the SMC-specific promoters at putative SBEs [126,127]. TGF β also induces serum response factor (SRF) protein expression and enhances its binding activity to CAR β elements within the promoters of SMC marker genes [128]. Interestingly, Qiu et al have shown that Smad3 is the primary mediator for TGF β 1-induced SM22 α expression, while Smad6 and Smad7 repress its activation [129]. Furthermore, the authors illustrated that Smad3 can bind to a SBE in the first exon of SM22 α and directly associate with the SRF complex in response to TGF β 1 treatment [129]. TGF β is also a potent inducer of the synthetic SMC phenotype, stimulating the production and secretion of collagen and proteoglycans by SMCs via direct and indirect interactions with the promoters of these genes [130–132]. However, the effects of TGF β on SMC behaviour are more variable, with studies showing that TGF β can both inhibit and stimulate SMC proliferation and migration. This may be due to the heterogeneous nature of SMCs, as evidenced by the varying gene expression patterns of human SMCs derived from primary atherosclerotic plaques, in-stent stenoses or healthy arteries [133]. At the molecular level, these differences have been attributed to varying levels of receptor expression, membrane localisation of receptors, availability of intracellular signalling mediators and presence of transcriptional co-regulators within the nucleus (reviewed in [54,134]). In the next section, we will highlight key findings on the regulation of SMC proliferation and migration by TGF β in the context of intimal hyperplasia and CAD.

7. TGF β -regulated SMC proliferation

SMC responses to TGF β *in vitro* are influenced by factors such as type of SMC (aortic, venous etc.), cellular density and concentration of TGF β [134]. For example, Majack et al found that TGF β 1 inhibited proliferation of rat aortic SMCs at sub-confluent densities but potentiated SMC growth at high seeding densities [135]. Furthermore,

treatment of cultured porcine coronary artery SMCs with low concentrations of TGF β 1 (0.025ng/mL) stimulated SMC proliferation, but attenuated SMC growth at concentrations of greater than 0.1 ng/mL [136]. The presence of other growth factors also appears to influence the effects of TGF β on SMC proliferation. For instance, treatment of rat aortic SMCs with TGF β 1 had no significant effect on cell number in quiescent SMC cultures maintained in 1 % FBS, but markedly inhibited SMC proliferation in response to 5 % FBS or PDGF-BB in a dose-dependent manner [135,137]. Other studies, however, have shown that TGF β potentiates the mitogenicity of FBS, PDGF-BB and bFGF, but only in confluent SMC cultures [138,139].

TGF β -induced inhibition of SMC proliferation *in vitro* has been associated with G0/G1 cell cycle arrest through downregulation of the cell cycle regulator, cyclin-dependent kinase 1 (CDK1) [140]. Treatment of mouse aortic SMCs with TGF β 1 for 24 hours substantially reduced the percentage of cells in S phase and G2/M phase and increased the number of cells in G0/G1 [141]. Pharmacological inhibition of the p38 MAPK pathway (using 10 μ M SB203580) resulted in complete attenuation of TGF β -dependent growth inhibition in the absence of any inhibitory effect on Smad2/3 signalling, as analysed by phosphorylation, nuclear translocation and reporter gene expression [141], indicating that p38 MAPK may mediate growth inhibition induced by TGF β in SMCs. More recently, TGF β has been shown to inhibit PDGF-induced SMC proliferation through downregulation of Cyclin D1 [142], a key regulator of cell cycle transition from G1 to S phase [143]. Here the authors demonstrated that treatment of human aortic SMCs with TGF β 1 significantly inhibited PDGF-BB-induced Cyclin D1 mRNA and protein expression after 24 hours. Interestingly, inhibition of ALK5 using 10 μ M SB431542 or siRNA-mediated knockdown of Smad4 completely abolished the inhibitory effect of TGF β on PDGF-induced Cyclin D1 expression and restored SMC proliferation in response to PDGF, suggesting that this occurs through a Smad-dependent mechanism [142].

In contrast, certain studies have shown that TGF β 1 can indirectly promote SMC proliferation in confluent cultures by inducing PDGF-A gene expression and autocrine production of PDGF-AA [138,144]. Both these studies found that TGF β -induced rat aortic SMC proliferation was mimicked by treatment with exogenous PDGF-AA (> 5 ng/ml) and partially inhibited by neutralising antibodies to PDGF-AA [138,144].

However, a later study showed that while TGF β induced an 8-fold increase in PDGF concentration after 24 hours, application of this conditioned medium (containing ~ 1 ng/mL PDGF-AA) to aortic SMCs did not increase mitogenic activity, indicating that induction of PDGF-AA production by TGF β cannot fully account for the effects of TGF β on the proliferation of rat aortic SMCs under all *in vitro* culture conditions [145]. Indeed, TGF β has also been shown to directly stimulate SMC proliferation through a Smad-dependent mechanism. For instance, Mao et al demonstrated that aortic SMCs from smooth muscle-specific Smad4 knockout mice display a 62 % reduction in proliferation *in vitro* (as determined by BrdU labelling), compared with SMCs from wild-type mice [146]. Furthermore, shRNA-mediated knockdown of Smad2 and Smad3 within wild-type SMCs significantly reduced SMC proliferation in response to 20 % FBS and the expression of SMC-specific marker genes [146].

Despite the contrasting *in vitro* data for the effects of TGF β on SMC proliferation, the majority of *in vivo* evidence indicates that TGF β is a potent stimulator of arterial SMC proliferation [96,97,111,147]. For instance, infusion of recombinant TGF β 1 into rats after carotid artery balloon injury resulted in a significant increase in the number of [3H]-thymidine labelled SMC nuclei within the neointima, compared with untreated rat coronary arteries [111]. Similarly, Schulick et al noted that localised adenoviral over-expression of TGF β 1 in the endothelium of uninjured rat carotid arteries resulted in substantial intimal thickening after 4 weeks with marked cellular proliferation (measured by BrdU incorporation) when compared with control arteries [96]. TGF β -induced SMC proliferation *in vivo* has been shown to be mediated via a Smad3-dependent mechanism, involving the phosphorylation and nuclear export of the cyclin-dependent kinase inhibitor p27 [97]. Adenoviral overexpression of Smad3 within balloon-injured rat carotid arteries significantly enhanced intimal thickening after 14 days and was associated with increased PCNA expression within intimal SMCs [97] and increased pERK MAPK expression within whole arteries and isolated SMCs [147]. Conflicting studies performed using a more damaging, inflammatory model of femoral artery wire injury showed enhanced neointimal hyperplasia and increased SMC proliferation in Smad3 knockout mice, indicating that the role of TGF β in the arterial response to injury can vary as a function of the inflammatory micro-environment [148]. Thus, TGF β /Smad3 can directly enhance SMC proliferation *in vivo* through transactivation of the ERK MAPK signalling pathway; other indirect mechanisms may account for the enhanced or repressed proliferative responses observed, including modulation of the inflammatory microenvironment or release of sequestered mitotic growth factors following ECM degradation.

8. TGF β -regulated SMC migration

Similar to SMC proliferation, TGF β has been shown to variably stimulate and inhibit SMC migration. Early *in vitro* studies performed in venous and arterial-derived SMCs showed that PDGF-BB, b-FGF or serum-induced migration is inhibited by TGF β 1 in a concentration-dependent manner and this effect is independent of cellular density [149,150]. TGF β 1 can suppress PDGF-BB-induced up-regulation of MMP-2 within rat arterial SMCs, suggesting that the indirect effects of TGF β 1 on SMC migration may partly be due to the inhibition of downstream pro-migratory genes [151]. Conversely, studies also show that TGF β can directly stimulate SMC migration. For instance, aortic SMCs from smooth muscle-specific Smad4 knockout mice displayed significantly reduced migration in response to serum or PDGF-BB *in vitro*, compared with SMCs from wild-type mice [146]. Furthermore, inhibition of ALK5 using the kinase inhibitor SB431542 or shRNA-mediated knockdown of Smad2 or Smad3 significantly attenuated SMC migration in response to serum stimulation [146]. *In vitro* studies performed on aortic SMCs have shown that TGF β can also regulate SMC migration via indirect mechanisms involving the up-regulation of α v β 3 mRNA expression, an integrin which is highly expressed following

vascular injury and is important in driving SMC migration [152–155]. Pre-treatment of human aortic SMCs with TGF β 1 was associated with enhanced migration in response to vitronectin, a serum glycoprotein which promotes cell spreading and attachment through integrin receptor binding [154]. Furthermore, treatment of injured rat carotid artery SMCs with a TGF β 1 neutralising antibody completely abrogated TGF β 1-induced integrin β 3 mRNA up-regulation [156]. Interestingly, treatment of rats with genistein (a tyrosine kinase inhibitor) following carotid artery injury markedly inhibited injury-induced up-regulation of TGF β 1, TGF β 3, integrin α v and β 3 mRNA expression, compared with vehicle-treated arteries, suggesting that induction of TGF β following vascular injury is broadly reliant on tyrosine kinases [156].

9. Therapeutic targeting of TGF β in CAD: challenges and opportunities

As documented above, TGF β plays a fundamental role in the regulation of vascular function by affecting SMC proliferation, migration, differentiation and ECM production in CAD. Mutations in genes encoding TGF β ligands and receptors are also associated with several developmental disorders and vascular diseases, including Marfan syndrome type 2, Loeys-Dietz syndrome, and other vasculopathies with clinical presentations that include thoracic aortic aneurysms and dissections [157–159]. Hence, components of the TGF β signalling pathway are important therapeutic targets for a wide range of vascular pathologies.

Numerous pre-clinical studies have employed different approaches to inhibit TGF β signalling after vascular injury, which have been shown to reduce intimal thickening compared with controls. However these approaches have yet to translate to significant clinical gain in the cardiovascular disease arena, with no TGF β therapeutics currently on the market. Promisingly, small-scale clinical trials demonstrated that oral administration of 600 mg/day tranilast (N-(3,4-dimethoxycinnamoyl) anthranilic acid), a non-specific inhibitor of TGF β biosynthesis, was associated with a significantly reduced risk of restenosis following PCI, compared with placebo (17.6% vs. 39.4% at 3 months) [160,161]. Originally developed as a treatment for allergic disorders such as chronic rhinitis and bronchial asthma, tranilast has also successfully been used (both orally and topically) as an anti-fibrotic agent in the treatment of hypertrophic scars or keloids [162–164]. However, the large-scale randomised double-blind clinical trial PRESTO (Prevention of REStenosis with Tranilast and its Outcomes) examining the effects of tranilast treatment in 11,484 patients after PCI failed to show improved clinical outcome (death, MI or repeat revascularisation) compared with placebo [165]. Worryingly, this trial highlighted some potential adverse effects of tranilast, including hyperbilirubinemia, increased serum creatinine and alanine transaminases, indicative of liver abnormalities. Fortunately these adverse effects were reversed upon cessation of treatment, however the lack of primary and secondary endpoint efficacy in this large-scale trial highlights the complexity of targeting TGF β using systemic approaches in multimorbid, highly diverse groups of patients.

Nevertheless, TGF β therapeutics are advancing in clinical trials for other indications, particularly fibrosis and oncology, and results appear to be positive [166,167]. Indeed, Pirfenidone (5-methyl-1-phenyl-2-[1H]-pyridone), which inhibits TGF β production and activity, was approved by the FDA in October 2014 for treatment of idiopathic pulmonary fibrosis (IPF). IPF is a devastating progressive lung disease, with a median survival from time of diagnosis of 3 years; Pirfenidone was approved on the basis of phase III clinical trials showing a reduction in forced vital capacity decline (a measure of lung function) and improved progression-free survival compared to placebo (ASCEND study [168]). In the oncology field, Galunisertib (LY2157299 monohydrate) a small molecule inhibitor of the ALK5 kinase, has been evaluated in > 10 clinical trials (alone or in combination with e.g alkylating agents) for different types of cancer [169]. The most advanced

trial currently in progress is a phase II/III randomised, placebo-controlled trial that has enrolled ~140 patients with myelodysplastic syndrome (MDS; NCT02008318); interim data from this trial shows good tolerance of the drug and haematological improvement in 26% of patients enrolled. Of note, recent trials investigating the use of Galunisertib have utilised an adapted, intermittent dosing regimen (14 days on, 14 days off) due to preclinical studies showing proliferative, inflammatory changes in the heart valves and aortae of rats when continuously dosed with Galunisertib [170]. Although no medically significant cardiotoxicities were observed in a first-in-human dose study administering Galunisertib to glioma patients [171], the potential for serious adverse events with high-dose, systemic TGF β agonists or antagonists should not be underestimated. Ultimately, localised and pathway-specific targeting of TGF β signalling will be required in order to achieve optimal therapeutic efficacy whilst avoiding undesired off-target effects.

While there are acknowledged challenges associated with using global approaches for targeting TGF β in multimorbid CAD patients, new avenues with the potential for more focused targeting of TGF β in SMCs have recently opened up. In the last decade, next-generation sequencing studies have identified non-coding RNA (ncRNA) sequences residing in intergenic regions of the genome. These non-coding transcripts are now known to have multiple functions, regulating the transcription and translation of proximal and distant protein-coding genes in a context-specific manner (reviewed in [172]). Recent studies have begun to elucidate the interactions between them and TGF β pathway components, identifying novel potential therapeutic targets for CAD. Early studies showed that TGF β could alter the expression of numerous microRNAs (miRs) in various human tissues and cells, the effects of which appear to be cell-type specific [173]. Microarray analysis in human carotid artery SMCs revealed a number of differentially expressed miRs following TGF β 1 treatment, including miR-143/145, which was significantly up-regulated by TGF β 1 in a concentration- and time-dependent manner [174]. Treatment of SMC with a specific inhibitor of p38MAPK completely blocked TGF β 1-induced miR-143/145 expression and attenuated the expression of SMC contractile genes (including CNN1, TAGLN and ACTA2) in response to TGF β 1 stimulation [174], identifying an additional mechanism through which TGF β 1 can promote SMC differentiation. Interestingly the miR-143/145 cluster, which is highly enriched in SMCs, has been shown to be significantly decreased following acute arterial injury [175] and in mouse atherosclerotic lesions [175]. Genetic knockout of miR-143/145 led to a reduction in the number of contractile arterial SMCs and a corresponding increase in synthetic SMCs, as determined by electron microscopy [176]. Neointimal lesions were also frequently observed in the femoral arteries of aged miR-143/145^{-/-} mice, with no lesions observed in wild-type animals [176]. TGF β has also been shown to regulate the expression of miR-21 through promoting the processing of pri-miR-21 into pre-miR-21 by the Drosha complex [177]. Importantly, miR-21 is over-expressed in murine and porcine models of vein grafting and is highly expressed within α SMA⁺ SMCs of failed human vein grafts [178]. Genetic ablation or antisense oligonucleotide-mediated knock-down of miR-21 significantly attenuated injury-induced neointima formation by inhibiting SMC proliferation and migration and inducing SMC apoptosis, highlighting the potential therapeutic benefit of miR-21 inhibition [178,179]. Together, these studies indicate that TGF β -regulated miRNAs play a critical role in controlling SMC phenotype transitions and the response of the vascular wall to injury, underlining their potential as therapeutic targets. Targeting SMC-enriched, disease-dysregulated miRs downstream of TGF β may be a more rational approach for achieving therapeutic efficacy whilst avoiding undesired side-effects.

10. Concluding remarks

TGF β was initially identified in the early 1980's, when Anita

Roberts and Michael Sporn purified a 'transformation factor' that could render healthy cells malignant [180]. The first observation that this Janus-like cytokine could have multifunctional effects was made shortly thereafter, in studies showing that TGF β could synergise with PDGF to stimulate fibroblast colony formation (CF) whilst inhibiting epidermal growth factor-induced CF [181]. From these early beginnings, the field of TGF β research – and indeed the TGF β superfamily – has expanded exponentially, with papers on TGF β now numbering in the tens of thousands. Nevertheless, important questions have yet to be fully answered, and our understanding of the many TGF β paradoxes remains incomplete. The advent of next-generation sequencing (NGS) has provided some clarification, identifying hitherto unknown genetic and phenotypic overlaps between patients who develop cardiovascular disease and those with inherited vascular conditions caused by mutations in TGF β genes. Alongside, investigations following on from the Human Genome Project have started unravelling the complexity of the transcriptome, identifying non-coding RNA sequences that both regulate and are regulated by TGF β signalling. These and other studies have greatly enhanced our mechanistic understanding of TGF β , and the many levels at which this pleiotropic cytokine is controlled. From early experiments showing that TGF β enhances the secretion of ECM proteins, we are now beginning to grasp how the cellular microenvironment in turn influences the actions of TGF β ; this is of particular relevance to coronary artery disease and intimal hyperplasia, during which extensive vascular remodelling occurs. Elucidation of these and other questions regarding the actions and interactions of TGF β will, we hope, lead to the development of localised and pathway-specific therapies that effectively and selectively target the pathological actions of TGF β .

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References

- [1] S.M. Schwartz, M.B. Stemerman, E.P. Benditt, The aortic intima. II. Repair of the aortic lining after mechanical denudation, *Am. J. Pathol.* 81 (1) (1975) 15–42.
- [2] A.W. Clowes, M.A. Reidy, M.M. Clowes, Kinetics of cellular proliferation after arterial injury. I. Smooth muscle growth in the absence of endothelium, *Lab. Invest.* 49 (3) (1983) 327–333.
- [3] J.A. Vita, C.B. Treasure, E.G. Nabel, J.M. McLenachan, R.D. Fish, A.C. Yeung, et al., Coronary vasomotor response to acetylcholine relates to risk factors for coronary artery disease, *Circulation* 81 (2) (1990) 491–497.
- [4] S. John, M. Schlaich, M. Langenfeld, H. Weihprecht, G. Schmitz, G. Weidinger, et al., Increased bioavailability of nitric oxide after lipid-lowering therapy in hypercholesterolemic patients: a randomized, placebo-controlled, double-blind study, *Circulation* 98 (3) (1998) 211–216.
- [5] V. Schachinger, M.B. Britten, M. Elsner, D.H. Walter, I. Scharrer, A.M. Zeiher, A positive family history of premature coronary artery disease is associated with impaired endothelium-dependent coronary blood flow regulation, *Circulation* 100 (14) (1999) 1502–1508.
- [6] E.M. Ruzicido, K.A. Martin, R.J. Powell, Regulation of vascular smooth muscle cell differentiation, *J. Vasc. Surg.* 45 (2007) Suppl A:A25–32.
- [7] M.R. de Vries, K.H. Simons, J.W. Jukema, J. Braun, P.H. Quax, Vein graft failure: from pathophysiology to clinical outcomes, *Nat. Rev. Cardiol.* 13 (8) (2016) 451–470.
- [8] P. ten Dijke, C.S. Hill, New insights into TGF-beta-Smad signalling, *Trends Biochem. Sci.* 29 (5) (2004) 265–273.
- [9] A.P. Hinck, Structural studies of the TGF-betas and their receptors - insights into evolution of the TGF-beta superfamily, *FEBS Lett.* 586 (14) (2012) 1860–1870.
- [10] A.P. Hinck, T. Huang, TGF-beta antagonists: same knot, but different hold, *Structure* 21 (8) (2013) 1269–1270.
- [11] R.J. Akhurst, S.A. Lehnert, A. Faissner, E. Duffie, TGF beta in murine morphogenetic processes: the early embryo and cardiogenesis, *Development* 108 (4) (1990) 645–656.
- [12] F.A. Millan, F. Denhez, P. Kondaiah, R.J. Akhurst, Embryonic gene expression patterns of TGF beta 1, beta 2 and beta 3 suggest different developmental functions in vivo, *Development* 111 (1) (1991) 131–143.

- [13] A. Bobik, A. Agrotis, P. Kanellakis, R. Dilley, A. Krushinsky, V. Smirnov, et al., Distinct patterns of transforming growth factor-beta isoform and receptor expression in human atherosclerotic lesions - Colocalization implicates TGF-beta in fibroblastic lesion development, *Circulation* 99 (22) (1999) 2883–2891.
- [14] P. ten Dijke, H.M. Arthur, Extracellular control of TGFbeta signalling in vascular development and disease, *Nat. Rev. Mol. Cell Biol.* 8 (11) (2007) 857–869.
- [15] D.B. Rifkin, Latent transforming growth factor-beta (TGF-beta) binding proteins: orchestrators of TGF-beta availability, *J. Biol. Chem.* 280 (9) (2005) 7409–7412.
- [16] J.S. Munger, J.G. Harpel, F.G. Giancotti, D.B. Rifkin, Interactions between growth factors and integrins: latent forms of transforming growth factor-beta are ligands for the integrin alphavbeta1, *Mol. Biol. Cell* 9 (9) (1998) 2627–2638.
- [17] J. Saharinen, M. Hyytiäinen, J. Taipale, J. Keski-Oja, Latent transforming growth factor-beta binding proteins (LTBPs) - structural extracellular matrix proteins for targeting TGF-beta action, *Cytokine Growth Factor Rev.* 10 (2) (1999) 99–117.
- [18] E. Piek, C.H. Heldin, P. Ten Dijke, Specificity, diversity, and regulation in TGF-beta superfamily signaling, *FASEB J.* 13 (15) (1999) 2105–2124.
- [19] S. Cheifetz, T. Bellon, C. Cales, S. Vera, C. Bernabeu, J. Massague, et al., Endoglin is a component of the transforming growth factor-beta receptor system in human endothelial cells, *J. Biol. Chem.* 267 (27) (1992) 19027–19030.
- [20] F. Lopez-Casillas, J.L. Wrana, J. Massague, Betaglycan presents ligand to the TGF beta signaling receptor, *Cell* 73 (7) (1993) 1435–1444.
- [21] M. Piao, O. Tokunaga, Significant expression of endoglin (CD105), TGFbeta-1 and TGFbeta R-2 in the atherosclerotic aorta: an immunohistological study, *J. Atheroscler. Thromb.* 13 (2) (2006) 82–89.
- [22] B. Gore, M. Izikki, O. Mercier, L. Dewachter, E. Fadel, M. Humbert, et al., Key role of the endothelial TGF-beta/ALK1/endoglin signaling pathway in humans and rodents pulmonary hypertension, *PLoS One* 9 (6) (2014) e100310.
- [23] M. Bilandzic, K.L. Stenvers, Reprint of: Betaglycan: a multifunctional accessory, *Mol. Cell. Endocrinol.* 359 (1–2) (2012) 13–22.
- [24] N.K. Kapur, K.J. Morine, M. Letarte, Endoglin: a critical mediator of cardiovascular health, *Vasc. Health Risk Manag.* 9 (2013) 195–206.
- [25] R. Derynck, Y.E. Zhang, Smad-dependent and Smad-independent pathways in TGF-beta family signalling, *Nature* 425 (6958) (2003) 577–584.
- [26] J. Massague, TGFbeta signalling in context, *Nat. Rev. Mol. Cell Biol.* 13 (10) (2012) 616–630.
- [27] T.D. Mueller, J. Nickel, Promiscuity and specificity in BMP receptor activation, *FEBS Lett.* 586 (14) (2012) 1846–1859.
- [28] S. Radaev, Z. Zou, T. Huang, E.M. Lafer, A.P. Hinck, P.D. Sun, Ternary complex of transforming growth factor-beta1 reveals isoform-specific ligand recognition and receptor recruitment in the superfamily, *J. Biol. Chem.* 285 (19) (2010) 14806–14814.
- [29] T.A. McCaffrey, S. Consigli, B. Du, D.J. Falcone, T.A. Sanborn, A.M. Spokojny, et al., Decreased type II/type I TGF-beta receptor ratio in cells derived from human atherosclerotic lesions. Conversion from an antiproliferative to profibrotic response to TGF-beta1, *J. Clin. Invest.* 96 (6) (1995) 2667–2675.
- [30] B. Schmierer, C.S. Hill, TGFbeta-SMAD signal transduction: molecular specificity and functional flexibility, *Nat. Rev. Mol. Cell Biol.* 8 (12) (2007) 970–982.
- [31] M.J. Goumans, G. Valdimarsdottir, S. Itoh, A. Rosendahl, P. Sideras, P. ten Dijke, Balancing the activation state of the endothelium via two distinct TGF-beta type I receptors, *EMBO J.* 21 (7) (2002) 1743–1753.
- [32] M.J. Goumans, G. Valdimarsdottir, S. Itoh, F. Lebrin, J. Larsson, C. Mummery, et al., Activin receptor-like kinase (ALK)1 is an antagonistic mediator of lateral TGFbeta/ALK5 signaling, *Mol. Cell* 12 (4) (2003) 817–828.
- [33] E.N. Blaney Davidson, D.F. Remst, E.L. Vitters, H.M. van Beuningen, A.B. Blom, M.J. Goumans, et al., Increase in ALK1/ALK5 ratio as a cause for elevated MMP-13 expression in osteoarthritis in humans and mice, *J. Immunol.* 182 (12) (2009) 7937–7945.
- [34] F. Curado, P. Spuul, I. Egana, P. Rottiers, T. Daubon, V. Veillot, et al., ALK5 and ALK1 play antagonistic roles in transforming growth factor beta-induced podosome formation in aortic endothelial cells, *Mol. Cell. Biol.* 34 (24) (2014) 4389–4403.
- [35] K.W. Finson, W.L. Parker, P. ten Dijke, M. Thorikay, A. Philip, ALK1 opposes ALK5/Smad3 signaling and expression of extracellular matrix components in human chondrocytes, *J. Bone Miner. Res.* 23 (6) (2008) 896–906.
- [36] E.M. Garrido-Martin, F.J. Blanco, M. Roque, L. Novensa, M. Tarocchi, U.E. Lang, et al., Vascular injury triggers Kruppel-like factor 6 mobilization and cooperation with specificity protein 1 to promote endothelial activation through upregulation of the activin receptor-like kinase 1 gene, *Circ. Res.* 112 (1) (2013) 113–127.
- [37] Y. Yao, B.J. Bennett, X. Wang, M.E. Rosenfeld, C. Giachelli, A.J. Lusis, et al., Inhibition of bone morphogenetic proteins protects against atherosclerosis and vascular calcification, *Circ. Res.* 107 (4) (2010) 485–494.
- [38] T. Seki, K.H. Hong, S.P. Oh, Nonoverlapping expression patterns of ALK1 and ALK5 reveal distinct roles of each receptor in vascular development, *Lab. Invest.* 86 (2) (2006) 116–129.
- [39] A. Moustakas, C.H. Heldin, Non-Smad TGF-beta signals, *J. Cell Sci.* 118 (2005) 3573–3584 Pt 16.
- [40] Y.E. Zhang, Non-Smad pathways in TGF-beta signaling, *Cell Res.* 19 (1) (2009) 128–139.
- [41] J. Massague, J. Seoane, D. Wotton, Smad transcription factors, *Genes Dev.* 19 (23) (2005) 2783–2810.
- [42] L. Attisano, S.T. Lee-Hoeflich, The Smads, *Genome Biol.* 2 (8) (2001) (REVIEWS3010).
- [43] Y. Shi, J. Massague, Mechanisms of TGF-beta signaling from cell membrane to the nucleus, *Cell* 113 (6) (2003) 685–700.
- [44] J. Massague, Y.G. Chen, Controlling TGF-beta signaling, *Genes Dev.* 14 (6) (2000) 627–644.
- [45] J. Massague, How cells read TGF-beta signals, *Nat. Rev. Mol. Cell Biol.* 1 (3) (2000) 169–178.
- [46] B. Schmierer, C.S. Hill, Kinetic analysis of Smad nucleocytoplasmic shuttling reveals a mechanism for transforming growth factor beta-dependent nuclear accumulation of Smads, *Mol. Cell. Biol.* 25 (22) (2005) 9845–9858.
- [47] X.H. Feng, R. Derynck, A kinase subdomain of transforming growth factor-beta (TGF-beta) type I receptor determines the TGF-beta intracellular signaling specificity, *EMBO J.* 16 (13) (1997) 3912–3923.
- [48] Y.G. Chen, A. Hata, R.S. Lo, D. Wotton, Y. Shi, N. Pavletich, et al., Determinants of specificity in TGF-beta signal transduction, *Genes Dev.* 12 (14) (1998) 2144–2152.
- [49] T. Tsukazaki, T.A. Chiang, A.F. Davison, L. Attisano, J.L. Wrana, SARA, a FYVE domain protein that recruits Smad2 to the TGFbeta receptor, *Cell* 95 (6) (1998) 779–791.
- [50] Y. Shi, Y.F. Wang, L. Jayaraman, H. Yang, J. Massague, N.P. Pavletich, Crystal structure of a Smad MH1 domain bound to DNA: insights on DNA binding in TGF-beta signaling, *Cell* 94 (5) (1998) 585–594.
- [51] L. Zavel, J.L. Dai, P. Buckhaults, S. Zhou, K.W. Kinzler, B. Vogelstein, et al., Human Smad3 and Smad4 are sequence-specific transcription activators, *Mol. Cell* 1 (4) (1998) 611–617.
- [52] G. Nishimura, I. Manabe, K. Tsushima, K. Fujiu, Y. Oishi, Imai Y, et al. delta EF1 mediates TGF-beta signaling in vascular smooth muscle cell differentiation, *Dev. Cell* 11 (1) (2006) 93–104.
- [53] P. Qiu, R.P. Ritchie, Z. Fu, D. Cao, J. Cumming, J.M. Miano, et al., Myocardin enhances Smad3-mediated transforming growth factor-beta1 signaling in a CARg box-independent manner: Smad-binding element is an important cis element for SM22alpha transcription in vivo, *Circ. Res.* 97 (10) (2005) 983–991.
- [54] David CJ, Massague J. Contextual determinants of TGF beta action in development, immunity and cancer (vol 19, pg 419, 2018). *Nat. Rev. Mol. Cell Biol.* 2018;19(7):479-.
- [55] C.S. Hill, Transcriptional Control by the SMADs, *Csh Perspect Biol.* 8 (10) (2016).
- [56] P. Libby, Changing concepts of atherogenesis, *J. Intern. Med.* 247 (3) (2000) 349–358.
- [57] K.J. Williams, I. Tabas, The response-to-retention hypothesis of early atherogenesis, *Arterioscler. Thromb. Vasc. Biol.* 15 (5) (1995) 551–561.
- [58] K.J. Williams, I. Tabas, The response-to-retention hypothesis of atherogenesis re-inforced, *Curr. Opin. Lipidol.* 9 (5) (1998) 471–474.
- [59] I. Tabas, K.J. Williams, J. Boren, Subendothelial lipoprotein retention as the initiating process in atherosclerosis: update and therapeutic implications, *Circulation* 116 (16) (2007) 1832–1844.
- [60] H.O. Steinberg, B. Bayazeed, G. Hook, A. Johnson, J. Cronin, A.D. Baron, Endothelial dysfunction is associated with cholesterol levels in the high normal range in humans, *Circulation* 96 (10) (1997) 3287–3293.
- [61] P. Libby, P.M. Ridker, G.K. Hansson, Progress and challenges in translating the biology of atherosclerosis, *Nature* 473 (7347) (2011) 317–325.
- [62] R. Ross, Atherosclerosis—an inflammatory disease, *N. Engl. J. Med.* 340 (2) (1999) 115–126.
- [63] A.L. Durham, M.Y. Speer, M. Scatena, C.M. Giachelli, C.M. Shanahan, Role of smooth muscle cells in vascular calcification: implications in atherosclerosis and arterial stiffness, *Cardiovasc. Res.* 114 (4) (2018) 590–600.
- [64] F. Cambien, S. Ricard, A. Troesch, C. Mallet, L. Genereaux, A. Evans, et al., Polymorphisms of the transforming growth factor-beta 1 gene in relation to myocardial infarction and blood pressure. The Etude Cas-Témoin de l'Infarctus du Myocarde (ECTIM) Study, *Hypertension* 28 (5) (1996) 881–887.
- [65] W. Koch, P. Hoppmann, J.C. Mueller, A. Schomig, A. Kastrati, Association of transforming growth factor-beta1 gene polymorphisms with myocardial infarction in patients with angiographically proven coronary heart disease, *Arterioscler. Thromb. Vasc. Biol.* 26 (5) (2006) 1114–1119.
- [66] M.P. Sie, A.G. Uitterlinden, M.J. Bos, P.P. Arp, M.M. Breteler, P.J. Koudstaal, et al., TGF-beta 1 polymorphisms and risk of myocardial infarction and stroke: the Rotterdam Study, *Stroke* 37 (11) (2006) 2667–2671.
- [67] D.R. Morris, J.V. Moxon, E. Biros, S.M. Krishna, J. Golledge, Meta-analysis of the association between transforming growth factor-beta polymorphisms and complications of coronary heart disease, *PLoS One* 7 (5) (2012) e37878.
- [68] Y. Lu, J.M. Boer, R.M. Barsova, O. Favorova, A. Goel, M. Muller, et al., TGFbeta1 genetic polymorphisms and coronary heart disease risk: a meta-analysis, *BMC Med. Genet.* 13 (2012) 39.
- [69] Y.Y. Li, Y.H. Zhou, G. Gong, H.Y. Geng, X.X. Yang, TGF-beta1 Gene -509C/T Polymorphism and Coronary Artery Disease: An Updated Meta-Analysis Involving 11,701 Subjects, *Front. Physiol.* 8 (2017) 108.
- [70] N. Verweij, R.N. Eppinga, Y. Hagemeijer, P. van der Harst, Identification of 15 novel risk loci for coronary artery disease and genetic risk of recurrent events, atrial fibrillation and heart failure, *Sci. Rep.* 7 (1) (2017) 2761.
- [71] C. Coronary Artery Disease, N.J. Samani, P. Deloukas, J. Erdmann, C. Hengstenberg, K. Kuulusmaa, et al., Large scale association analysis of novel genetic loci for coronary artery disease, *Arterioscler. Thromb. Vasc. Biol.* 29 (5) (2009) 774–780.
- [72] M. Nikpay, A. Goel, H.H. Won, L.M. Hall, C. Willenborg, S. Kanoni, et al., A comprehensive 1,000 Genomes-based genome-wide association meta-analysis of coronary artery disease, *Nat. Genet.* 47 (10) (2015) 1121–1130.
- [73] A.W. Turner, A. Martinuk, A. Silva, P. Lau, M. Nikpay, P. Eriksson, et al., Functional Analysis of a Novel Genome-Wide Association Study Signal in SMAD3 That Confers Protection From Coronary Artery Disease, *Arterioscler. Thromb. Vasc. Biol.* 36 (5) (2016) 972–983.
- [74] D.J. Grainger, P.R. Kemp, J.C. Metcalfe, A.C. Liu, R.M. Lawn, N.R. Williams, et al., The serum concentration of active transforming growth factor-beta is severely depressed in advanced atherosclerosis, *Nat. Med.* 1 (1) (1995) 74–79.

- [75] M. Erren, H. Reinecke, R. Junker, M. Fobker, H. Schulte, J.O. Schurek, et al., Systemic inflammatory parameters in patients with atherosclerosis of the coronary and peripheral arteries, *Arterioscler. Thromb. Vasc. Biol.* 19 (10) (1999) 2355–2363.
- [76] X.L. Wang, S.X. Liu, D.E. Wilcken, Circulating transforming growth factor beta 1 and coronary artery disease, *Cardiovasc. Res.* 34 (2) (1997) 404–410.
- [77] D.J. Grainger, D.E. Mosedale, J.C. Metcalfe, TGF-beta in blood: a complex problem, *Cytokine Growth Factor Rev.* 11 (1–2) (2000) 133–145.
- [78] R.A. van Dijk, C.C. Engels, A.F. Schaapherder, A. Mulder-Stapel, P. Ten Dijke, J.F. Hamming, et al., Visualizing TGF-beta and BMP signaling in human atherosclerosis: a histological evaluation based on Smad activation, *Histol. Histopathol.* 27 (3) (2012) 387–396.
- [79] D.J. Grainger, Transforming growth factor beta and atherosclerosis: so far, so good for the protective cytokine hypothesis, *Arterioscler. Thromb. Vasc. Biol.* 24 (3) (2004) 399–404.
- [80] D.J. Grainger, D.E. Mosedale, J.C. Metcalfe, E.P. Bottinger, Dietary fat and reduced levels of TGFbeta1 act synergistically to promote activation of the vascular endothelium and formation of lipid lesions, *J. Cell Sci.* 113 (2000) 2355–2361 Pt 13.
- [81] Z. Mallat, A. Gojova, C. Marchiol-Fournigault, B. Esposito, C. Kamate, R. Merval, et al., Inhibition of transforming growth factor-beta signaling accelerates atherosclerosis and induces an unstable plaque phenotype in mice, *Circ. Res.* 89 (10) (2001) 930–934.
- [82] E. Lutgens, M. Gijbels, M. Smook, P. Heeringa, P. Gotwals, V.E. Kotliansky, et al., Transforming growth factor-beta mediates balance between inflammation and fibrosis during plaque progression, *Arterioscler. Thromb. Vasc. Biol.* 22 (6) (2002) 975–982.
- [83] D. Li, Y. Liu, J. Chen, N. Velchala, F. Amani, A. Nemarkommula, et al., Suppression of atherogenesis by delivery of TGFbeta1ACT using adeno-associated virus type 2 in LDLR knockout mice, *Biochem. Biophys. Res. Commun.* 344 (3) (2006) 701–707.
- [84] A.D. Frutkin, G. Otsuka, A. Stempien-Otero, C. Sesti, L. Du, M. Jaffe, et al., TGF-beta 1 Limits Plaque Growth, Stabilizes Plaque Structure, and Prevents Aortic Dilation in Apolipoprotein E-Null Mice, *Arterioscler. Thromb. Vasc. Biol.* 29 (9) (2009) 1251–U41.
- [85] J. Rodriguez-Vita, E. Sanchez-Galan, B. Santamaria, E. Sanchez-Lopez, R. Rodriguez-Diez, L.M. Blanco-Colio, et al., Essential role of TGF-beta/Smad pathway on statin dependent vascular smooth muscle cell regulation, *PLoS One* 3 (12) (2008) e3959.
- [86] D.J. Grainger, TGF-beta and atherosclerosis in man, *Cardiovasc. Res.* 74 (2) (2007) 213–222.
- [87] S.M. Evrard, L. Lecce, K.C. Michelis, A. Nomura-Kitabayashi, G. Pandey, K.R. Purushothaman, et al., Endothelial to mesenchymal transition is common in atherosclerotic lesions and is associated with plaque instability, *Nat. Commun.* 7 (2016) 11853.
- [88] V. Cavieres, K. Valdes, B. Moreno, R. Moore-Carrasco, D.R. Gonzalez, Vascular hypercontractility and endothelial dysfunction before development of atherosclerosis in moderate dyslipidemia: role for nitric oxide and interleukin-6, *Am J Cardiovasc Dis.* 4 (3) (2014) 114–122.
- [89] E. Schonherr, H.T. Jarvelainen, M.G. Kinsella, L.J. Sandell, T.N. Wight, Platelet-derived growth factor and transforming growth factor-beta 1 differentially affect the synthesis of biglycan and decorin by monkey arterial smooth muscle cells, *Arterioscler. Thromb.* 13 (7) (1993) 1026–1036.
- [90] P.J. Little, L. Tannock, K.L. Olin, A. Chait, T.N. Wight, Proteoglycans synthesized by arterial smooth muscle cells in the presence of transforming growth factor-beta 1 exhibit increased binding to LDLs, *Arterioscler. Thromb. Vasc. Biol.* 22 (1) (2002) 55–60.
- [91] V.M. Kahari, H. Larjava, J. Uitto, Differential regulation of extracellular matrix proteoglycan (PG) gene expression. Transforming growth factor-beta 1 up-regulates biglycan (PGI), and versican (large fibroblast PG) but down-regulates decorin (PGII) mRNA levels in human fibroblasts in culture, *J. Biol. Chem.* 266 (16) (1991) 10608–10615.
- [92] T.N. Wight, A role for proteoglycans in vascular disease, *Matrix Biol.* 71–72 (2018) 396–420.
- [93] T. Tang, P.G. Wilson, J.C. Thompson, C. Nelson, M.H. Yoder, L.R. Tannock, Prevention of TGFbeta induction attenuates angII-stimulated vascular biglycan and atherosclerosis in *Ldlr*^{-/-} mice, *J. Lipid Res.* 54 (8) (2013) 2255–2264.
- [94] V.J. Thannickal, D.Y. Lee, E.S. White, Z. Cui, J.M. Larios, R. Chacon, et al., Myofibroblast differentiation by transforming growth factor-beta1 is dependent on cell adhesion and integrin signaling via focal adhesion kinase, *J. Biol. Chem.* 278 (14) (2003) 12384–12389.
- [95] P.J. Gao, Y. Li, A.J. Sun, J.J. Liu, K.D. Ji, Y.Z. Zhang, et al., Differentiation of vascular myofibroblasts induced by transforming growth factor-beta1 requires the involvement of protein kinase Calpha, *J. Mol. Cell. Cardiol.* 35 (9) (2003) 1105–1112.
- [96] A.H. Schulick, A.J. Taylor, W. Zuo, C.B. Qiu, G. Dong, R.N. Woodward, et al., Overexpression of transforming growth factor beta1 in arterial endothelium causes hyperplasia, apoptosis, and cartilaginous metaplasia, *Proc. Natl. Acad. Sci. U. S. A.* 95 (12) (1998) 6983–6988.
- [97] S. Tsai, S.T. Hollenbeck, E.J. Ryer, R. Edlin, D. Yamanouchi, R. Kundi, et al., TGF-beta through Smad3 signaling stimulates vascular smooth muscle cell proliferation and neointimal formation, *Am. J. Physiol. Heart Circ. Physiol.* 297 (2) (2009) H540–H549.
- [98] R.A. Nemenoff, H. Horita, A.C. Ostriker, S.B. Furgeson, P.A. Simpson, V. VanPutten, et al., SDF-1alpha induction in mature smooth muscle cells by inactivation of PTEN is a critical mediator of exacerbated injury-induced neointima formation, *Arterioscler. Thromb. Vasc. Biol.* 31 (6) (2011) 1300–1308.
- [99] J. Chappell, J.L. Harman, V.M. Narasimhan, H. Yu, K. Foote, B.D. Simons, et al., Extensive Proliferation of a Subset of Differentiated, yet Plastic, Medial Vascular Smooth Muscle Cells Contributes to Neointimal Formation in Mouse Injury and Atherosclerosis Models, *Circ. Res.* 119 (12) (2016) 1313–1323.
- [100] A.E. Walts, M.C. Fishbein, J.M. Matloff, Thrombosed, ruptured atheromatous plaques in saphenous vein coronary artery bypass grafts: ten years' experience, *Am. Heart J.* 114 (4 Pt 1) (1987) 718–723.
- [101] R.D. Safian, Accelerated atherosclerosis in saphenous vein bypass grafts: a spectrum of diffuse plaque instability, *Prog. Cardiovasc. Dis.* 44 (6) (2002) 437–448.
- [102] K. Yahagi, F.D. Kolodgie, F. Otsuka, A.V. Finn, H.R. Davis, M. Joner, et al., Pathophysiology of native coronary, vein graft, and in-stent atherosclerosis, *Nat. Rev. Cardiol.* 13 (2) (2016) 79–98.
- [103] T. Higo, Y. Ueda, J. Oyabu, K. Okada, M. Nishio, A. Hirata, et al., Atherosclerotic and thrombotic neointima formed over sirolimus drug-eluting stent: an angiographic study, *JACC Cardiovasc. Imaging* 2 (5) (2009) 616–624.
- [104] G. Nakazawa, A.V. Finn, M. Vorpahl, E.R. Ladich, F.D. Kolodgie, R. Virmani, Coronary responses and differential mechanisms of late stent thrombosis attributed to first-generation sirolimus- and paclitaxel-eluting stents, *J. Am. Coll. Cardiol.* 57 (4) (2011) 390–398.
- [105] F. Otsuka, R.A. Byrne, K. Yahagi, H. Mori, E. Ladich, D.R. Fowler, et al., Neoatherosclerosis: overview of histopathologic findings and implications for intravascular imaging assessment, *Eur. Heart J.* 36 (32) (2015) 2147–2159.
- [106] D. Nakamura, G.F. Attizzani, C. Toma, T. Sheth, W. Wang, M. Soud, et al., Failure Mechanisms and Neoatherosclerosis Patterns in Very Late Drug-Eluting and Bare-Metal Stent Thrombosis, *Circ Cardiovasc Interv.* 9 (9) (2016).
- [107] S. Nikol, J.M. Isner, J.G. Pickering, M. Kearney, G. Leclerc, L. Weir, Expression of transforming growth factor-beta 1 is increased in human vascular restenosis lesions, *J. Clin. Invest.* 90 (4) (1992) 1582–1592.
- [108] C. Yutani, M. Imakita, H. Ishibashi-Ueda, Y. Tsukamoto, N. Nishida, Y. Ikeda, Coronary atherosclerosis and interventions: pathological sequences and restenosis, *Pathol. Int.* 49 (4) (1999) 273–290.
- [109] I.M. Chung, H.K. Gold, S.M. Schwartz, Y. Ikari, M.A. Reidy, T.N. Wight, Enhanced extracellular matrix accumulation in restenosis of coronary arteries after stent deployment, *J. Am. Coll. Cardiol.* 40 (12) (2002) 2072–2081.
- [110] J.A. Madri, M.A. Reidy, O. Koher, L. Bell, Endothelial cell behavior after denudation injury is modulated by transforming growth factor-beta1 and fibronectin, *Lab. Invest.* 60 (6) (1989) 755–765.
- [111] M.W. Majesky, V. Lindner, D.R. Twardzik, S.M. Schwartz, M.A. Reidy, Production of transforming growth factor beta 1 during repair of arterial injury, *J. Clin. Invest.* 88 (3) (1991) 904–910.
- [112] J. Chamberlain, J. Gunn, S.E. Francis, C.M. Holt, N.D. Arnold, D.C. Cumberland, et al., TGF beta is active, and correlates with activators of TGF beta, following porcine coronary angioplasty, *Cardiovasc. Res.* 50 (1) (2001) 125–136.
- [113] Z. Jiang, M. Tao, K.A. Omalley, D. Wang, C.K. Ozaki, S.A. Berceci, Established neointimal hyperplasia in vein grafts expands via TGF-beta-mediated progressive fibrosis, *Am. J. Physiol. Heart Circ. Physiol.* 297 (4) (2009) H1200–H1207.
- [114] E.G. Nabel, L. Shum, V.J. Pompili, Z.Y. Yang, H. San, H.B. Shu, et al., Direct transfer of transforming growth factor beta 1 gene into arteries stimulates fibrocellular hyperplasia, *Proc. Natl. Acad. Sci. U. S. A.* 90 (22) (1993) 10759–10763.
- [115] R.A. Wolff, R.L. Malinowski, N.S. Heaton, D.A. Hullett, J.R. Hoch, Transforming growth factor-beta1 antisense treatment of rat vein grafts reduces the accumulation of collagen and increases the accumulation of h-caldesmon, *J. Vasc. Surg.* 43 (5) (2006) 1028–1036.
- [116] J.D. Smith, S.R. Bryant, L.L. Couper, C.P. Vary, P.J. Gotwals, V.E. Kotliansky, et al., Soluble transforming growth factor-beta type II receptor inhibits negative remodeling, fibroblast transdifferentiation, and intimal lesion formation but not endothelial growth, *Circ. Res.* 84 (10) (1999) 1212–1222.
- [117] K. Yamamoto, R. Morishita, N. Tomita, T. Shimozato, H. Nakagami, A. Kikuchi, et al., Ribozyme oligonucleotides against transforming growth factor-beta inhibited neointimal formation after vascular injury in rat model: potential application of ribozyme strategy to treat cardiovascular disease, *Circulation* 102 (11) (2000) 1308–1314.
- [118] P.A. Kingston, S. Sinha, C.E. Appleby, A. David, T. Verakis, M.G. Castro, et al., Adenovirus-mediated gene transfer of transforming growth factor-beta3, but not transforming growth factor-beta1, inhibits constrictive remodeling and reduces luminal loss after coronary angioplasty, *Circulation* 108 (22) (2003) 2819–2825.
- [119] J. Ghosh, M. Baguneid, N. Khwaja, M.O. Murphy, N. Turner, A. Halka, et al., Reduction of myointimal hyperplasia after arterial anastomosis by local injection of transforming growth factor beta3, *J. Vasc. Surg.* 43 (1) (2006) 142–149.
- [120] R. Kundi, S.T. Hollenbeck, D. Yamanouchi, B.C. Herman, R. Edlin, E.J. Ryer, et al., Arterial gene transfer of the TGF-beta signalling protein Smad3 induces adaptive remodelling following angioplasty: a role for CTGF, *Cardiovasc. Res.* 84 (2) (2009) 326–335.
- [121] C.M. Mallawaarachchi, P.L. Weissberg, R.C. Slow, Smad7 gene transfer attenuates arterial cell migration and vascular remodeling after balloon injury, *Arterioscler. Thromb. Vasc. Biol.* 25 (7) (2005) 1383–1387.
- [122] J. Rodriguez-Vita, E. Sanchez-Lopez, V. Esteban, M. Ruperez, J. Egido, M. Ruiz-Ortega, Angiotensin II activates the Smad pathway in vascular smooth muscle cells by a transforming growth factor-beta-independent mechanism, *Circulation* 111 (19) (2005) 2509–2517.
- [123] J.H. Li, X.R. Huang, H.J. Zhu, R. Johnson, H.Y. Lan, Role of TGF-beta signaling in extracellular matrix production under high glucose conditions, *Kidney Int.* 63 (6) (2003) 2010–2019.
- [124] K. Fu, M.J. Corbley, L. Sun, J.E. Friedman, F. Shan, J.L. Papadatos, et al., SM16, an orally active TGF-beta type I receptor inhibitor prevents myofibroblast induction

- and vascular fibrosis in the rat carotid injury model, *Arterioscler. Thromb. Vasc. Biol.* 28 (4) (2008) 665–671.
- [125] X. Guo, S.Y. Chen, Transforming growth factor-beta and smooth muscle differentiation, *World J. Biol. Chem.* 3 (3) (2012) 41–52.
- [126] S. Chen, M. Kulik, R.J. Lechleider, Smad proteins regulate transcriptional induction of the SM22alpha gene by TGF-beta, *Nucleic Acids Res.* 31 (4) (2003) 1302–1310.
- [127] B. Hu, Z. Wu, S.H. Phan, Smad3 mediates transforming growth factor-beta-induced alpha-smooth muscle actin expression, *Am. J. Respir. Cell Mol. Biol.* 29 (3) (2003) 397–404 Pt 1.
- [128] K.K. Hirschi, L. Lai, N.S. Belaguli, D.A. Dean, R.J. Schwartz, W.E. Zimmer, Transforming growth factor-beta induction of smooth muscle cell phenotype requires transcriptional and post-transcriptional control of serum response factor, *J. Biol. Chem.* 277 (8) (2002) 6287–6295.
- [129] P. Qiu, X.H. Feng, L. Li, Interaction of Smad3 and SRF-associated complex mediates TGF-beta1 signals to regulate SM22 transcription during myofibroblast differentiation, *J. Mol. Cell. Cardiol.* 35 (12) (2003) 1407–1420.
- [130] K. Kubota, J. Okazaki, O. Louie, K.C. Kent, B. Liu, TGF-beta stimulates collagen (I) in vascular smooth muscle cells via a short element in the proximal collagen promoter, *J. Surg. Res.* 109 (1) (2003) 43–50.
- [131] F. Verrecchia, A. Mauviel, Transforming growth factor-beta signaling through the Smad pathway: role in extracellular matrix gene expression and regulation, *J. Invest. Dermatol.* 118 (2) (2002) 211–215.
- [132] A. Leask, D.J. Abraham, TGF-beta signaling and the fibrotic response, *FASEB J.* 18 (7) (2004) 816–827.
- [133] Q.J. Zhang, M. Goddard, C. Shanahan, L. Shapiro, M. Bennett, Differential gene expression in vascular smooth muscle cells in primary atherosclerosis and in stent stenosis in humans, *Arterioscler. Thromb. Vasc. Biol.* 22 (12) (2002) 2030–2036.
- [134] R. Khan, A. Agrotis, A. Bobik, Understanding the role of transforming growth factor-beta1 in intimal thickening after vascular injury, *Cardiovasc. Res.* 74 (2) (2007) 223–234.
- [135] R.A. Majack, Beta-type transforming growth factor specifies organizational behavior in vascular smooth muscle cell cultures, *J. Cell Biol.* 105 (1) (1987) 465–471.
- [136] D.L. Hwang, L.J. Latus, A. Lev-Ran, Effects of platelet-contained growth factors (PDGF, EGF, IGF-I, and TGF-beta) on DNA synthesis in porcine aortic smooth muscle cells in culture, *Exp. Cell Res.* 200 (2) (1992) 358–360.
- [137] G.K. Owens, A.A. Geisterfer, Y.W. Yang, A. Komoriya, Transforming growth factor-beta-induced growth inhibition and cellular hypertrophy in cultured vascular smooth muscle cells, *J. Cell Biol.* 107 (2) (1988) 771–780.
- [138] Majack RA, Majesky MW, Goodman LV. Role of PDGF-A expression in the control of vascular smooth muscle cell growth by transforming growth factor-beta. *J. Cell Biol.* 1990;111(1):239-47.
- [139] P.J. Little, T.J. Allen, K. Hashimura, J. Nigro, C.A. Farrelly, R.J. Dilley, High glucose potentiates mitogenic responses of cultured ovine coronary smooth muscle cells to platelet derived growth factor and transforming growth factor-beta1, *Diabetes Res. Clin. Pract.* 59 (2) (2003) 93–101.
- [140] K.B. Reddy, P.H. Howe, Transforming growth factor beta 1-mediated inhibition of smooth muscle cell proliferation is associated with a late G1 cell cycle arrest, *J. Cell. Physiol.* 156 (1) (1993) 48–55.
- [141] U. Seay, D. Sedding, S. Krick, M. Hecker, W. Seeger, O. Eickelberg, Transforming growth factor-beta-dependent growth inhibition in primary vascular smooth muscle cells is p38-dependent, *J. Pharmacol. Exp. Ther.* 315 (3) (2005) 1005–1012.
- [142] A. Martin-Garrido, H.C. Williams, M. Lee, B. Seidel-Rogol, X. Ci, J.T. Dong, et al., Transforming Growth Factor beta Inhibits Platelet Derived Growth Factor-Induced Vascular Smooth Muscle Cell Proliferation via Akt-Independent, Smad-Mediated Cyclin D1 Downregulation, *PLoS One* 8 (11) (2013).
- [143] E.A. Klein, R.K. Assoian, Transcriptional regulation of the cyclin D1 gene at a glance, *J. Cell Sci.* 121 (23) (2008) 3853–3857.
- [144] E.J. Battegay, E.W. Raines, R.A. Seifert, D.F. Bowen-Pope, R. Ross, TGF-beta induces bimodal proliferation of connective tissue cells via complex control of an autocrine PDGF loop, *Cell* 63 (3) (1990) 515–524.
- [145] G.A. Stouffer, G.K. Owens, Tgf-Beta Promotes Proliferation of Cultured Smc Via Both Pdgf-Aa-Dependent and Pdgf-Aa-Independent Mechanisms, *J. Am. Coll. Cardiol.* A162-A (1994).
- [146] X. Mao, P. DeBenedittis, Y. Sun, J.F. Chen, K.Y. Yuan, K. Jiao, et al., Vascular Smooth Muscle Cell Smad4 Gene Is Important for Mouse Vascular Development, *Arterioscler. Thromb. Vasc. Biol.* 32 (9) (2012) 2171.
- [147] P.A. Suwanabol, S.M. Seedial, X. Shi, F. Zhang, D. Yamanouchi, D. Roenneburg, et al., Transforming growth factor-beta increases vascular smooth muscle cell proliferation through the Smad3 and extracellular signal-regulated kinase mitogen-activated protein kinases pathways, *J. Vasc. Surg.* 56 (2) (2012) 446–454.
- [148] K. Kobayashi, K. Yokote, M. Fujimoto, K. Yamashita, A. Sakamoto, M. Kitahara, et al., Targeted disruption of TGF-beta-Smad3 signaling leads to enhanced neointimal hyperplasia with diminished matrix deposition in response to vascular injury, *Circ. Res.* 96 (8) (2005) 904–912.
- [149] S. Mii, J.A. Ware, K.C. Kent, Transforming growth factor-beta inhibits human vascular smooth muscle cell growth and migration, *Surgery* 114 (2) (1993) 464–470.
- [150] L. Engel, U. Ryan, TGF-beta 1 reverses PDGF-stimulated migration of human aortic smooth muscle cells in vitro, *In Vitro Cell Dev Biol Anim.* 33 (6) (1997) 443–451.
- [151] G.M. Risinger Jr., D.L. Updike, E.C. Bullen, J.J. Tomasek, E.W. Howard, TGF-beta suppresses the upregulation of MMP-2 by vascular smooth muscle cells in response to PDGF-BB, *Am. J. Phys. Cell Phys.* 298 (1) (2010) C191–C201.
- [152] C.T. Basson, O. Kocher, M.D. Basson, A. Asis, J.A. Madri, Differential modulation of vascular cell integrin and extracellular matrix expression in vitro by TGF-beta 1 correlates with reciprocal effects on cell migration, *J. Cell. Physiol.* 153 (1) (1992) 118–128.
- [153] M.F. Janat, W.S. Argraves, G. Liao, Regulation of vascular smooth muscle cell integrin expression by transforming growth factor beta1 and by platelet-derived growth factor-BB, *J. Cell. Physiol.* 151 (3) (1992) 588–595.
- [154] S.L. Brown, C.H. Lundgren, T. Nordt, S. Fujii, Stimulation of migration of human aortic smooth muscle cells by vitronectin: implications for atherosclerosis, *Cardiovasc. Res.* 28 (12) (1994) 1815–1820.
- [155] Srivatsa SS, Fitzpatrick LA, Tsao PW, Reilly TM, Holmes DR, Jr., Schwartz RS, et al. Selective alpha v beta 3 integrin blockade potentially limits neointimal hyperplasia and lumen stenosis following deep coronary arterial stent injury: evidence for the functional importance of integrin alpha v beta 3 and osteopontin expression during neointima formation. *Cardiovasc. Res.* 1997;36(3):408-28.
- [156] M.R. Ward, A. Agrotis, P. Kanellakis, R. Dilley, G. Jennings, A. Bobik, Inhibition of protein tyrosine kinases attenuates increases in expression of transforming growth factor-beta isoforms and their receptors following arterial injury, *Arterioscler. Thromb. Vasc. Biol.* 17 (11) (1997) 2461–2470.
- [157] A.M. Bertoli-Avella, E. Gillis, H. Morisaki, J.M.A. Verhagen, B.M. de Graaf, G. van de Beek, et al., Mutations in a TGF-beta Ligand, TGFβ3, Cause Syndromic Aortic Aneurysms and Dissections, *J. Am. Coll. Cardiol.* 65 (13) (2015) 1324–1336.
- [158] E. Pardali, M.J. Goumans, P. ten Dijke, Signaling by members of the TGF-beta family in vascular morphogenesis and disease, *Trends Cell Biol.* 20 (9) (2010) 556–567.
- [159] E. Pardali, P. Ten Dijke, TGFbeta signaling and cardiovascular diseases, *Int. J. Biol. Sci.* 8 (2) (2012) 195–213.
- [160] H. Tamai, O. Katoh, S. Suzuki, K. Fujii, T. Aizawa, S. Takase, et al., Impact of tranilast on restenosis after coronary angioplasty: tranilast restenosis following angioplasty trial (TREAT), *Am. Heart J.* 138 (5) (1999) 968–975 Pt 1.
- [161] H. Tamai, K. Katoh, T. Yamaguchi, H. Hayakawa, K. Kamatsuse, K. Haze, et al., The impact of tranilast on restenosis after coronary angioplasty: the Second Tranilast Restenosis Following Angioplasty Trial (TREAT-2), *Am. Heart J.* 143 (3) (2002) 506–513.
- [162] H. Azuma, K. Banno, T. Yoshimura, Pharmacological properties of N-(3',4'-dimethoxycinnamoyl) anthranilic acid (N-5'), a new anti-atopic agent, *Br. J. Pharmacol.* 58 (4) (1976) 483–488.
- [163] H. Suzuki, K. Tanaka, H. Kaneshige, K. Nakagami, E. Noguchi, The effects of long term Tranilast administration on bronchial hypersensitivity in asthmatics, *Panninerva Med.* 31 (2) (1989) 88–93.
- [164] L. Kohavi, E. Sprecher, E. Zur, O. Artzi, The Effect of Tranilast 8% Liposomal Gel Versus Placebo on Post-Cesarean Surgical Scars: A Prospective Double-Blind Split-Scar Study, *Dermatol. Surg.* 43 (9) (2017) 1157–1163.
- [165] D.R. Holmes Jr., M. Savage, J.M. LaBlanche, L. Grip, P.W. Serruys, P. Fitzgerald, et al., Results of Prevention of REStenosis with Tranilast and its Outcomes (PRESTO) trial, *Circulation* 106 (10) (2002) 1243–1250.
- [166] R.J. Aikhurst, Targeting TGF-beta Signaling for Therapeutic Gain, *Cold Spring Harb. Perspect. Biol.* 9 (10) (2017).
- [167] R.J. Aikhurst, A. Hata, Targeting the TGFbeta signalling pathway in disease, *Nat. Rev. Drug Discov.* 11 (10) (2012) 790–811.
- [168] T.E. King Jr., W.Z. Bradford, S. Castro-Bernardini, E.A. Fagan, I. Glaspol, M.K. Glassberg, et al., A phase 3 trial of pirfenidone in patients with idiopathic pulmonary fibrosis, *N. Engl. J. Med.* 370 (22) (2014) 2083–2092.
- [169] S. Herberitz, J.S. Sawyer, A.J. Stauber, I. Gueorguieva, K.E. Driscoll, S.T. Estrem, et al., Clinical development of galunisertib (LY2157299 monohydrate), a small molecule inhibitor of transforming growth factor-beta signaling pathway, *Drug Des Devel Ther.* 9 (2015) 4479–4499.
- [170] M.J. Anderton, H.R. Mellor, A. Bell, C. Sadler, M. Pass, S. Powell, et al., Induction of heart valve lesions by small-molecule ALK5 inhibitors, *Toxicol. Pathol.* 39 (6) (2011) 916–924.
- [171] J. Rodon, M.A. Carducci, J.M. Sepulveda-Sanchez, A. Azaro, E. Calvo, J. Seoane, et al., First-in-human dose study of the novel transforming growth factor-beta receptor I kinase inhibitor LY2157299 monohydrate in patients with advanced cancer and glioma, *Clin. Cancer Res.* 21 (3) (2015) 553–560.
- [172] M.D. Ballantyne, R.A. McDonald, A.H. Baker, lncRNA/MicroRNA interactions in the vasculature, *Clin. Pharmacol. Ther.* 99 (5) (2016) 494–501.
- [173] H. Butz, K. Racz, L. Hunyady, A. Patocs, Crosstalk between TGF-beta signaling and the microRNA machinery, *Trends Pharmacol. Sci.* 33 (7) (2012) 382–393.
- [174] X. Long, J.M. Miano, Transforming growth factor-beta1 (TGF-beta1) utilizes distinct pathways for the transcriptional activation of microRNA 143/145 in human coronary artery smooth muscle cells, *J. Biol. Chem.* 286 (34) (2011) 30119–30129.
- [175] K.R. Cordes, N.T. Sheehy, M.P. White, E.C. Berry, S.U. Morton, A.N. Muth, et al., miR-145 and miR-143 regulate smooth muscle cell fate and plasticity, *Nature* 460 (7256) (2009) 705–U80.
- [176] T. Boettger, N. Beetz, S. Kostin, J. Schneider, M. Kruger, L. Hein, et al., Acquisition of the contractile phenotype by murine arterial smooth muscle cells depends on the Mir143/145 gene cluster, *J. Clin. Invest.* 119 (9) (2009) 2634–2647.
- [177] B.N. Davis, A.C. Hilyard, G. Lagna, A. Hata, SMAD proteins control DROSHA-mediated microRNA maturation, *Nature* 454 (7200) (2008) 56–61.
- [178] R.A. McDonald, K.M. White, J. Wu, B.C. Cooley, K.E. Robertson, C.A. Halliday, et al., miRNA-21 is dysregulated in response to vein grafting in multiple models and genetic ablation in mice attenuates neointima formation, *Eur. Heart J.* 34 (22) (2013) 1636–1643.
- [179] R. Ji, Y. Cheng, J. Yue, J. Yang, X. Liu, H. Chen, et al., MicroRNA expression signature and antisense-mediated depletion reveal an essential role of MicroRNA in vascular neointimal lesion formation, *Circ. Res.* 100 (11) (2007) 1579–1588.
- [180] A.B. Roberts, M.A. Anzano, L.C. Lamb, J.M. Smith, C.A. Frollich, H. Marquardt,

- et al., Isolation from murine sarcoma cells of novel transforming growth factors potentiated by EGF, *Nature* 295 (5848) (1982) 417–419.
- [181] A.B. Roberts, M.A. Anzano, L.M. Wakefield, N.S. Roche, D.F. Stern, M.B. Sporn, Type beta transforming growth factor: a bifunctional regulator of cellular growth, *Proc. Natl. Acad. Sci. U. S. A.* 82 (1) (1985) 119–123.
- [182] W. Zhao, C. Wang, R. Liu, C. Wei, J. Duan, K. Liu, et al., Effect of TGF-beta1 on the Migration and Recruitment of Mesenchymal Stem Cells after Vascular Balloon Injury: Involvement of Matrix Metalloproteinase-14, *Sci. Rep.* 6 (2016) 21176.
- [183] M. Merrilees, B. Beaumont, L. Scott, V. Hermanutz, P. Fennessy, Effect of TGF-beta (1) antisense S-oligonucleotide on synthesis and accumulation of matrix proteoglycans in balloon catheter-injured neointima of rabbit carotid arteries, *J. Vasc. Res.* 37 (1) (2000) 50–60.
- [184] M.R. Ward, T. Sasahara, A. Agrotis, R.J. Dille, G.L. Jennings, A. Bobik, Inhibitory effects of tranilast on expression of transforming growth factor-beta isoforms and receptors in injured arteries, *Atherosclerosis* 137 (2) (1998) 267–275.
- [185] N. Ohashi, A. Matsumori, Y. Furukawa, K. Ono, M. Okada, A. Iwasaki, et al., Role of p38 mitogen-activated protein kinase in neointimal hyperplasia after vascular injury, *Arterioscler. Thromb. Vasc. Biol.* 20 (12) (2000) 2521–2526.
- [186] E.H. Yao, N. Fukuda, T. Ueno, H. Matsuda, H. Nagase, Y. Matsumoto, et al., A pyrrole-imidazole polyamide targeting transforming growth factor-beta1 inhibits restenosis and preserves endothelialization in the injured artery, *Cardiovasc. Res.* 81 (4) (2009) 797–804.