

Vennin, C., Rath, N., Pajic, M., Olson, M. F. and Timpson, P. (2017) Targeting ROCK activity to disrupt and prime pancreatic cancer for chemotherapy. *Small GTPases*, (doi:<u>10.1080/21541248.2017.1345712</u>)

This is the author's final accepted version.

There may be differences between this version and the published version. You are advised to consult the publisher's version if you wish to cite from it.

http://eprints.gla.ac.uk/149547/

Deposited on: 23 October 2017

Enlighten – Research publications by members of the University of Glasgow http://eprints.gla.ac.uk

1	Targeting ROCK activity to disrupt and prime pancreatic cancer for
2	chemotherapy.
3	
4	Claire Vennin <sup>1,2,*</sup> , Nicola Rath <sup>3*</sup> , Marina Pajic <sup>1,2</sup> , Michael F. Olson <sup>3,4#</sup> and Paul
5	Timpson <sup>1,2,#</sup>
6	
7	<sup>1</sup> The Garvan Institute of Medical Research & The Kinghorn Cancer Centre,
8	2010 Sydney Australia
9	<sup>2</sup> St Vincent's Clinical School, Faculty of Medicine, University of New South
10	Wales, 2010 Sydney Australia
11	<sup>3</sup> Cancer Research UK Beatson Institute, Glasgow G61 BD UK
12	<sup>4</sup> Institute of Cancer Sciences, University of Glasgow, Glasgow G12 8QQ UK
13	<sup>*</sup> authors contributed equally to this work
14	<sup>#</sup> authors for correspondence:
15	Prof. Michael F. Olson: m.olson@beatson.gla.ac.uk and
16	Dr. Paul Timpson: <u>p.timpson@garvan.org.au</u> .
17	
18	

### 19 Abstract

20 Pancreatic ductal adenocarcinoma (PDAC) is a devastating disease; the identification of novel targets and development of effective treatment 21 22 strategies are urgently needed to improve patient outcomes. Remodeling of 23 the pancreatic stroma occurs during PDAC development, which drives 24 disease progression and impairs responses to therapy. The actomyosin 25 regulatory ROCK1 and ROCK2 kinases govern cell motility and contractility, 26 and have been suggested to be potential targets for cancer therapy, 27 particularly to reduce the metastatic spread of tumor cells. However, ROCK 28 inhibitors are not currently used for cancer patient treatment, largely due to 29 the overwhelming challenge faced in the development of anti-metastatic 30 drugs, and a lack of clarity as to the cancer types most likely to benefit from 31 ROCK inhibitor therapy. In two recent publications, we discovered that 32 ROCK1 and ROCK2 expression were increased in PDAC, and that increased 33 ROCK activity was associated with reduced survival and PDAC progression 34 by enabling extracellular matrix (ECM) remodeling and invasive growth of 35 pancreatic cancer cells. We also used intravital imaging to optimize ROCK 36 inhibition using the pharmacological ROCK inhibitor fasudil (HA-1077), and 37 demonstrated that short-term ROCK targeting, or 'priming', improved 38 chemotherapy efficacy, disrupted cancer cell collective movement, and 39 impaired metastasis. This body of work strongly indicates that the use of 40 ROCK inhibitors in pancreatic cancer therapy as 'priming' agents warrants 41 further consideration, and provides insights as to how transient mechanical 42 manipulation, or fine-tuning the ECM, rather than chronic stromal ablation

43 might be beneficial for improving chemotherapeutic efficacy in the treatment44 of this deadly disease.

45

# 46 Introduction

47 Despite there being a number of new therapeutics that have been 48 developed for pancreatic cancer patient therapy, survival remains the lowest 49 of all solid cancers, with 5-year survival rate being less than 7% and a median survival of 6 months <sup>1</sup>. Despite pre-clinical efforts to develop new therapeutics 50 <sup>2</sup>, patient survival has not significantly improved over the last 4 decades, 51 52 which highlights not only the need to identify new targets, but also to develop 53 innovative treatment strategies to improve the outcomes of patients suffering from this disease. In addition, development of diagnostic tools, for example 54 based on detection of cancer-derived exosomes<sup>3</sup>, to enable early detection of 55 56 pancreatic cancer remains a critical challenge for this disease. Pancreatic 57 ductal adenocarcinoma (PDAC) is characterized by extensive remodeling of 58 the pancreatic stroma, with increased deposition and crosslinking of 59 extracellular matrix (ECM) components and poor vascularization compared to normal pancreas<sup>4, 5</sup>. Alterations of the biochemical and mechanical properties 60 61 of the ECM are known to influence cancer progression, invasion and responses to chemotherapy 6-9, however, recent studies assessing the 62 63 efficacy of ECM-based pancreatic cancer therapies, for example via inhibition 64 of Sonic Hedgehog signaling pathway, targeting of lysyl oxidase activity or inhibition of hyaluronic acid (HA), have yielded conflicting results <sup>4, 10-16</sup>. 65

66 Rho-associated protein kinases 1 and 2 (ROCK1 and ROCK2) are 67 master regulators of the actomyosin cytoskeleton and govern force

generation, cell invasion, proliferation and contractility <sup>17-19</sup>. Numerous studies 68 69 have established that ROCK inhibition disrupts tumor progression and metastasis in cell based and *in vivo* models of various solid cancers <sup>20-23</sup>. 70 However, to date no compounds have progressed into the clinic for cancer 71 72 The development of anti-metastatic therapy for several reasons. 73 chemotherapeutics for clinical use is very challenging due to the need to 74 detect a reduction in metastasis in patients over sustained periods (likely years) as a positive outcome <sup>24</sup>, in contrast to chemotherapeutics that induce 75 76 acute positive responses, such as tumor regression, which can be monitored in a clinical trial in a defined and relatively brief time period <sup>24</sup>. Furthermore, 77 78 the absence of correlations between defined genetic alterations, such as 79 ROCK1 or ROCK2 mutations, with ROCK inhibitor sensitivity means that 80 there is no simple genetic test for convenient patient stratification. As a result, ROCK inhibition has not been adopted as a cancer chemotherapy. In this 81 commentary, we describe our recent findings <sup>25, 26</sup> demonstrating that ROCK 82 83 activity promotes pancreatic cancer invasive growth via ECM remodeling. We 84 also highlight how transient ROCK inhibition, or mechanical 'priming' with the 85 pharmacological inhibitor fasudil affects tumor tissue tension, which in turn 86 improves chemotherapy efficacy in primary and secondary tumor sites, while also disrupting collective movement of metastatic cancer cells <sup>26</sup>. Lastly, we 87 88 discuss potential translation of our findings into the clinic for pancreatic cancer 89 therapy, where balancing cellular contractility via transient ROCK inhibition, 90 rather than long-term ablation of the matrix, enables re-establishment of the 91 normal mechanical features of the stroma.

92

### 93 **ROCK activity promotes PDAC progression.**

94 Genomic analyses have previously shown that the ROCK1 gene is amplified in 15% of pancreatic patient tumors <sup>27</sup>, however the role of ROCK-95 mediated actomyosin contractility in PDAC had not been clearly established. 96 97 To address this, we assessed ROCK expression in a patient tissue microarray (78 samples from patients with pancreatic cancers and 5 healthy human 98 99 pancreas) and in human TCGA datasets, and determined that ROCK1 and ROCK2 expression increase with tumor stage and grade <sup>25</sup>. In line with this, 100 101 genomic alterations or mRNA amplification of ROCK1 and/or ROCK2 were 102 found to be positively correlated with poorer survival, suggesting that ROCK signaling promotes pancreatic cancer progression <sup>25</sup>. 103

104 To further understand how ROCK influences the fate and behavior of 105 pancreatic cancer cells, Cre-recombinase was expressed from the pancreatic 106 epithelial selective Pdx1 promoter to induce pancreas-targeted recombination of LOX-STOP-LOX (LSL)-Kras<sup>G12D/+</sup> and LSL-Trp53<sup>R172H/+</sup> (KPC) alleles in 107 mice, which spontaneously develop PDAC that closely resembles human 108 pancreatic cancer <sup>28, 29</sup>. In addition, KPC mice were crossed with LSL-109 ROCK2:ER mice <sup>30</sup> to conditionally activate ROCK2 during PDAC 110 progression. This model closely recapitulates the genomic features of human 111 PDAC, where an initiating Kras<sup>G12D</sup> mutation is found in almost 90% of patient 112 tumors, while the p53<sup>R175H</sup> mutation is found in 50-75% of patient tumors <sup>31</sup>. 113 114 Consistent with the observed increased ROCK2 protein levels in advanced 115 PDAC stages, as well as the correlation between increased ROCK1 and 116 ROCK2 mRNA expression, along with a potentially activating truncation 117 mutation (I383F-frameshift deletion; TCGA-HZ-8005-01), with poor survival

118 from the TCGA human dataset, conditional ROCK2 activation was associated with reduced PDAC mouse survival. Conditional ROCK2 activation in non 119 metastatic PDAC cells isolated from genetically modified mice promoted 120 121 pancreatic cancer cell invasion into 3D collagen matrices (see schematic 122 representation of ROCK inhibition at the cellular level, Fig. 1A)<sup>25</sup>. 123 Interestingly, analyses of cell-ECM interactions using Second Harmonic 124 Generation (SHG) imaging, a label free imaging technique used to detect non-125 centrosymmetric entities such as crosslinked collagen fibers, or tannic acid-126 glutaraldehyde fixation of collagen fibers for transmission electron 127 microscopy, revealed that ROCK activation induced extensive remodeling of the collagen matrix surrounding invading cancer cells <sup>25</sup>. 128

129 While ROCK is well known to induce force generation via its action on 130 actomyosin structures <sup>19</sup>, ROCK signaling also induces gene transcription <sup>32</sup>. 131 To identify ROCK induced gene expression changes, we performed RNA sequencing and identified 285 genes that were consistently and significantly 132 133 found to be changed greater than twofold relative to control cells. 134 conditional ROCK activation increased expression of Interestingly. 135 metalloproteinases (MMP) Mmp10 and Mmp13, which was associated with 136 increased release of these MMPs into the surrounding environment (see 137 schematic representation of ROCK inhibition at the cellular level, Fig. 1A). 138 These results indicated that ROCK mediates collagen remodeling by 139 pancreatic cancer cells via transcription, synthesis and release of MMPs, in line with previous observations in melanoma cells <sup>33</sup>, and in pancreatic cancer 140 cells in which dasatinib-induced reduction of KPC cell migration was 141 correlated with reduced production of MMP2 and MMP9<sup>34</sup>. We also 142

143 determined that ROCK-mediated remodeling of the surrounding matrix 144 facilitated invasive growth of pancreatic cancer cells (see schematic representation of ROCK inhibition at the cellular and whole-body levels, Fig. 145 1A, B). These findings highlight the ability of cancer cells to adapt to the 146 147 mechanical environment and to remodel the ECM to support their aberrant growth. These cell-based observations were further extended in KPC mice, 148 149 where ROCK inhibition with fasudil significantly prolonged survival, and 150 reduced collagen remodeling (see schematic representation of ROCK inhibition at the cellular and whole-body levels Fig. 1A, B)<sup>25</sup>. Together, these 151 152 results shed light on novel roles of ROCK in driving pancreatic cancer 153 progression, suggesting that targeting ROCK might be beneficial for the 154 clinical management of the disease.

155

# 156 **Transient ROCK inhibition with fasudil disrupts pancreatic cancer.**

157 Although ROCK-driven cell contractility and stromal remodeling are known to play crucial roles in cancer progression <sup>7, 19, 35</sup>, ROCK inhibitors and 158 159 ECM-based therapies have yet to be translated to the clinic. In our recent 160 publication, we assessed the efficacy of fasudil to impair PDAC progression and to influence cell responses to chemotherapy <sup>26</sup>. Fasudil is a ROCK 161 inhibitor currently used clinically as a monotherapy for the treatment of 162 cerebral vasospasm <sup>36</sup>, and Fasudil has also been shown to inhibit, in a less 163 164 potent manner than for ROCK, other kinases such as PKA, PKC and MLCK <sup>37</sup>. Meta-analysis of post-marketing surveillance data (>3,000 patients) has 165 demonstrated the safety of fasudil for clinical use in humans <sup>38</sup>, which 166 167 prompted us to assess the repurposing of fasudil for the treatment of

168 pancreatic cancer. We combined mouse and stratified patient-derived models 169 of pancreatic cancer with biosensor FLIM-FRET intravital imaging to monitor the effect of ROCK inhibition in real-time and in live tissues <sup>39-42</sup>. Using an 170 early, transient 'priming' regimen, where fasudil was administered for 3 days 171 172 prior to chemotherapy, in line with its treatment regimen in patients with stable angina <sup>43</sup>, we demonstrated that short-term ROCK inhibition with fasudil 173 174 synchronized pancreatic cancer cell cycle progression, and rendered them 175 more sensitive to subsequent treatment with anti-microtubule drugs and 176 standard-of-care chemotherapy, both in primary tumors and metastatic sites 177 (see schematic representation of ROCK inhibition at the whole-body level, Fig. 1B) <sup>26</sup>. We also observed that 'priming' with fasudil in the adjuvant setting 178 179 disturbed coordinated cancer cell movement and impaired metastatic 180 colonization in the liver (see schematic representation of ROCK inhibition at the whole-body level, Fig. 1B). 181

182

183 Assessment of the effect of 'priming' on key metastatic events revealed 184 that ROCK inhibition rendered circulating tumor cells more sensitive to shear 185 stress to which they are subjected in the blood circulation and in turn impaired 186 their ability to extravasate and colonize host tissues (see schematic 187 representation of ROCK inhibition at the whole-body level, Fig. 1B), consistent with previous studies <sup>44, 45</sup>. Additionally, analysis of collective cell movement, 188 189 or streaming, upon 'priming' suggested that transient ROCK inhibition 190 impaired coordinated cell migration and 3D cell movement of the metastatic 191 emboli in the liver (see schematic representation of ROCK inhibition at the whole-body level, Fig. 1B) <sup>26</sup>, possibly due to disrupted durotaxis - where cell 192

movement is directed by stiffness gradients - in the metastatic niche <sup>46</sup>. The 193 194 observed reduction of coordinated PDAC cell spread that we observed upon ROCK inhibition was also in line with previous work highlighting how the Rho-195 ROCK-LIMK pathway leads tumor cell invasion by driving path generation <sup>47</sup>. 196 197 ROCK inhibition was also found to reduce the ability of metastatic cells to 198 remodel the host ECM and to create a favorable environment to support their 199 growth in a distant site (see schematic representation of ROCK inhibition at 200 the whole-body level Fig. 1B), as recently demonstrated in pancreatic cancer and melanoma <sup>48-50</sup>. Assessment of the effects of 'priming' with fasudil on the 201 202 stroma demonstrated that transient ROCK inhibition reduced ECM remodeling 203 and tissue stiffness, thereby altering integrin signaling and depriving cancer cells of mechanical cues provided by the matrix <sup>26</sup>. In addition, decompression 204 205 of the tumor tissue upon 'priming' with fasudil was accompanied by relaxation 206 and increased permeability of the tumor vasculature, as assessed by the 207 imaging of quantum dots diffusing from blood vessels and into tumor tissue 208 (see schematic representation of ROCK inhibition at the whole-body level Fig. 1B and Movie 1)<sup>26</sup>. This is in line with the current clinical use of fasudil for the 209 treatment of cerebral vasospasm 36, 43 and with recent work demonstrating 210 that ROCK regulates vascular patency, or obstruction <sup>51</sup>. Our findings 211 212 therefore demonstrate that fasudil has a dual effect on both the ECM and the 213 intratumoral vasculature, which together increased drug delivery and 214 improved cancer cell responses to chemotherapy. This aligns with recent 215 stromal-based strategies in metastatic colorectal cancer, where the 216 combination of anti-VEGF therapy and anti-hyaluronic acid treatment 217 significantly improved chemotherapy efficacy and prolonged survival

compared to anti-VEGF therapy alone <sup>52</sup>. Our work also indicates that rather 218 219 than chronic treatment, which has a greater potential for adverse effects and toxicity <sup>11, 14</sup>, acute fasudil treatment to induce transient mechanical 'priming' 220 was sufficient to re-equilibrate the pancreatic tumor stroma and to impair 221 222 PDAC progression. Together, our findings demonstrate that 'priming' with 223 fasudil might be beneficial both in the neo-adjuvant and adjuvant settings, 224 which strongly suggests that further clinical assessment of fasudil in 225 combination with standard-of-care chemotherapy, such as Gemcitabine and 226 Abraxane, is warranted to improve PDAC patient outcomes.

227

### Balancing cell contractility: a new approach to treat pancreatic cancer.

229 studies While numerous have demonstrated that extensive 230 transformation of the pancreatic stroma occurs during cancer development<sup>5</sup>, <sup>53</sup>, previous work assessing ECM-based therapies have vielded conflicting 231 data regarding the efficacy of stromal therapies in pancreatic cancer. As such, 232 while pharmacological inhibition of the Hedgehog (Hh) signaling pathway<sup>4</sup>, 233 hyaluronic acid (HA) deposition <sup>13, 15</sup> or lysyl oxidase (LOX) activity <sup>12</sup> resulted 234 235 in impaired tumor growth and increased survival in mouse models of pancreatic cancer, genetic ablation of Hh signaling <sup>14</sup> or myofibroblasts <sup>11</sup> 236 237 resulted in decreased survival. Importantly, ablation of fibrosis triggered 238 adverse effects on the pancreatic stroma, such as profound alterations of the immune microenvironment, which in turn promoted cancer progression <sup>11, 14</sup>. 239 240 Identification of new ECM targets and development of innovative therapeutic 241 regimens to 'fine-tune' and manipulate the pancreatic stroma are therefore 242 needed to improve pancreatic cancer patient outcomes. We believe that this

balance is key to future development of stromal targeting strategies for thisdisease.

Our two recent publications <sup>25, 26</sup> establish ROCK as a key regulator of 245 246 matrix remodeling in pancreatic cancer, both via generation of contractile 247 force, and regulation of MMP synthesis and release into the surrounding 248 matrix (see schematic representation of ROCK inhibition at the cellular level, 249 Fig. 1A). These findings align with recent work in pancreatic cancer 250 demonstrating that the JAK/ROCK/STAT3 signaling pathway governs cancer 251 cellular tension and promotes tumor progression via remodeling of the surrounding matrix in close proximity to the tumor <sup>53</sup>. Our observations also 252 253 highlight the intricate effects of ROCK-induced remodeling of the ECM. While 254 prolonged exposure to fasudil significantly increased mechanical constraints 255 and reduced tumor growth in the KPC model, potentially via reduced release of MMPs into the environment, transient 'priming' with fasudil led to reduced 256 257 ECM crosslinking and relaxation of tumor tissue. This aligns with the 258 emerging concept that the pancreatic stroma can both promote and restrain disease progression <sup>8, 16</sup>. Importantly, our work provides pre-clinical evidence 259 260 that fine-tuning the ECM via transient ROCK inhibition using our 'priming' 261 approach might provide new avenues for the treatment of pancreatic cancer. 262 Potential hypotensive effects of ROCK inhibition with fasudil might be 263 expected given its use for cerebral vasospasm, however the actions on the 264 vasculature that we observe also have the potential beneficial effect of 265 increasing drug delivery. Consistent with recently published work from the 266 Weaver lab, we report no significant change in patient survival associated with bulk tumor stroma <sup>26, 53</sup>, however our study demonstrates a graded response 267

268 to the 'priming' strategy in patient-derived xenografts that had been stratified based on their ECM signature <sup>26</sup>. Where in tumors with high ECM content, 269 'priming' with fasudil greatly improved cancer cell responses to chemotherapy, 270 271 delayed metastasis and approximately doubled survival compared to 272 chemotherapy alone, this had a modest effect in tumors with low ECM content 273 <sup>26</sup>. This observation suggested that initial collagen content could be used as a 274 surrogate biomarker alone, or because of the dual effects of fasudil 'priming' 275 on the ECM and the intratumoral vasculature, in combination with tumor 276 vasculature markers, such as CD31 (cluster of differentiation 31), to identify 277 patients most likely to benefit from transient ROCK inhibition prior to 278 chemotherapy (see schematic representation companion biomarker strategy, 279 Fig. 1C). Additionally, non-invasive PET-reporters of fibrotic tissue are being 280 developed for diagnosis of pulmonary fibrosis, which could be repurposed in this context <sup>54</sup>. We propose that the repurposing of a low-cost, off-patent drug 281 282 such as fasudil as a 'priming' agent might be beneficial for pancreatic cancer 283 therapy. In addition, novel ROCK inhibitors such as AT13148, KD025 or 284 CCT129254, currently in the clinical testing pipeline as anti-fibrotic agents, or in phase I clinical trial for the treatment of solid tumors (AT13148, 285 NCT01585701<sup>55</sup>) could also have similar applications<sup>56-59</sup>. Remodeling of the 286 stroma has also been reported to occur in other solid cancers and to influence 287 disease progression <sup>7, 48, 60, 61 62 63</sup>. Therefore, we envisage that fine-tuning the 288 289 ECM via ROCK inhibition prior to standard-of-care therapies might lead to 290 substantial therapeutic benefits in additional diseases.

291

### 292 Acknowledgements

- 293 Funding was provided from Cancer Research UK to MFO (A18276) and to the
- 294 Cancer Research UK Beatson Institute (A17196), NHMRC, Cancer Council

295 NSW, Cancer Australia, Tour de Cure grants, Cancer Institute NSW, ARC

296 Future, Lens Ainsworth and Philip Hemstritch Pancreatic Cancer Fellowships,

- 297 Sydney Catalyst scholarship.
- 298
- 299

# 300 References

3011.Siegel RL, Miller KD, Jemal A. Cancer Statistics, 2017. CA Cancer J Clin3022017; 67:7-30.

Spadi R, Brusa F, Ponzetti A, Chiappino I, Birocco N, Ciuffreda L, et al.
Current therapeutic strategies for advanced pancreatic cancer: A review for
clinicians. World J Clin Oncol 2016; 7:27-43.

306 3. Yang KS, Im H, Hong S, Pergolini I, Del Castillo AF, Wang R, et al.
307 Multiparametric plasma EV profiling facilitates diagnosis of pancreatic
308 malignancy. Science translational medicine 2017; 9.

309 4. Olive KP, Jacobetz MA, Davidson CJ, Gopinathan A, McIntyre D, Honess D,
310 et al. Inhibition of Hedgehog signaling enhances delivery of chemotherapy in a
311 mouse model of pancreatic cancer. Science 2009; 324:1457-61.

5. Neesse A, Michl P, Frese KK, Feig C, Cook N, Jacobetz MA, et al. Stromal biology and therapy in pancreatic cancer. Gut 2011; 60:861-8.

6. Harris NL, Vennin C, Conway JR, Vine KL, Pinese M, Cowley MJ, et al.
SerpinB2 regulates stromal remodelling and local invasion in pancreatic cancer.
Oncogene 2017; 63:1-11.

7. Pickup MW, Mouw JK, Weaver VM. The extracellular matrix modulates thehallmarks of cancer. EMBO reports 2014; 15:1243-53.

8. Rath N, Olson MF. Regulation of pancreatic cancer aggressiveness by
stromal stiffening. Nature Medicine 2016; 22:462-3.

9. Nobis M, McGhee EJ, Morton JP, Schwarz JP, Karim SA, Quinn J, et al.
Intravital FLIM-FRET imaging reveals dasatinib-induced spatial control of src in
pancreatic cancer. Cancer Res 2013; 73:4674-86.

10. Chang J, Lucas MC, Leonte LE, Garcia-Montolio M, Singh LB, Findlay AD, et
al. Pre-clinical evaluation of small molecule LOXL2 inhibitors in breast cancer.
Oncotarget 2017; 8:26066-78.

327 11. Ozdemir BC, Pentcheva-Hoang T, Carstens JL, Zheng X, Wu CC, Simpson

328 TR, et al. Depletion of carcinoma-associated fibroblasts and fibrosis induces

immunosuppression and accelerates pancreas cancer with reduced survival.Cancer Cell 2014; 25:719-34.

331 12. Miller BW, Morton JP, Pinese M, Saturno G, Jamieson NB, McGhee E, et al.
332 Targeting the LOX/hypoxia axis reverses many of the features that make

333 pancreatic cancer deadly: inhibition of LOX abrogates metastasis and enhances 334 drug efficacy. EMBO Mol Med 2015; 7:1063-76. Jacobetz MA, Chan DS, Neesse A, Bapiro TE, Cook N, Frese KK, et al. 335 13. 336 Hyaluronan impairs vascular function and drug delivery in a mouse model of 337 pancreatic cancer. Gut 2013; 62:112-20. 338 Rhim AD, Oberstein PE, Thomas DH, Mirek ET, Palermo CF, Sastra SA, et 14. 339 al. Stromal elements act to restrain, rather than support, pancreatic ductal 340 adenocarcinoma. Cancer Cell 2014; 25:735-47. 341 15. Provenzano PP, Cuevas C, Chang AE, Goel VK, Von Hoff DD, Hingorani SR. 342 Enzymatic targeting of the stroma ablates physical barriers to treatment of 343 pancreatic ductal adenocarcinoma. Cancer Cell 2012; 21:418-29. 344 Cox T. Erler JT. Fibrosis and cancer: partner in crime or opposing forces? 16. 345 Trends in Cancer 2016; Volume 2 279 - 82. 346 Riento K, Ridley AJ. Rocks: multifunctional kinases in cell behaviour. Nat 17. 347 Rev Mol Cell Biol 2003; 4:446-56. 348 Julian L, Olson MF. Rho-associated coiled-coil containing kinases (ROCK): 18. 349 structure, regulation, and functions. Small GTPases 2014; 5:e29846. 350 Rath N, Olson MF. Rho-associated kinases in tumorigenesis: re-19. 351 considering ROCK inhibition for cancer therapy. EMBO reports 2012; 13:900-8. 352 Sadok A, McCarthy A, Caldwell J, Collins I, Garrett MD, Yeo M, et al. Rho 20. 353 kinase inhibitors block melanoma cell migration and inhibit metastasis. Cancer 354 Res 2015; 75:2272-84. 355 21. Olson MF, Sahai E. The actin cytoskeleton in cancer cell motility. Clinical & 356 experimental metastasis 2009; 26:273-87. 357 Rodriguez-Hernandez I, Cantelli G, Bruce F, Sanz-Moreno V. Rho, ROCK 22. 358 and actomyosin contractility in metastasis as drug targets. F1000Research 2016; 359 5. 360 23. Prudnikova TY, Rawat SJ, Chernoff J. Molecular pathways: targeting the 361 kinase effectors of RHO-family GTPases. Clin Cancer Res 2015; 21:24-9. 362 24. Steeg PS. Targeting metastasis. Nat Rev Cancer 2016; 16:201-18. 363 25. Rath N, Morton JP, Julian L, Helbig L, Kadir S, McGhee EJ, et al. ROCK 364 signaling promotes collagen remodeling to facilitate invasive pancreatic ductal 365 adenocarcinoma tumor cell growth. EMBO Mol Med 2017; 9:198-218. 366 Vennin C, Chin VT, Warren SC, Lucas MC, Herrmann D, Magenau A, et al. 26. 367 Transient tissue priming via ROCK inhibition uncouples pancreatic cancer progression, sensitivity to chemotherapy, and metastasis. Science translational 368 369 medicine 2017; 9. 370 Waddell N, Pajic M, Patch AM, Chang DK, Kassahn KS, Bailey P, et al. 27. 371 Whole genomes redefine the mutational landscape of pancreatic cancer. Nature 372 2015; 518:495-501. 373 Hingorani SR, Wang L, Multani AS, Combs C, Deramaudt TB, Hruban RH, et 28. 374 al. Trp53R172H and KrasG12D cooperate to promote chromosomal instability 375 and widely metastatic pancreatic ductal adenocarcinoma in mice. Cancer Cell 376 2005: 7:469-83. 377 29. Hingorani SR, Petricoin EF, Maitra A, Rajapakse V, King C, Jacobetz MA, et 378 al. Preinvasive and invasive ductal pancreatic cancer and its early detection in 379 the mouse. Cancer Cell 2003; 4:437-50.

- 380 30. Samuel MS, Rath N, Masre SF, Boyle ST, Greenhalgh DA, Kochetkova M, et 381 al. Tissue-selective expression of a conditionally-active ROCK2-estrogen receptor 382 fusion protein. Genesis 2016; 54:636-46. 383 Bardeesy N, DePinho RA. Pancreatic cancer biology and genetics. Nat Rev 31. 384 Cancer 2002; 2:897-909. 385 Rajakyla EK, Vartiainen MK. Rho, nuclear actin, and actin-binding proteins 32. 386 in the regulation of transcription and gene expression. Small GTPases 2014; 387 5:e27539. 388 33. Orgaz JL, Pandya P, Dalmeida R, Karagiannis P, Sanchez-Laorden B, Viros 389 A, et al. Diverse matrix metalloproteinase functions regulate cancer amoeboid 390 migration. Nat Commun 2014; 5:4255. 391 Morton JP, Karim SA, Graham K, Timpson P, Jamieson N, Athineos D, et al. 34. 392 Dasatinib inhibits the development of metastases in a mouse model of pancreatic 393 ductal adenocarcinoma. Gastroenterology 2010; 139:292-303. 394 Pajic M, Herrmann D, Vennin C, Conway JR, Chin VT, Johnsson AK, et al. 35. 395 The dynamics of Rho GTPase signaling and implications for targeting cancer and 396 the tumor microenvironment. Small GTPases 2015; 6:123-33. 397 Chin VT, Nagrial AM, Chou A, Biankin AV, Gill AJ, Timpson P, et al. Rho-36. 398 associated kinase signalling and the cancer microenvironment: novel biological 399 implications and therapeutic opportunities. Expert Rev Mol Med 2015; 17:e17. 400 37. Tamura M, Nakao H, Yoshizaki H, Shiratsuchi M, Shigyo H, Yamada H, et al. 401 Development of specific Rho-kinase inhibitors and their clinical application. 402 Biochim Biophys Acta 2005; 1754:245-52. 403 Liu GJ WZ, Wang YF, Xu LL, Wang XL, Liu Y, Luo GJ, He GH, Zeng YJ. 38. 404 Systematic assessment and meta-analysis of the efficacy and safety of fasudil in 405 the treatment of cerebral vasospasm in patients with subarachnoid 406 haemorrhage. Eur J Clin Pharmacol 2012; 68:131-139. 407 Conway JRW, Warren SC, Timpson P. Context-dependent intravital 39. 408 imaging of therapeutic response using intramolecular FRET biosensors. Methods 409 2017 (in press). 410 Conway JR, Carragher NO, Timpson P. Developments in preclinical cancer 40. 411 imaging: innovating the discovery of therapeutics. Nat Rev Cancer 2014; 14:314-412 28. 413 41. Vennin C, Herrmann D., Lucas M.C., Timpson P. Intravital imaging reveals 414 new ancillary mechanisms co-opted by cancer cells to drive tumor progression. 415 F1000 Research 2016; 5:892. 416 42. Nobis M, Carragher NO, McGhee EJ, Morton JP, Sansom OJ, Anderson KI, et 417 al. Advanced intravital subcellular imaging reveals vital three-dimensional signalling events driving cancer cell behaviour and drug responses in live tissue. 418 419 The FEBS journal 2013; 280:5177-97. 420 43. Vicari RM, Chaitman B, Keefe D, Smith WB, Chrysant SG, Tonkon MJ, et al. 421 Efficacy and safety of fasudil in patients with stable angina: a double-blind, placebo-controlled, phase 2 trial. J Am Coll Cardiol 2005; 46:1803-11. 422 Wojciak-Stothard B, Ridley AJ. Shear stress-induced endothelial cell 423 44. 424 polarization is mediated by Rho and Rac but not Cdc42 or PI 3-kinases. J Cell Biol 425 2003; 161:429-39. 426 45. Barnes JM, Nauseef JT, Henry MD. Resistance to fluid shear stress is a
- 427 conserved biophysical property of malignant cells. PLoS One 2012; 7:e50973.

428 46. Sunyer R, Conte V, Escribano J, Elosegui-Artola A, Labernadie A, Valon L, 429 et al. Collective cell durotaxis emerges from long-range intercellular force 430 transmission. Science 2016; 353:1157-61. 431 Scott RW, Hooper S, Crighton D, Li A, Konig I, Munro J, et al. LIM kinases 47. 432 are required for invasive path generation by tumor and tumor-associated 433 stromal cells. J Cell Biol 2010; 191:169-85. 434 Hirata E, Girotti MR, Viros A, Hooper S, Spencer-Dene B, Matsuda M, et al. 48. 435 Intravital imaging reveals how BRAF inhibition generates drug-tolerant 436 microenvironments with high integrin beta1/FAK signaling. Cancer Cell 2015; 437 27:574-88. 438 Hoshino A, Costa-Silva B, Shen TL, Rodrigues G, Hashimoto A, Tesic Mark 49. 439 M, et al. Tumour exosome integrins determine organotropic metastasis. Nature 440 2015; 527:329-35. 441 Costa-Silva B, Aiello NM, Ocean AJ, Singh S, Zhang H, Thakur BK, et al. 50. 442 Pancreatic cancer exosomes initiate pre-metastatic niche formation in the liver. 443 Nat Cell Biol 2015; 17:816-26. 444 51. Johansson-Percival A, Li ZJ, Lakhiani DD, He B, Wang X, Hamzah J, et al. 445 Intratumoral LIGHT Restores Pericyte Contractile Properties and Vessel 446 Integrity. Cell reports 2015; 13:2687-98. Rahbari NN, Kedrin D, Incio J, Liu H, Ho WW, Nia HT, et al. Anti-VEGF 447 52. 448 therapy induces ECM remodeling and mechanical barriers to therapy in 449 colorectal cancer liver metastases. Science translational medicine 2016; 450 8:360ra135. 451 53. Laklai H, Miroshnikova YA, Pickup MW, Collisson EA, Kim GE, Barrett AS, 452 et al. Genotype tunes pancreatic ductal adenocarcinoma tissue tension to induce 453 matricellular fibrosis and tumor progression. Nat Med 2016; 22:497-505. 454 Desogere P, Tapias LF, Hariri LP, Rotile NJ, Rietz TA, Probst CK, et al. Type 54. 455 I collagen-targeted PET probe for pulmonary fibrosis detection and staging in 456 preclinical models. Sci Transl Med 2017; 9. 457 Kumar R, Mateo J, Smith AD, Khan KH, Ruddle R, Swales KE, et al. First-in-55. 458 human, first-in-class phase 1 study of a novel oral multi-AGC kinase inhibitor 459 AT13148 in patients (pts) with advanced solid tumors. Journal of Clinical 460 Oncology 2014; 15:2554. 461 Flynn R, Paz K, Du J, Reichenbach DK, Taylor PA, Panoskaltsis-Mortari A, 56. 462 et al. Targeted Rho-associated kinase 2 inhibition suppresses murine and human chronic GVHD through a Stat3-dependent mechanism. Blood 2016; 127:2144-54. 463 464 57. Sadok A, Marshall CI. Rho GTPases: masters of cell migration. Small 465 GTPases 2014; 5:e29710. Zanin-Zhorov A, Weiss JM, Trzeciak A, Chen W, Zhang J, Nyuydzefe MS, et 466 58. al. Cutting Edge: Selective Oral ROCK2 Inhibitor Reduces Clinical Scores in 467 468 Patients with Psoriasis Vulgaris and Normalizes Skin Pathology via Concurrent Regulation of IL-17 and IL-10. J Immunol 2017 (in press). 469 Bertolini F, Sukhatme VP, Bouche G. Drug repurposing in oncology--470 59. patient and health systems opportunities. Nat Rev Clin Oncol 2015: 12:732-42. 471 472 Samuel MS, Lopez JI, McGhee EJ, Croft DR, Strachan D, Timpson P, et al. 60. 473 Actomyosin-mediated cellular tension drives increased tissue stiffness and beta-474 catenin activation to induce epidermal hyperplasia and tumor growth. Cancer 475 Cell 2011; 19:776-91.

- 476 61. Levental KR, Yu H, Kass L, Lakins JN, Egeblad M, Erler JT, et al. Matrix
- 477 crosslinking forces tumor progression by enhancing integrin signaling. Cell 2009;478 139:891-906.
- 479 62. Madsen CD, Pedersen JT, Venning FA, Singh LB, Moeendarbary E, Charras
- 480 G, et al. Hypoxia and loss of PHD2 inactivate stromal fibroblasts to decrease
- 481 tumour stiffness and metastasis. EMBO reports 2015; 16:1394-408.
- 482 63. Kular J, Scheer KG, Pyne NT, Allam AH, Pollard AN, Magenau A, et al. A
- 483 Negative Regulatory Mechanism Involving 14-3-3zeta Limits Signaling
- 484 Downstream of ROCK to Regulate Tissue Stiffness in Epidermal Homeostasis.
- 485 Developmental cell 2015; 35:759-74.
- 486

488

# 489 Figure and movie legends

## 490 Figure 1 Schematic of the roles of ROCK and ROCK inhibition in

### 491 pancreatic cancer: from cell-to-global effects to translation to patients.

492 A. ROCK inhibition at the cellular level impairs ECM remodeling via

493 decreased MMP release and impaired contractility. B. ROCK inhibition at the

- 494 whole body, global level. Schematic representation of the effects of ROCK
- inhibition in primary tumor tissue (left hand panel), on circulating tumor cells
- 496 (CTC, middle panel) and at secondary sites (right hand panel). Adapted from
- 497 (Vennin et al., Science Translational Medicine 2017)<sup>26</sup>. Reprinted with

498 permission from AAAS. C. Combination of ECM and vasculature markers as

499 companion biomarkers for priming strategy. Left hand panel: Schematic

- 500 representation of in-house automated Second Harmonic Generation (SHG)
- analysis of the ECM in the ICGC human TMA cohort, with examples of SHG
- 502 signals in cores (triplicates) from patients with high, medium, or low SHG
- signal. Right hand panel: representative images of quantum dots and CD31

504 (cluster of differentiation 31) staining in tumors with high and low vascularity.

505 Adapted from (Vennin et al., Science Translational Medicine 2017)<sup>26</sup>.

506 Reprinted with permission from AAAS.

507

508 **Movie 1:** Intravital imaging of quantum dots circulating in tumor associated

509 blood vessels and diffusing into the surrounding tumor tissue. Red: Quantum

510 Dot, Blue: Collagen fibers (SHG signal).

### A. ROCK inhibition at the cellular level



### B. ROCK inhibition at the whole body level



C. Companion biomarkers: initial ECM/vasculature markers to predict tailored patient response to 'priming' approach

