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1 **Proteomics in cardiovascular disease: recent progress and clinical implication and**  
2 **implementation**

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10

11 **Abstract**

12 Introduction: Although multiple efforts have been initiated to shed light into the molecular  
13 mechanisms underlying cardiovascular disease, it still remains one of the major causes of  
14 death worldwide. Proteomics approaches are unequivocally powerful tools that may provide  
15 deeper understanding into the molecular mechanisms associated with cardiovascular  
16 disease and improve its management.

17 Areas covered: Cardiovascular proteomics is an emerging field and significant progress has  
18 been made during the past few years with the aim of defining novel candidate biomarkers  
19 and obtaining insight into molecular pathophysiology. To summarize the recent progress in  
20 the field, a literature search was conducted in PubMed and Web of Science. As a result, 704  
21 studies from PubMed and 320 studies from Web of Science were retrieved. Findings from  
22 original research articles using proteomics technologies for the discovery of biomarkers for  
23 cardiovascular disease in human are summarized in this review.

24 Expert commentary: proteins associated with cardiovascular disease represent pathways in  
25 inflammation, wound healing and coagulation, proteolysis and extracellular matrix  
26 organization, handling of cholesterol and LDL. Future research in the field should target to  
27 increase proteome coverage as well as integrate proteomics with other omics data to  
28 facilitate both drug development as well as clinical implementation of findings.

29 **Keywords:** cardiovascular disease, vascular disease, proteome, biomarker, clinical  
30 proteomics

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32

## 33 **1. Introduction**

34 Cardiovascular disease (CVD) is the leading cause of death worldwide with a high mortality  
35 rate and an increasing incidence annually. Since it represents a major contributor to the  
36 global disease burden, gaining insights into the mechanisms of CVDs is emergent. CVD is a  
37 complex disorder describing pathophysiological conditions of the heart and blood vessels  
38 and includes a wide range of diseases such as coronary artery disease (CAD), heart failure  
39 (HF), myocardial infarction (MI), abdominal aortic stenosis, ischemic stroke, cardiomyopathy  
40 etc [1]. Several factors such as lifestyle, diet, (epi)genetics, dyslipidemia, hypertension, and  
41 inflammation are associated with the onset and development of CVD. Addressing the  
42 standard risk factors for CVD including hypertension, diabetes mellitus, smoking, and  
43 hypercholesterolemia is a major aim in the prevention of CVD. Nevertheless, the current  
44 treatment schemes, by addressing these individual risk factors, have not fully provided  
45 either a significant progress in curative treatment or a full understanding of the disease.  
46 Over the past years, efforts to elucidate the key molecular mechanisms underlying CVD were  
47 undertaken. Its complex etiology and the plethora of the different factors that are  
48 responsible for the observed pathology, require the development of more holistic treatment  
49 approaches targeting the underlying mechanisms [2]. The development and application of  
50 systems biology approaches for CVD investigation is an active and challenging research field  
51 [2-4].

52 Integrated omics approaches, including genomics/epigenomics, transcriptomics, proteomics,  
53 metabolomics have gained momentum the past few years. They provide information at  
54 multiple biological levels from the DNA and RNA to the protein and metabolite patterns,  
55 offering a different perspective on the molecular and cellular networks of whole organ  
56 system. Although, genomic and transcriptomic strategies have been recently applied in CVD

57 research, the weakness of these strategies is that they cannot correlate easily the observed  
58 changes to the expression of proteins and additionally, they cannot characterize the post-  
59 translational modifications (PTM) which play critical role in the regulation of many biological  
60 processes. These shortcomings can be addressed by the application of proteomic  
61 approaches. **In fact, proteomic changes are likely the cause of most non-communicable**  
62 **diseases, including CVD, and interfering drugs generally act via targeting specific proteins.** As  
63 graphically depicted in **Figure 1**, based on these facts it appears evident that the proteome  
64 will inform with superior accuracy on (any) disease, and on the best suited therapeutic  
65 intervention, leading towards personalized medicine.

66 Two main strategies appear of relevance in the exploitation of proteomic changes in the  
67 context of CVD: 1) obtain information about molecular pathophysiology and enable  
68 identifying relevant targets for intervention, and 2) identification of biomarkers that support  
69 patient management and guide therapy. As depicted in **Figure 2**, these two strategies do  
70 have different requirements. While implementation of a biomarker requires very high  
71 confidence in a biomarker and the test to be used (needs to be verified in large cohorts,  
72 likely exceeding 1000 subjects, and needs to demonstrate a significant improvement over  
73 the current state of the art), knowledge on the specific role of the biomarker in the  
74 molecular pathology is not a prerequisite for its employment (e.g even if we do not know  
75 why a patient has fever or elevated blood pressure, we nevertheless typically act upon  
76 detection of a significant change from the reference range in these biomarkers). In contrast,  
77 identifying drug targets requires in depth knowledge on molecular pathology. A major  
78 prerequisite to unravel the biological processes and pathways (and to consequently enable  
79 identifying the best suited drug targets) is a comprehensive coverage of the proteomic  
80 changes in the respective disease. To enable such in depth coverage, reduced stringency in

81 statistical evaluation needs to be accepted; it is impossible to verify ideally thousands of  
82 (sometimes) subtle changes observed in disease.

83 To review the current status of proteomic biomarkers associated with CVD, we conducted a  
84 literature search in PubMed using the keywords [proteomic; biomarker; cardiovascular  
85 disease] or the search query [cardiovasc\*and proteom\* and marker or biomarker] in the  
86 Web of Science. Following this combined search, 704 studies from PubMed and 320 from  
87 Web of Science published between January 2010 and October 2016) were initially retrieved.  
88 Studies focusing on animal models and reviews were excluded. Only studies in human using  
89 proteomics technologies for the discovery of biomarkers in cardiovascular disease were  
90 included (all listed as supplementary table). Based on these articles, we discuss in this review  
91 the recent advances in the development of specific biomarkers for CVD through the  
92 application of state of the art proteomic technologies and the progress that was made  
93 towards their clinical implementation. The general workflow that is followed in these studies  
94 and the molecules that are currently being used as CVD biomarkers in the clinical routine is  
95 shown in **Figure 3**. A brief overview of main employed proteomics platforms and types of  
96 specimens analyzed in CVD research, followed by a more detailed presentation of biomarker  
97 findings, as retrieved from the aforementioned literature search, follow.

98

## 99 **2. Overview of the state of the art proteomics technologies in CVD**

100 Quantitative methods can be divided into global and targeted proteomic approaches. The  
101 global approaches compare two or more proteomes for the identification of the  
102 differentially expressed proteins under physiological and pathological conditions and can be  
103 applied to the unbiased discovery of biomarkers. Global approaches are further categorized  
104 into gel based and gel free [Liquid chromatography-tandem mass spectrometry (LC-MS/MS)

105 based]. Gel based approaches, such as two-dimensional electrophoresis (2DE) and two-  
106 dimensional Differential-In-Gel-Electrophoresis (2D-DIGE), have contributed in CVD [5-8].  
107 However, 2DE is of lower resolving power in comparison to LC-MS/MS, allowing, by now, to  
108 assess with good confidence > 1000 proteins in complex mixtures per experiment.

109 As a result, LC-MS/MS has become a commonly employed method due to its sensitivity and  
110 high resolving power. There are two main strategies to perform quantitative proteome  
111 analysis, by either using label-free strategies where the peptides from 2 or more samples are  
112 injected into the mass spectrometer and analyzed in individual LC-MS experiments or by  
113 using the isotope labeling approaches. Labeling proteomic approaches include stable isotope  
114 labeling by amino acids in cell culture (SILAC), in which isotopically labeled amino acids are  
115 incorporated into cell culture media, as well as isobaric tags for relative and absolute  
116 quantification (iTRAQ) [9] or tandem mass tags (TMT), applicable to the study of any type of  
117 biological samples (beyond cell cultures) [10]. In a recent study, employing multiplexed  
118 iTRAQ, intensive depletion of abundant plasma proteins, optimized fractionation methods  
119 along with the use of the latest mass spectrometry (MS) instrumentation on plasma samples  
120 from patients undergoing a therapeutic, planned myocardial infarction (PMI) for treatment  
121 of hypertrophic cardiomyopathy [11], about 5000 proteins were identified in one analysis.

122 In studies that focus on analyzing a large number of heterogeneous samples containing  
123 interfering compounds such as lipids, the use of capillary electrophoresis coupled to mass  
124 spectrometry (CE-MS) appears advantageous [12]. This technique has been applied for the  
125 proteomic analysis of urine and has enabled the identification of novel candidate biomarkers  
126 in CAD [13], stroke [14], HF [15] and deep vein thrombosis [16]. These studies are  
127 summarized in the following sections.

128 Targeted proteomic approaches characterize and quantify a limited number of known target  
129 proteins from a complex sample. Their application includes a follow-up on proteins  
130 identified from global/unbiased proteomics screening, conducting functional studies such as  
131 post-translational modification (PTM) analysis and validation of biomarker candidates.  
132 Selected reaction monitoring (SRM) or multiple reaction monitoring (MRM) is frequently  
133 employed in targeted proteomics and has been applied for biomarker [17-19] and PTM  
134 quantification [20,21]. In some instances SRM has been combined with the Protein standard  
135 absolute quantification (PSAQ) strategy, which uses full-length isotope-labeled protein  
136 standards to quantify target proteins. The PSAQ-SRM method was used for the  
137 quantification of four biomarkers (LDH-B, CKMB, myoglobin and troponin I) in MI patients'  
138 sera with good accuracy and reproducibility [22].

139 In recent years, array-based techniques have also been applied in order to identify  
140 biomarkers in atherosclerosis [23,24], abdominal aortic aneurism (AAA) [25] and CAD [26].  
141 Additionally, aptamer-based proteomics have been applied for the identification of low-  
142 abundance biomarkers in CVD [27]. However, due to the small sample size and the lack of  
143 validation studies, the value of these approaches cannot be assessed yet.

144

### 145 **3. Samples for proteome analysis in CVD**

#### 146 **3.1 Tissue**

147 Tissue proteomics in CVD have generated insight into the proteome of atherosclerotic  
148 plaques [6,7] and thrombi [28]. Although a number of studies have focused on the  
149 proteomic analysis of the whole atheromatous tissue of carotid and coronary arteries, this  
150 approach suffers from the existence of different layers; the adventitial, the medial and the  
151 intima. The latter represents the layer where a wide variety of changes occur upon



152 atherosclerosis development and progression. To overcome this barrier, arterial tissue  
153 subproteomes isolated by laser microdissection (LMD) from human atherosclerotic and pre-  
154 atherosclerotic coronaries have been studied [7]. This selective approach is also considered  
155 essential for studying the different cell populations or the cellular/extracellular components  
156 to better understand their contribution in the pathogenesis of the disease [29]. Finally,  
157 tissue secretome represents an alternative sample source. Tissue explants directly obtained  
158 from healthy and diseased arteries are cultured and their supernatant which is enriched in  
159 secreted proteins, is used for proteomics analysis [30].

### 160 **3.2 Sub-cellular proteomes**

161 The majority of the sub-cellular studies in the field of cardiovascular proteomics focus on the  
162 analysis of the mitochondrial [31] and extracellular matrix (ECM) proteomes [32]. Other sub-  
163 cellular proteomes, including myofilaments [33], sacromere [34] and proteasomes [35] have  
164 been less extensively investigated. Isolation of the sub-cellular fractions in general involves  
165 application of differential centrifugation, immune-based isolation, and membrane protein  
166 enrichment [36] methods.

#### 167 **3.2.1 ECM**

168 The ECM is involved in a broad spectrum of signaling events during growth, differentiation,  
169 injury, and remodeling [37]. Technical challenges are the insolubility and extensive PTMs of  
170 the ECM proteins, and their low abundance (~1% of proteome) compared to the highly  
171 abundant cellular or plasma proteins. ECM enrichment strategies via decellularization,  
172 sequential solubilization and extraction of ECM constituents have been applied to overcome  
173 these obstacles [38-41]. Of note, Didangelos et al, through the application of these  
174 procedures, analyzed the ECM proteome in human aortic samples [39] and identified 103  
175 ECM proteins with the one third of them not previously described in the context of vascular

176 tissue proteomics. The same group characterized ECM remodeling in abdominal aortic  
177 aneurysms [42]. ECM remodeling has also been described in HF and cardiac fibrosis [32,43].

### 178 **3.2.2 Mitochondria**

179 In a healthy cardiomyocyte, mitochondria represent ~40% of the volume. Mitochondria are  
180 essential for providing energy (for the contraction of the muscle), but also for regulating  
181 programmed cell death. Accumulating evidence supports that CVD progression is associated  
182 with changes in mitochondria structure and function [44,45]. An increasing number of  
183 studies focus on the characterization of alterations of the mitochondrial proteome in CVD  
184 [31], including pressure overload-induced HF [46], atrial fibrillation (AF) [47] and type I [48]  
185 and type II diabetic heart [49]. Goudarzi et al., performed LC-MS/MS analysis of isolated  
186 mitochondrial enriched fractions from right atrial tissues and identified 32 differentially  
187 expressed proteins in patients with AF compared to non-AF patients [47]. Other studies have  
188 also tried to analyze the effect of oxidative stress on cardiac mitochondrial protein dynamics  
189 by using *in vivo* models [50].

### 190 **3.3 Biological fluids**

191 Biological fluids, such as serum, plasma and urine, represent the ideal specimen for  
192 biomarker detection because they are easily accessible. Since urine and plasma account for  
193 the vast majority of biomarker studies, they will be reviewed extensively in the section on  
194 biomarkers (below).

### 195 **3.4 Circulating cells and extracellular vesicles**

196 Proteome analysis of circulating cells such as monocytes, platelets and endothelial cells may  
197 provide a better understanding of the involvement of these cells in CVD and enable  
198 identification of disease associated biomarkers. Monocytes play important roles in  
199 inflammation and atherosclerosis and therefore their proteomic profiling may provide novel

200 insights into their function, as indicated from recent studies [51,52]. Additionally, proteomic  
201 analyses of platelets in CVD [60,61], as well as circulating endothelial cells (CECs) and the  
202 endothelial progenitor cells (EPCs) in patients with ACS have also been described [53].  
203 Plasma microvesicles (including microparticles and exosomes) are advocated as source of  
204 vascular-specific disease biomarkers. Membrane microvesicles, mainly released from  
205 activated platelets into the circulation, represent an important mode of intercellular  
206 communication and their number is increased in patients with acute coronary syndromes  
207 [54]. Comprehensive proteomic analysis of the plaque microparticles derived from human  
208 atherosclerotic lesions revealed that they derived primarily from leukocytes and are  
209 implicated in inflammation [55].

#### 210 **4. PTMs**

211 Protein PTMs, including phosphorylation, ubiquitylation, acetylation, *N*- and *O*-linked  
212 glycosylation, and methylation, are key regulators of the protein conformation, stability and  
213 activity [56].

214 Protein phosphorylation is critical to myocardial function since it underpins cellular  
215 processes associated with energy metabolism, signal transduction and contractile function of  
216 the myocyte [57]. Thus, phosphorylation is the most commonly described PTM in the cardiac  
217 proteome [58,59]. Changes in the abundance of 25 phosphoproteins have been previously  
218 identified in hypertensive cardiac remodeling through the application of 2D-DIGE on  
219 phosphoenriched proteins [60]. Among them, myofilament proteins such as Alpha-  
220 tropomyosin and Myosin; mitochondrial proteins such as Pyruvate dehydrogenase A;  
221 phosphatases such as Protein phosphatase 2A; other proteins including Proteasome subunits  
222  $\alpha$ -type 3 and  $\beta$  type 7 and Eukaryotic translation initiation factor 1A, were included.  
223 Employing top-down MS-based quantitative proteomics, the phosphorylation of cardiac

224 Troponin I has been identified as a biomarker for congestive heart failure (CHF) [61], as  
225 described below.

## 226 **5. Biomarkers currently applied**

227 Biomarkers must provide accurate and reliable information about disease in order to aid in  
228 prognosis, diagnosis, or therapy monitoring. Currently, there are four biomarkers  
229 recommended for clinical use in CVD (summarized in **Figure 3**): 1) Cardiac Troponin T and  
230 cardiac Troponin I [62] are used for the diagnosis of acute coronary syndrome (ACS) and MI,  
231 while elevated plasma levels of troponin T represent the gold standard approach to detect  
232 MI [63] with a high predictive value [64]. The development of high-sensitivity assays for  
233 Troponin I and T has improved the diagnostic sensitivity for acute myocardial infarction  
234 (AMI) [65]. 2) B-type natriuretic peptide (BNP) and its N-terminal form are employed to  
235 detect congestive heart failure [66]. Circulating concentrations of BNP reveal strong  
236 associations with CVD risk under a range of different clinical conditions. 3) C-reactive protein  
237 (CRP) [67,68] and D-dimer [69], are inflammatory markers associated with ischemic heart  
238 disease. 4) Apolipoprotein A-I , the major protein component of HDL, is an excellent risk  
239 predictor for CVD related to the metabolism of high-density lipoprotein (HDL) [70]. While  
240 these biomarkers have clearly demonstrated value in multiple studies, a significant limitation  
241 appears in that they detect mainly late stage of CVD.

242 In the following section we present an overview of potential biomarkers for CVD that have  
243 been recently identified using proteomic technologies, categorized by the biological source  
244 investigated, and summarized in the respective tables.

245

## 246 **6. Proteomic biomarkers in CVD**

### 247 **6.1 Tissue**

248 The affected tissue certainly holds the most relevant information on molecular pathology,  
249 enabling identifying potential therapeutic targets. Tissue is not easily accessible, and  
250 certainly cannot be assessed for routine diagnosis. However, results from tissue proteomics  
251 may enable generating hypotheses on circulating biomarkers, which can subsequently be  
252 tested [71]. Several studies therefore focused on tissue, as summarized in **Table 1**.

### 253 **6.1.1 Arterial Tissue**

254 Atherosclerotic plaque may hold proteomic information on molecular alterations that  
255 discriminate patients with adverse cardiovascular events from those that remain stable  
256 during follow-up. De Kleijn et al. investigated the carotid atherosclerotic plaque proteome by  
257 MS in relation to outcome, during a 3-year follow-up. With this approach, Osteopontin  
258 (OPN) was identified as a potential instable plaque biomarker and subsequently validated in  
259 femoral plaque samples, indicating predictive value independent of plaque localization. The  
260 authors concluded that high levels of OPN from atherosclerotic plaques could be prognostic  
261 of the occurrence of cardiovascular events [72]. Hao et al. fractionated proteins from human  
262 carotid atherosclerotic plaques using Electrostatic Repulsion-Hydrophilic Interaction  
263 Chromatography followed by offline LC-MS/MS. Several previously undetected low-  
264 abundant proteins with important functions in atherosclerosis, such as TGF- $\beta$ , interleukins  
265 and other growth factors were identified. The authors highlighted three proteins;  
266 Myeloperoxidase (MPO), Fibrinogen gamma chain (FGG) and Fibrinogen beta chain (FGB), as  
267 potential CVD biomarkers [73].

268 To overcome the barrier of early studies using whole tissue proteome, LMD of the intimal  
269 layer from human atherosclerotic and pre-atherosclerotic coronaries was used in  
270 combination with a 2D-DIGE approach. This study identified 13 proteins, with three of them,

271 Annexin 4 (ANXA4), Myosin regulatory light 2 smooth muscle isoform (MYL9) and Ferritin  
272 light chain (FTL), representing novel findings in the atherosclerotic coronary intima [7].

### 273 **6.1.2 Cardiac tissue**

274 The number of proteomic studies using human heart tissue samples is low, due to obvious  
275 difficulties in sampling. One study performed by Kakimoto et al. included the proteomic  
276 analysis of human cardiac tissue obtained from cases of sudden death due to AMI. In this  
277 study, using LC-MS/MS, reduced levels of the sarcoplasmic protein, Sorbin and SH3 domain-  
278 containing protein 2 (SORBS2) was detected in the infarcted myocardia. SORBS2 levels were  
279 also found increased in serum of AMI patients, indicating its potential to be used as a  
280 biomarker [74].

281 Aiming at identification of novel biomarkers for early detection of CHF, Zhang et al.,  
282 characterized the PTMs associated with disease progression. Top-down quantitative  
283 proteomics using affinity chromatography and high resolution MS was applied onto human  
284 heart tissue samples. Phosphorylation of cardiac Troponin I was identified as a candidate  
285 biomarker for detection of CHF [61].

### 286 **6.1.3 Atherosclerotic vulnerable plaques**

287 Aiming at identifying circulating biomarkers derived from atherosclerotic vulnerable plaques  
288 that could predict adverse cardiovascular events, Malaud et al. performed protein  
289 enrichment followed by 2DE in protein extracts obtained from human fibrotic and  
290 hemorrhagic carotid atherosclerotic plaques. Several proteins were found to be differentially  
291 released by vulnerable hemorrhagic human carotid plaques when compared with stable  
292 fibrotic plaques. The authors suggest that combinations of the circulating biomarkers  
293 Calponin-1 (CNN1), IL-8, DJ-1 (PARK7), Vascular endothelial growth factor (VEGF) and PCPE-1  
294 (PCOLCE) could be used for coronary patient stratification [75].

#### 295 **6.1.4 Thrombus**

296 Intra-luminal thrombus (ILT) secretions from AAA patients were analyzed by LC-MS/MS and  
297 two thrombus associated proteins, serum Thrombospondin-1 (TSP-1) and Clusterin (CLU),  
298 were identified at reduced levels in circulation of patients with AAA compared to non  
299 aneurismal controls [76]. Targeting the identification of biomarkers in AAA progression,  
300 Martinez-Pinna et al. analyzed the proteins released by the different layers  
301 (luminal/abluminal) of the ILT by 2D-DIGE. This analysis identified Peroxiredoxin-1 (PRDX1)  
302 as prominently released by the luminal layer of the ILT. PRDX1 levels were further analyzed  
303 by ELISA in the serum of patients with AAA and were found increased compared to the  
304 healthy individuals. A significant positive correlation between PRDX1 serum levels with AAA  
305 size and growth rate had an additive predictive value in AAA, indicating PRDX1 as a potential  
306 biomarker for AAA progression [8]. The same group analyzed the tissue-conditioned media  
307 from patients with AAA by LC-MS/MS and identified that decreased levels and activity of  
308 systemic C3 in advanced AAA stages are associated with AAA evolution [77]. In another  
309 study, emphasis was given to the identification of peptides and low-molecular-weight  
310 proteins released by the different layers of abdominal aortic aneurysm thrombus. By  
311 employing SELDI-TOF mass spectrometry, LVV- Hemorphin7 (H7) and VV-H7, both generated  
312 from hemoglobin proteolysis, were found more abundantly released by recently formed  
313 luminal layer of AAA thrombus relative to intermediate and abluminal older layers. The  
314 levels of the H7 peptides were also found increased in the serum of AAA patients compared  
315 to controls and were positively correlated with the AAA diameter and thrombus volume.  
316 These peptides may be used as biological markers of pathological vascular remodeling [78].  
317 The proteomic profiling of the coronary thrombus from MI patients has been recently  
318 performed using three different proteomic approaches: 2DE-MALDI MS/MS, 1DE- LC-MALDI-

319 MS/MS or 1DE-LC-ESI-MS/MS. Co-expression of 5 focal adhesion proteins [Fermitin homolog  
320 3 (FERMT3), Thrombospondin-1 (THBS1), Myosin-9 (MYH9), Beta parvin (PARVB) and Ras-  
321 related protein Rap-1b (RAP1B)] with CD41 (ITGA2B) was found to be potentially implicated  
322 in platelet activation during thrombus formation. Additionally, Death-inducer obliterator 1a  
323 (DIDO1), was found up-regulated in the plasma of patients and was suggested as potential  
324 biomarker of thrombosis [28]. In an effort to better understand the characteristics of  
325 coronary thrombus, Ramaiola et al. employed 2DE-MS and found that the aged ischemic  
326 thrombi T6 (more than 6h of evolution) in STEMI patients are characterized by reduced  
327 levels of Profilin-1 (PFN-1) compared to T3 (thrombus of less than 3h of evolution) likely due  
328 to its release in the peripheral circulation since its levels were found by ELISA significantly  
329 increased in coronary and systemic blood in T6 patients compared to T3 [79].

### 330 **6.1.5 Tissue secretome**

331 Several proteomics approaches aimed at elucidating the molecular mechanisms involved in  
332 plaque formation to identify potential biomarkers for early diagnosis of disease. The  
333 advantage of using tissue secretome instead of whole tissue proteome is that secretome  
334 may reflect the *in vivo* condition with reduced complexity. Early studies revealed a significant  
335 reduction of soluble TNF-like weak inducer of apoptosis (sTWEAK) in carotid plaque  
336 supernatants compared to normal mammary endartery conditioned media [80]. The same  
337 trend was also described in chronic stable HF [81], peripheral artery disease (PAD) [82], AAAs  
338 [83] and CAD [84]. The potential of sTWEAK as prognostic biomarker in CVD has also been  
339 suggested [85,86].

340 The carotid plaque secretome has also been studied using LC-MS approaches. Among the 31  
341 differentially secreted proteins from plaques, extracellular and intracellular proteins such as  
342 Thrombospondin-1 (THBS1), Vitamin D binding protein (GC), and Vinculin (VCL) were



343 identified. A significantly higher concentration of THBS1 and GC was confirmed in  
344 atherosclerotic subjects in comparison to controls [87].

345 In an attempt to identify novel circulating CAD biomarkers Reiser et al, analyzed the  
346 secretome derived from atherosclerotic plaques of carotid/iliac arteries and control arterial  
347 tissue by antibody phage display followed by MS. In parallel, gene expression analysis of the  
348 coronary thrombi versus peripheral blood mononuclear cells was performed. Integrating the  
349 data from the proteomic and gene expression studies enabled identification of Fatty acid-  
350 binding protein 4 (FABP4) as a candidate biomarker for CAD. FABP4 was found in the plaque  
351 but not in the control secretome and its expression was higher in thrombi than in peripheral  
352 blood mononuclear cells. FABP4 failed to serve either as a clinically relevant diagnostic  
353 marker in stable CAD and ACS or as prognostic biomarker in an asymptomatic population.  
354 However, FABP4 could identify ACS patients at risk for adverse cerebrovascular or  
355 cardiovascular events and therefore the authors concluded that circulating FABP4 may be  
356 utilized as a prognostic biomarker in risk stratification of ACS patients [88].

357

## 358 **6.2 Plasma and Serum biomarkers**

359 Plasma has been the main target specimen of biomarker research in cardiovascular disease,  
360 owed to its availability, and proximity to the affected organ (vessel or heart). The results  
361 obtained are summarized in **Table 2**. Darde et al. [89], investigated the plasma proteome  
362 from patients, healthy controls, and stable CAD patients using immunodepletion of the six  
363 most abundant proteins, and 2DE and 2D-DIGE analysis. Along with proteins previously  
364 associated with ACS, four novel proteins associated with the pathology [Alpha-1- B-  
365 glycoprotein (A1BG), Hakata antigen (FCN3), Tetranectin (CLEC3B) and Tropomyosin 4  
366 (TPM4)] were identified [89]. More recently, Kristensen et al., employed plasma-based

367 quantitative proteomics for the analysis of four well-phenotyped patient cohorts: individuals  
368 1) without cardiovascular symptoms and without the presence of coronary calcium, 2)  
369 without cardiovascular symptoms but with high amounts of coronary calcium, 3) operated  
370 because of atherosclerotic diseases, and 4) with ACS [90]. Through a 5-plex SRM-MS assay,  
371 statistically significant increased levels of the cytoskeletal protein Vinculin (VCL) were  
372 verified, along with other known risk markers of CVD such as CRP, Serum amyloid protein A  
373 (SAA1) and Apolipoprotein-A (APOA1) in the ACS group [90]. Lepedda et al., performed a  
374 comprehensive analysis of the differentially expressed Apolipoprotein component of plasma  
375 VLDL, LDL and HDL from patients undergoing carotid endarterectomy compared to healthy  
376 individuals. A panel of twenty three proteins was identified through the application of 2DE  
377 coupled with Matrix-Assisted Laser Desorption/Ionization Time-of-Flight (MALDI-TOF) MS  
378 analysis. Among them, Acute-phase serum amyloid A protein (SAA1) was over-expressed in  
379 all lipoprotein fractions indicating a potential association with the presence of advanced  
380 carotid lesions [91]. Recently, a novel targeted proteomics approach, based on the proximity  
381 extension assay proteomics chip, allowed the discovery of associations of Osteoprotegerin  
382 (TNFRSF11B), Growth hormone (GH1), Tumor necrosis factor ligand superfamily member 14  
383 (TNFSF14) and Renin (REN) with plaque occurrence, independently of each other and  
384 traditional CVD risk factors [24]. Cheng et al. selected 13 candidate biomarkers possibly  
385 associated with plaque vulnerability and investigated their prognostic value in CAD patients  
386 in a nested-case-control study involving 768 patients undergoing coronary angiography.  
387 Increased plasma levels of circulating Osteoglycin (OGN) and Neutrophil gelatinase-  
388 associated lipocalin/matrix metalloproteinase 9 (NGAL/MMP9) were independently  
389 predictive for occurrence of major adverse cardiovascular event (MACE) within 1 year after

390 coronary angiography. Combining these two proteins with conventional risk factors  
391 significantly improved risk reclassification [92].

392 AAA is associated with inflammatory responses with cytokines as important mediators. By  
393 analyzing the cytokine profile in plasma of patients with AAA via a protein array approach,  
394 elevated Insulin Like Growth Factor Binding Protein 1 (IGFBP1) was identified as a potential  
395 novel disease biomarker [25]. Burillo et al. performed differential quantitative proteome  
396 analysis using iTRAQ labeling and nano-LC-MS/MS of plasma proteins from patients at  
397 different AAA stages of evolution [small (aortic size 3-5cm) vs large AAA (aortic size > 5cm)].  
398 Among the differentially expressed proteins APOA1 was identified significantly reduced in  
399 patients with large AAA compared to small AAA. Furthermore, a strong correlation of APOA1  
400 levels with HDL-Cholesterol (HDL-C) concentration as well as with aortic size and thrombus  
401 volume (negative correlations) were also observed. The authors concluded that the systemic  
402 levels of APOA1/HDL-C are negatively associated with AAA progression [93].

403 To identify novel plasma protein biomarkers that could improve risk assessment for MI and  
404 atherosclerotic cardiovascular disease (ASCVD), Yin et al., compared plasma proteins in  
405 patients and controls through the application of a discovery-validation pipeline. In the  
406 discovery phase, plasma proteins in MI case-control pairs were analyzed using iTRAQ with  
407 multidimensional LC-MS/MS (discovery MS). The most promising protein biomarkers were  
408 subsequently chosen for the validation phase where plasma proteins from ASCVD case-  
409 control pairs were analyzed using MRM. This approach enabled identification of single- and  
410 multiple-marker protein panels significantly associated with CVD, the latter including 4  
411 proteins in combination [ $\alpha$ -1-acid glycoprotein 1 (ORM1), Paraoxonase 1 (PON1), Tetranectin  
412 (CLEC3B), and CD5 antigen-like (CD5L)] [94].

413 Comparative proteome analysis was performed in depleted plasma protein samples from  
414 patients with AMI, unstable angina pectoris (UAP), stable angina pectoris (SAP) and control  
415 subjects, using 2D SDS-PAGE followed by nanoLC-MS/MS analysis. Although two of the  
416 identified proteins, Alpha-1 microglobulin (AMB1) and Vitamin D-binding protein (GC) failed  
417 as suitable CVD markers, Apolipoprotein-A1 (APOA1) isoforms detected by 1D and 2D SDS-  
418 PAGE only in patients groups were presented as possible biomarkers for CVD [95]. In an  
419 effort to identify prognostic biomarkers for AMI, proteome analysis of depleted plasma  
420 samples from AMI patients was performed by 2D-DIGE. A 1-year follow-up was used to  
421 identify patients with heart failure and these data were used for the hierarchical clustering  
422 of the proteomic findings. Among the 22 differentially expressed proteins were isoforms of  
423 Haptoglobin (HP). Their distinct patterns were associated with stratification of the disease  
424 and HP plasma levels predicted the occurrence of HF at 1-year [96].

425 Identification of plasma biomarkers for early detection of myocardial injury is an active  
426 research field; one study applied integrated proteomic technologies in a human model that  
427 faithfully reproduces clinical aspects of spontaneous MI [patients undergoing a therapeutic,  
428 planned myocardial infarction (PMI) for treatment of hypertrophic cardiomyopathy].  
429 Coherent, MS-intensive pipeline, including high-performance LC-MS/MS, accurate inclusion  
430 mass screening (AIMS), stable isotope dilution (SID)-MRM-MS and immunoassays were  
431 employed which collectively revealed a panel of candidate biomarkers that may be specific  
432 to MI. This included previously known cardiovascular biomarkers such as Creatine kinase  
433 MB, Fatty Acid Binding Protein (FABP) and Myeloperoxidase (MPO) and novel candidates  
434 such as Aortic Carboxypeptidase-Like Protein ACLP1, Four And A Half LIM Domains 1 (FHL1),  
435 Myosin Light Chain 3 (MYL3) and Tropomyosin 1 (TPM1) [97]. Applying an aptamer-based  
436 platform in patients with planned and spontaneous MI, 79 candidate infarct markers were

437 detected, including previously known markers (eg. Troponin I and Creatinine kinase MB) and  
438 novel candidates [eg. Fibroblast growth factor 18 (FGF18) and Interleukin-11 (IL11)]. This  
439 study allowed also the identification of low abundance cytokines and cell-surface proteins in  
440 association to disease, not previously reported [27].

441 Analyzing plasma samples from 90 CVD patients with a highly multiplexed MRM-based  
442 assay, Domanski et al assessed 67 putative CVD plasma biomarkers involved in coagulation  
443 and thrombolysis pathways, acute-phase reaction, inflammation as well as lipoprotein  
444 formation [19]. An MRM assay panel was also developed for the detection of ischemic heart  
445 disease and MI, based on 11 proteins with association to CVDs. Comparative analysis of  
446 plasma samples obtained from patients with ST-segment elevation MI and chest pain due to  
447 other causes identified increased levels of Apolipoprotein C1 (APOC1), Apolipoprotein C2  
448 (APOC2) and Apolipoprotein E (APOE) in the former, rising the interest for further studying  
449 the role of these apolipoproteins in the pathophysiology of AMI [98].

450 Plasma proteomics also enables identification of potential biomarkers for acute  
451 decompensated heart failure (ADHF). Employing a discovery platform where depleted  
452 plasma protein samples from ADHF patients were analyzed by nano LC-MALDI-TOF/TOF and  
453 a verification platform based on nano LC-SRM analysis, Mebazaa et al. identified increased  
454 levels of Quiescin Q6 (QSOX1) as a novel biomarker further validated in human studies and  
455 also in animal models. Combination of QSOX1 to the gold standard biomarker BNP improved  
456 diagnostic accuracy and specificity for ADHF diagnosis [99].

457 To gain insight into the pathology of valvular aortic stenosis (AS), plasma proteins from AS  
458 patients and controls were analyzed by 2D-DIGE followed by MS. In this analysis, crude and  
459 pre-fractionated plasma either depleted or equalized was analyzed leading to the  
460 identification of 36 differentially expressed proteins in AS. These were further clustered into

461 functional groups including protease inhibitors and proteases, blood homeostasis and  
462 coagulation, inflammation and immune response, lipid metabolism and transport and others  
463 [100].

464 Degraba et al. investigated the proteomic signature of serum of patients with carotid artery  
465 disease compared to healthy individuals in order to identify protein biomarkers associated  
466 with increased atherosclerotic risk. Using 2D-DIGE in fractionated serum, decreased levels of  
467  $\alpha(1)$ -antitrypsin (A1AT), Haptoglobin (HP) and Vitamin D binding protein (GC) and increased  
468 levels of  $\alpha(2)$ -glycoprotein precursor (AZGP1) in the subset of patients with symptomatic  
469 carotid atherosclerosis were detected [101]. Targeting to correlate CAD imaging findings  
470 with circulating biomarker expression, proteome analysis of the serum of 66 patients with  
471 stable or unstable angina and AMI after 3 and 6 months follow up was performed. Pro-  
472 inflammatory markers [6C Kine (CCL21), CTAK (CCL27), MIG (CXCL9) and Platelet Factor 4  
473 (PF4)], pro-coagulable markers [D-dimer (FDP), Platelet Factor 4 (PF4) and Hepatocyte  
474 Growth Factor (HGF)] and marker of shear stress and remodeling [Follistatin (FST)] all  
475 decreased over time whereas anti-apoptotic markers [PAI-10 (CXCL10) and I-309 (CCL1)  
476 chemokine] were up-regulated [26]. Han et al. employed 2DE coupled with MS to  
477 characterize protein expression patterns in the serum of CAD patients. Serum cyclin-  
478 dependent kinase 9 (CDK9) was found elevated in the serum with a concomitant increase in  
479 monocytes and artery plaque samples of CAD patients. Further analysis indicated CDK9 as a  
480 potential biomarker of atherosclerotic inflammation [102].

481 In an effort to identify biomarkers for ventricular dysfunction and HF, Watson et al. analyzed  
482 the coronary sinus serum from asymptomatic, hypertensive patients. By employing 2DE,  
483 they detected increased levels of Leucine-rich  $\alpha 2$ -glycoprotein (LRG) in the serum of the

484 patients with increased BNP. Further investigation in an independent cohort demonstrated  
485 significant association of LRG with HF, independent of BNP [103].

486 Cubedo et al. compared the serum proteome of new-onset AMI patients with healthy  
487 individuals to identify biomarkers of early stages of AMI. Applying 2DE followed by MALDI-  
488 TOF, decreased levels of Apolipoprotein J/clusterin (APOJ) within the first 6h after the onset  
489 of AMI accompanied by alterations in its glycosylation pattern were detected [104].  
490 Following the same approach, the same group compared the serum proteome from early  
491 and late phase AMI patients. Late phase AMI was associated with a decrease in immune  
492 response-inflammation proteins [ $\alpha$ -1B-glycoprotein (A1BG), Fetuin-A (AHSG), Complex-  
493 forming glycoprotein HC (AMBP), Complement C1r Subcomponent (C1R), Complement  
494 Component 3 (C3) and Factor-B (CFB)] and an increase in the levels of Serum amyloid P-  
495 component (SAP) in comparison to early AMI [105]. In a more recent study, the same group  
496 also found that decreased serum Retinol Binding Protein 4 (RBP4) levels are implicated in  
497 acute new-onset MI in male patients [106].

498 The detection of novel serum biomarkers could be also very helpful for both diagnosis and  
499 development of novel treatment modalities for transient ischemic attacks (TIA). To this end,  
500 George et al., attempted to identify novel serum TIA biomarkers through the application of  
501 mass spectrometry-based proteomics. The study highlighted Platelet basic protein (PBP) as a  
502 candidate TIA/minor stroke serum biomarker [107].

503 To identify biomarkers for acute aortic dissection (AAD), Gu et al., compared serum samples  
504 from AAD patients, AMI patients and healthy individuals using iTRAQ MS proteomics.  
505 Analysis of the differentially expressed proteins identified increased levels of Lumican (LUM)  
506 as a potential AAD-related serum marker [108].

507

### 508 **6.3 Urine**

509 Urinary proteomics benefits from the considerable stability of the urinary proteome and the  
510 establishment of large scientific networks [109], and hence has been applied in multiple  
511 studies also in the context of CVD (**Table 3**). Hou et al. applied 2D-DIGE for the analysis of  
512 urinary protein extracts of CHF patients and healthy donors. Among the twenty detected  
513 differentially expressed proteins, Orosomucoid 1 (ORM1) was selected based on  
514 bioinformatics analysis for further verification by Western blot analysis and ELISA. The  
515 authors conclude that increased levels of ORM1 may be used as potential novel urinary  
516 biomarker for the early detection of CHF [110].

517 Applying 1DE-LC-MS/MS, Lee et al. detected increased levels of monocyte antigen CD14 in  
518 the urine samples from CAD patients compared to controls. Furthermore, the proportion of  
519 CD14+ monocytes was found elevated in CAD patients compared to controls. The authors  
520 suggested that the increased release of CD14 in urine coupled with the elevated number of  
521 CD14+ monocytes in CAD patients may be associated to CAD severity [111].

522 Recently, efforts have been made towards employing multiple proteomic biomarkers, in  
523 order to more comprehensively depict complex disease pathophysiology. To this end, Delles  
524 et al., analyzed urine samples from 623 individuals with and without CAD by CE-MS. A panel  
525 of 238 CAD-specific urinary polypeptides with good sensitivity and specificity for CAD  
526 diagnosis was reported. The identified discriminatory polypeptides included fragments of  
527 alpha-1-antitrypsin (SERPINA1), Collagen types 1 and 3, Granin-like neuroendocrine peptide  
528 precursor (PCSK1N), Membrane-associated progesterone receptor component 1 (PGRMC1),  
529 Sodium/potassium-transporting ATPase gamma chain (FXVD2) and Fibrinogen-alpha chain  
530 (FGA) [13]. The same group, by following the same approach, recently demonstrated the  
531 value of the CAD238 panel as predictor of coronary events in asymptomatic subjects with



532 hypertensive atherosclerotic cardiovascular disease (HACVD) [112] and in the diagnosis of  
533 stable angina [113].

534 The development of urinary biomarker panels has also been applied targeting early  
535 detection of ischemic stroke specifically reliable detection of minor ischemic stroke or TIA,  
536 especially in cases of inconclusive brain imaging [14]. To this end, the urinary proteome of  
537 patients with acute stroke or TIA and controls with elevated cardiovascular risk was explored  
538 through the application of CE-MS. Candidate biomarkers were further identified following  
539 LC-MS/MS. A biomarker-based classifiers was developed that enabled differentiating  
540 between patients with acute stroke or TIA and controls [14]. Peptides of the discriminatory  
541 panels included FXFD domain-containing ion transport regulator 4 (FXFD4), Inter-alpha-  
542 trypsin inhibitor heavy chain H4 (ITIH4), Uromodulin (UMOD), Polymeric-immunoglobulin  
543 receptor (PIGR) and Collagen fragments.

544 A pilot study, using CE-MS based urine proteome analysis, enabled identification of a vast  
545 array (103) of HFREF-related urinary peptide biomarkers which mainly included fragments of  
546 fibrillar type I and III Collagen but also Fibrinogen beta (FGB) and alpha-1-antitrypsin  
547 (SERPINA1) peptides [15]. This opens the possibility of early diagnosis of HFREF even before  
548 the disease progresses to an overt symptomatic stage. Application of the same technology  
549 identified a set of HFREF-specific urinary peptide biomarkers on a background of chronic  
550 kidney disease (CKD). Consistent with the previous study, the majority of sequenced  
551 peptides were fragments of Collagen type I and III [114].

552 Urinary proteome-based classifiers have been assessed for their value in the early detection  
553 of asymptomatic left ventricular diastolic dysfunction (LVDD) in hypertensive patients. In one  
554 study, a set of urinary polypeptides was identified by CE-MS to distinguish hypertensive  
555 patients with overt HF from healthy controls: HF1 and HF2 are classifiers based on 85 and

556 671 urinary peptides, respectively, and were generated to distinguish between patients with  
557 LVDD and controls [115]. These classifiers were further assessed in a population study by  
558 analyzing the urinary proteome by CE-MS [116]. The same group, in a subsequent study  
559 investigated whether HF1 classifier predicts cardiovascular end points in a population of 791  
560 randomly recruited normal subjects over a 5 year follow-up period. The urinary proteomic  
561 signature was found to predict CVD incidence at higher accuracy than systolic pressure [117].  
562 While the advantage of CE-MS in the management of CVD is very promising and attributed  
563 not only to the robustness of the technique (i.e. inter-laboratory variability, stability,  
564 interference with drugs) [118], but also to its multiple applications in patient management  
565 and drug development [119], it has a low urine loading capacity and is only suitable for small  
566 proteins (< 20 kDa) [119]. CE-MS identifies naturally occurring peptides with distinct c- and  
567 n-terminal that enables a specific evaluation of the disease pathology. Frequently a highly  
568 significant change in abundance of specific collagen fragments is detected. The most likely  
569 hypothesis today is that these specific peptides reflect altered protease activity which has a  
570 specific impact on collagen (as well as likely multiple other proteins), and which is associated  
571 with several pathologies [120].

572

#### 573 **6.4 Circulating Cells**

574 Circulating cells and vesicles may play a major role in CVD pathophysiology. Several  
575 proteomics studies targeting these specimens have been reported, summarized in **Table 4**.

576 Polymorphonuclear neutrophils (PMNs) play a key role in the pathophysiology of AAA  
577 progression [121,122], and as such, Ramos-Mozo et al., comparatively analyzed their  
578 proteome in AAA patients and controls. Using 2D-DIGE-MALDI MS, they identified decreased

579 levels of Catalase (CAT) further confirmed by an apparent decrease activity of Catalase in  
580 circulating PMNs and plasma in AAA patients [123].

581 To gain insight into the association of circulating cells with coronary atherosclerosis,  
582 Bleijerveld et al. performed in-depth proteomic profiling of granulocytes in a cohort of  
583 patients suffering from chronic (sub)total coronary occlusion in comparison to matched  
584 control patients. Using stable isotope peptide labeling and 2DE LC-MS/MS, 57 candidate  
585 biomarker proteins were identified including Bactericidal/permeability-increasing protein  
586 (BPI), Ficolin-1 (FCN1), Charcot–Leyden crystal protein (CLC), Eosinophil granule major basic  
587 protein (MBP) and Eosinophil-derived neurotoxin (EDN). Following further verification in an  
588 independent cohort by label-free proteome analysis, the down-regulation of  
589 Bactericidal/permeability-increasing protein (BPI) in circulating granulocytes was proposed  
590 as a promising biomarker for severe atherosclerotic coronary stenosis [124].

591 To investigate the profiles of macrophages from AAA patients and peripheral arterial  
592 occlusion (PAO) patients without AAA, transcriptomics and proteomics approaches were  
593 followed. Differentially expressed proteins were identified through the application of 2D-  
594 DIGE followed by MALDI TOF. An antibody protein array was used to validate selected  
595 proteins found (or predicted) to be differentially expressed in macrophages and plasma  
596 based on the transcriptomic and proteomic analysis. TIMP-3, ADAMTS5, and ADAMTS8 were  
597 found differentially expressed between the macrophages and plasma of AAA and PAO  
598 patients [125].

599 Changes in the proteome profile of circulating endothelial cells (CECs) and EPCs have also  
600 been described in ACS. CECs and EPCs were isolated by flow cytometry from blood obtained  
601 from ACS patients and control subjects. Proteome analysis by LC-MS/MS identified  
602 differences in EPC and CEC proteins between control and ACS patients, predicted to

603 segregate into 6 molecular pathways (5HT4 type receptor mediated signaling pathway, 5HT3  
604 type receptor mediated signaling pathway, 5HT1 type receptor mediated signaling pathway,  
605 Adrenaline and noradrenaline biosynthesis and Heterotrimeric G-protein signaling pathway-  
606 rod outer segment phototransduction exclusively represented in ACS). Although this study  
607 indicated the potential of CEC and EPC to reflect CVD, the development of a “gold standard”  
608 protocol for their isolation is imperative for their further implementation in biomarker  
609 development and diagnostics [53].

610

## 611 **6.5 Extracellular vesicles**

612 Membrane microvesicles (MVs) are released into the circulation from activated cells, such as  
613 platelets, they mediate intercellular communication, and their number is increased in  
614 patients with ACS [126]. To develop a unique panel of proteins discriminating patients with  
615 STEMI from stable CAD controls, Velez et al. performed comparative proteome analysis in  
616 plasma MVs based on 2D-DIGE MS. An up-regulation of  $\alpha$ 2-macroglobulin isoforms,  
617 Fibrinogen, and Viperin (RSAD2) was detected in MVs from STEMI patients [54].

618 In a recent study by Cheow et al. the proteome profile of plasma EVs derived from patients  
619 with MI in comparison to EVs from patients with stable angina was analyzed by LC-MS/MS. A  
620 biomarker panel for MI detection was developed, comprising of six up-regulated proteins:  
621 Complement C1q subcomponent subunit A (C1QA) and Complement C5 (C5), both  
622 associated with complement activation; Apolipoprotein D (APOD) and Apolipoprotein C-III  
623 (APOC3), implicated in lipoprotein metabolism; and Platelet glycoprotein Ib alpha chain  
624 (GP1BA) and Platelet basic protein (PPBP), both related to platelet activation [127].

625 Martinez-Pinna et al. employed label-free LC-MS for the quantitative proteome analysis of  
626 plasma-derived microvesicles (exosomes and microparticles) from AAA patients and control

627 subjects. Differentially expressed proteins not previously associated with AAA were  
628 detected. Among the proteins upregulated in exosomes from AAA patients were Ferritin  
629 light chain (FTL), C-reactive protein (CRP) and Platelet factor 4 (PLF4) whereas Dermcidin  
630 (DCD), Annexin A2 (ANXA2) and Oncoprotein-induced transcript 3 protein (OIT3) increased in  
631 microparticles of AAA patients [128].

632 The proteome of plasma-microparticles has also been investigated in DVT through the  
633 application of 2D MALDI MS/MS. Galectin-3 binding protein precursor (LGALS3BP) and  
634 Alpha-2 macroglobulin (A2M) were significantly enriched in DVT patients whereas among  
635 others Alpha-1-antitrypsin precursor (SERPINA1), Histidine-rich glycoprotein precursor  
636 (HRG), Hemopexin precursor (HPX), Fibrinogen beta chain precursor (FGB), Isoform Gamma-  
637 B of Fibrinogen gamma chain precursor (FGG), Fibronectin 1 isoform 4 preproprotein (FN1),  
638 and Rheumatoid factor RF-IP16 (RF) were reduced in DVT versus controls [129].

639

## 640 **6.6 Platelets**

641 Banfi et al. compared the proteome of isolated platelets from patients with stable angina or  
642 non-ST elevation ACS compared to control subjects with no history of CAD, using 2DE-MS. Six  
643 differentially expressed proteins were identified associated with energy metabolism [2-  
644 oxoglutarate dehydrogenase (OGDH), and Lactate dehydrogenase (LDH)]; cytoskeleton-  
645 based processes [Gamma-actin (ACTG1), Coronin 1B (CORO1B), and Pleckstrin (PLEK)]; or  
646 involved in protein degradation [Proteasome subunit type 8 (PSMB8)] [130]. Comparing the  
647 platelet proteome from patients with ACS to that of patients with stable CAD with 2DE-MS,  
648 the levels of proteins involved in cellular cytoskeleton [F-actin capping (CAPZA2),  $\beta$ -tubulin  
649 (TUBB),  $\alpha$ -tubulin isotypes 1 and 2, Vinculin (VCL), Vimentin (VIM) and two Ras-related  
650 protein Rab-7b isotypes (RAB7B)], glycolysis [Glyceraldehyde-3-phosphate dehydrogenase

651 (GAPDH), Lactate dehydrogenase and two pPruvate kinase isotypes]; cellular-related  
652 antioxidant system (Manganese superoxide dismutase), cell survival [Proteasome subunit  $\beta$   
653 type 1 (PSMB1)] and the expression and activity of Glutathione-S-transferase were  
654 significantly reduced in platelets from ACS patients compared to CAD patients [131].  
655 Platelets from patients with non-ST segment elevation ACS versus stable CAD controls were  
656 analyzed by 2DE-MS. The identified differentially expressed proteins were involved in  
657 cellular cytoskeleton [Actin Cytoplasmatic-1 (ACTB), Alpha-actinin-1 (ACTN1), Caldesmon  
658 (CALD1), F-actin-capping protein subunit beta (CAPZB), Filamin-A (FLNA), Myosin-9 (MYH9),  
659 Talin-1 (TLN1), Tropomyosin alpha chain 3 (TPM3), Zyxin (ZYG1)], signaling [Adenylyl cyclase-  
660 associated protein 1 (CAP1), FYN-binding protein (FYB), Integrin-linked protein kinase (ILK),  
661 Proto-oncogene tyrosine-protein kinase Src (SRC), Rho GDP-dissociation inhibitor 2  
662 (ARHGDI1)] and ECM [Serum Albumin, Secreted protein acidic and rich in cysteine (SPARC)],  
663 vesicles/secretory trafficking pathway [Dynamin-1-like protein (DNM1L), Ras-related protein  
664 Rab-27B (RAB27B), -6B (RAB6B), -11A (RAB11A)] [132].

665 Alterations in the platelet proteome have also been associated with HFpEF. Raphael et al.  
666 analyzed the proteome of platelets from three groups: 1) patients hospitalized with  
667 symptoms of HFpEF, 2) the same subjects several weeks later without symptoms and 3)  
668 control subjects. Among the 6102 proteins identified by mass spectrometry, S100A8 was  
669 found to be consistently expressed in HFpEF patients compared to controls. The levels of this  
670 protein were also found increased in the plasma of subjects with HFpEF in an external  
671 cohort. To investigate whether S100A8 is causal or only associated with the disease, its  
672 effects were further assessed in human induced pluripotent stem cell-derived  
673 cardiomyocytes. The results indicated direct effects of S100A8 on the electrophysiological

674 and calcium handling profile and suggest that this protein may be causally contributing to  
675 HFpEF [133].

676

## 677 **7. Factors with an impact on cardiovascular biomarkers**

### 678 **7.1 Sex and ethnicity**

679 Alterations in the plasma levels of CVD biomarkers are also associated with sex and ethnic  
680 differences and may have value in individualized CVD risk assessment. In a Mayo Clinic study,  
681 uniplex and multiplex assays were used to measure the circulating levels of 47 candidate  
682 CVD markers in men and women of African-American (AA) and non-Hispanic White (NHW)  
683 ethnicity. Female gender was associated with higher levels of inflammatory markers,  
684 adipokines, lipoproteins, natriuretic peptides, vasoconstrictor peptides and markers of  
685 calcification and thrombosis. AA ethnicity was associated with higher levels of inflammatory  
686 markers, leptin, vasoconstrictor-antidiuretic peptides and markers of calcification and  
687 thrombosis and with lower levels of adiponectin and vasodilator-natriuretic peptides [134].

### 688 **7.2 Diet**

689 Polyphenol rich diets are associated with reduced risk of CVD, hence their potential effects  
690 on CVD biomarkers may be relevant. A pilot study was conducted to detect quantitative  
691 differences in the urinary CAD biomarkers (CAD238 [13]) caused by short-term consumption  
692 of a polyphenol rich (P-R) drink. A group of overweight healthy subjects were randomized to  
693 P-R drink or placebo for two weeks. Following urinary analysis by CE-MS, 27 polypeptides  
694 that displayed more than 4-fold difference between the two groups were detected. Among  
695 these, seven were included in CAD238, five of which [Fibrinogen alpha chain (FGA) [607-  
696 622], Collagen alpha-1(I) chain (COL1A1) [543-588], Collagen alpha-2(V) chain (COL1A2)  
697 [1209-1225], Xylosyltransferase 1 (XYLT1) [51-66], Ig kappa chain C region (IGKC) [9-23]]

698 changing towards the healthy profile while two of them [Alpha-1-antitrypsin (SERPINA1)  
699 [276-295] and Collagen alpha-1(I) chain (COL1A1)[1095-1106]] changing towards CAD. Based  
700 on these results the authors suggested that P-R drink may have beneficial effects in CAD  
701 prevention [135]. More recently, the same group, in an effort to evaluate the impact of olive  
702 oil (OO) consumption in CVD prevention, investigated the impact of diet supplementation  
703 with OO, either low or high in phenolics, on urinary proteomic biomarkers (CAD238 panel). A  
704 significant improvement in the scoring of the CAD238 panel after supplementation with OO  
705 for 6 weeks was demonstrated, indicating a benefit of OO in preventing CVD [136].

### 706 **7.3 CKD**

707 CVD could also be a secondary endpoint in patients with CKD disease. To address this issue,  
708 Schiffer et al., performed a diagnostic phase I/II study in which CE-MS was applied to analyze  
709 plasma specimens from CKD stage 5D patients suffering from vascular disease. Comparative  
710 analysis identified 13 novel biomarkers for CVD of which four were identified by tandem MS  
711 as fragments of Collagen alpha-1 type I and III and one as fragment of Apolipoprotein C3  
712 (APOC3). The markers were validated in an independent blinded cohort and enabled  
713 distinguishing mild and severe CVD with good sensitivity and specificity, suggesting that this  
714 specific polypeptide pattern in the plasma of CKD patients reflects CVD [137].

715

### 716 **7.4 Ageing**

717 Urinary proteomics was also employed to investigate the proteomic transition from normal  
718 ageing to age-related pathological complications including CVD. Using CE-MS in a study  
719 including over 10000 subjects, 112 age-correlated peptides (mainly originated from collagen,  
720 uromodulin and fibrinogen) were identified. Pathway analysis revealed perturbations in  
721 collagen homeostasis, trafficking of toll-like receptors and endosomal pathways being



722 associated with ageing in general, whereas increased degradation of insulin-like growth  
723 factor-binding proteins (IGFBPs) was observed in pathological ageing only [138].

724

## 725 **7.5 Hemodialysis**

726 Plasma proteomic analysis of the HDL fraction may enable identification of candidate  
727 biomarkers associated with cardiovascular risk in end-stage renal disease (ESRD) patients  
728 undergoing hemodialysis. To this end, Mange et al. investigated the proteome of HDL  
729 isolated from plasma of hemodialysis patients at high risk for CVDs and healthy volunteers.  
730 Using iTRAQ labeling and nano-LC/MS/MS analysis, forty proteins differentially expressed in  
731 the two groups were identified. These proteins were primarily involved in lipid metabolism,  
732 inflammation, complement activation and metal cation homeostasis. Among them,  
733 Apolipoprotein C2 (APOC2) and Apolipoprotein C3 (APOC3) were found elevated in  
734 hemodialysis patients whereas Serotransferrin was reduced after validation in an  
735 independent population set. The authors conclude that the identified proteins are linked to  
736 HDL dysfunction in chronic hemodialysis patients [139].

737

## 738 **8. Data integration in CVD**

739 The advantage of integrating different omics approaches has previously been shown in the  
740 management of chronic kidney disease [140]. Not only is the identification of a broader  
741 range of biomarkers possible, but also the systemic understanding of related molecular  
742 mechanisms is achievable, both contributing to improving disease management [3,4].  
743 Towards this end, we performed gene ontology analysis of all the biomarkers summarized in  
744 the tables of this review through Cytoscape. Inflammation (acute, humoral, complement

745 activation etc.), wound healing and coagulation (platelet activation, degranulation, fibrin clot  
746 formation), proteolysis and extracellular matrix organization, and handling of cholesterol  
747 and LDL were among the most significantly pathways and biological functions reflected by  
748 the proteomic changes observed in CVD (data provided in supplementary table 2).

749 First reports on combining different omics traits were published recently. The integration of  
750 genomics and metabolomics has been explored in CVD and findings have shown an added  
751 value in the approach [141]. An integrated metabolomics and genome-wide association  
752 study (GWAS) has revealed that single nucleotide polymorphisms can explain variances  
753 observed with some metabolite levels [142]. Through the integration of the transcriptomic  
754 with proteomic findings from myocardial tissues global changes in dilated cardiomyopathy  
755 (DCM) in genes, proteins and pathways involved in cardiac function were unraveled [143]. A  
756 network biology approach that combined genomic and proteomic data was developed and  
757 distinguished pathological from physiological left ventricular hypertrophy (LVH) [144]. In  
758 another study, genetic, transcriptome and protein analyses of titin truncations provided  
759 evidence into the mechanisms of end-stage DCM [145]. A strategy combining proteomics  
760 with metabolomics was applied in human cardiac tissues from individuals with normal heart  
761 or patients with ischaemic (ISCM) and idiopathic dilated cardiomyopathies (IDCM). This  
762 study enabled the characterization of the differential regulation of molecular pathways and  
763 metabolism in ISCM and IDCM for the first time [146]. Apart from the potential to elucidate  
764 mechanisms of disease pathogenesis, the integrative omics approaches also hold promise  
765 for personalised medicine [147]. Combined, the published reports suggest that systems  
766 biology approaches underpinned by omics strategies in animal models and patients have the  
767 potential to aid in the dissection of the complicated networks interfering in cardiovascular  
768 pathophysiology and lead to the development of novel diagnostic and therapeutic

769 modalities [119,148-150]. However, although the benefits of omics integration are quite  
770 obvious, it is very challenging to achieve a successful integration. A major challenge  
771 associated with data integration is the heterogeneity of data generated by omics  
772 technologies. There is clearly an urgent need to develop robust algorithms that will facilitate  
773 this process.

## 774 **9. Expert commentary**

775 Significant progress towards the development of state of the art proteomic technologies  
776 applicable in the context of CVD has been made. These advancements include improvement  
777 of sample preparation methods for detection even of the low-abundance proteins as well as  
778 the development of high-throughput proteomic profiling techniques that enable the  
779 detection of multiple candidate biomarkers rapidly and with high accuracy. Although early  
780 studies focused mainly on tissue and plasma proteomics, recent studies also targeted other  
781 biological sources such as urine, extracellular vesicles and sub-cellular proteomes for  
782 biomarker discovery. The number of potential biomarkers that have arisen through the  
783 application of proteomic technologies has substantially increased in the past few years. In a  
784 considerable number of studies, efforts have been put on unraveling panels of proteomic  
785 biomarkers. These appear to be of higher stability and accuracy than single biomarkers, since  
786 panels are better suited to display the complex disease pathophysiology [151].

787 However, we also see substantial shortcomings in unfortunately many studies. As outlined  
788 by us and others in detail (e.g. [152-154]), the mere detection of the association of a protein  
789 with disease is insufficient to support claims for biomarkers. As a minimum a potential  
790 biomarker must have a defined context-of-use and demonstrate a significant improvement  
791 of the current state-of-the-art, in the population it is intended to be used. Unfortunately,  
792 many studies fall short of this minimal requirement, and consequently the biomarker value

793 and reliability of findings cannot be readily attributed. In addition, although other factors  
794 and diseases, like diabetes type 2 and CKD, have significant impact on cardiovascular  
795 biomarkers and should be taken into consideration when performing such studies, this is  
796 frequently not attempted. Progress has been made in recent years in the development of  
797 proteomics technologies. Application of these novel approaches in the discovery process  
798 may result in additional, novel findings. In addition, major efforts should be directed towards  
799 actual assessment of the value of existing biomarkers for a specific context of use, in the  
800 relevant population, and in the light of the current state-of-the-art (currently routinely used  
801 biomarkers and pertinent demographic information). This has been outlined several years  
802 ago for bladder cancer [155], but the exact same considerations hold true for cardiovascular  
803 disease.

804

#### 805 **10. Five-year view**

806 Summarizing the recent progress in the discovery of proteomic biomarkers for CVD, it is  
807 obvious that despite the plethora of studies performed in this field, only few of them appear  
808 close to clinical implementation. A significant number of studies stops at the discovery phase  
809 without subsequent validation of the findings. Since many of the studies cited in this review  
810 are very recent, we hope for a rapid progress in biomarker validation the next years.

811 Nevertheless, some biomarker panels have arisen the recent years holding promise for  
812 better management of CVD based on reported validation studies. It appears that the most  
813 relevant context of use for proteomics biomarkers in CVD is enabling early detection and  
814 prognosis.

815 The next step for these panels is to be validated in blinded studies and subsequently  
816 approved by the regulatory authorities for clinical application. An example is the application

817 of a proteomic biomarker panel for CKD detection: the panel, CKD273, was developed based  
818 on over 600 samples [156], and subsequently evaluated and validated in multiple studies  
819 [157-159]. As a result, this urinary proteome-based panel is now being used in patient  
820 stratification in PRIORITY, a large multicentric randomized controlled trial [160,161] and has  
821 also received a letter-of-support from the US-FDA  
822 (<http://www.fda.gov/downloads/Drugs/DevelopmentApprovalProcess/UCM518268.pdf>).

823 As depicted in **Figure 4**, the current state-of-the-art allows assessment of CVD with high  
824 confidence at late stage, when significant and frequently irreversible organ damage has  
825 occurred. Hence it seems biomarkers enabling detecting established disease are of very  
826 little, if any value. The potential of proteomic biomarkers appears in the early detection of  
827 disease, when disease-initiating molecular processes are already ongoing, but clinical  
828 symptoms are not detectable. This is also supported by the thoughts outlined in **Figure 1**:  
829 the hypothesis that proteins are the cause of CVD onset and progression. At an early stage  
830 also intervention will be most effective, ideally even preventing onset of disease.

831 Such approaches also depend on a holistic understanding of CVD processes, which will be  
832 one of the biggest challenges in forthcoming studies. In the near future, the development of  
833 systems biology approaches through the integration of omics technologies will provide a  
834 more systematic insight into CVD [3,4]. Integration of the proteomics data with findings from  
835 other omics approaches and bioinformatics analysis will pave the way for the development  
836 of novel biomarker panels detecting early stages of the disease leading to CVD prevention. A  
837 new era of proteomics towards personalized cardiovascular medicine will hopefully begin  
838 soon.

839

## 840 **11. Key issues**

- 841 • CVD remains the major cause of morbidity and mortality worldwide, knowledge on  
842 molecular mechanisms and potential drug targets as well as validated biomarkers  
843 guiding intervention are urgently required.
- 844 • Proteomics has the ability to allow insights into the mechanisms underlying  
845 cardiovascular disease, should enable defining disease on a molecular level.
- 846 • Advancements and development of state of the art proteomics technologies have  
847 facilitated the high-throughput analysis of many and different samples.
- 848 • Progress has been also made the recent years towards the development of  
849 biomarker panels, demonstrating superiority over single biomarkers.
- 850 • A list of promising results and multiple potential biomarkers exists, but has to be  
851 investigated and validated in detail to have an actual impact, improve patient care in  
852 the near future.
- 853 • Other factors and systemic disorders have an impact on cardiovascular biomarkers  
854 and therefore should be considered when analyzing data from discovery and  
855 validation phases.
- 856 • Verification of the proteomic findings in appropriately powered studies appears  
857 imperative now, to receive approval from the regulatory authorities and enable  
858 implementation.

859

860 **Figure 1: Graphic depiction of the information obtained from the different approaches.**

861 Histology can give information on microscopic structural changes. Subcellular structures, and  
862 even more the molecules involved in the molecular pathology can generally not be assessed.  
863 In contrast, proteome analysis does not give information on morphological changes, but  
864 gives information on global protein changes, which can be associated with the molecular  
865 changes in disease. Proteins represent the most appropriate targets of therapeutic drugs,  
866 hence, proteome analysis can give guidance on the molecular structures to be targeted in  
867 therapy, an information that can generally not be obtained from histology. Adapted with  
868 permission from [162].

869

870 **Figure 2: Graphical depiction of the main two different routes clinical proteomics**  
871 **approaches can take: towards biomarker discovery, or a Systems Medicine approach**

872 **towards "modelling disease":** The biomarker approach (left) requires identification of  
873 distinct potential biomarkers that are subsequently verified, and then applied in the  
874 appropriate clinical setting. If benefit can be demonstrated, they should be  
875 implemented/routinely applied. The Systems medicine approach takes advantage of the  
876 breadth of data, including literature sources, aims at molecular modelling of disease and  
877 predicting key structures. These are then verified (first valid result), and subsequently their  
878 value as potential targets is investigated in appropriate interference studies. If positive,  
879 drugs can be developed, tested, and, if beneficial, should be applied. Starting with the same  
880 or similar data, the downstream utilization (and the associated issues) is quite different, but  
881 ultimately in both cases we aim at investigating proteomics to improve patient care.

882

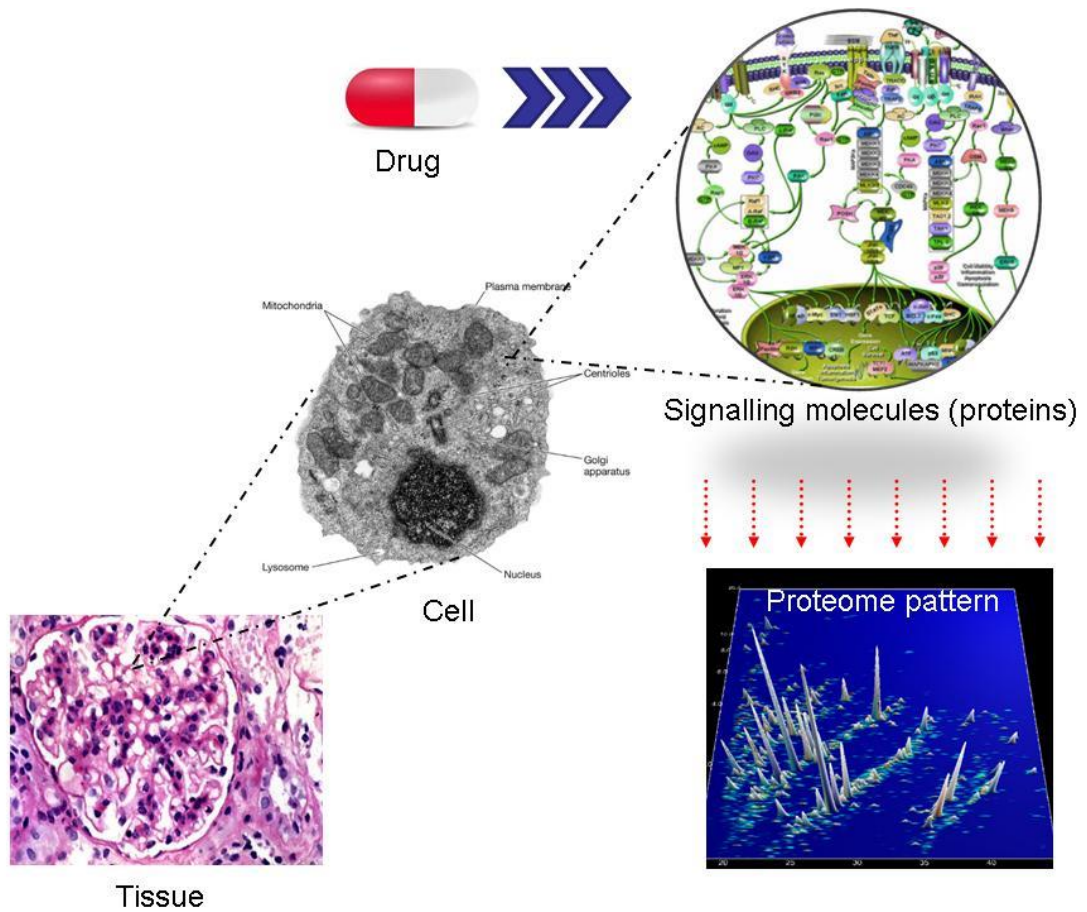
883 **Figure 3. Schematic representation of the workflow followed in cardiovascular proteomic**  
884 **studies.** A plethora of biological sources including arterial tissue, cardiac tissue, tissue  
885 secretome, blood, urine, thrombus, circulating cells and platelets have already been used to  
886 investigate the proteome alterations in cardiovascular disease (CVD). Analysis of these  
887 proteomes through global proteomics (discovery phase) and targeted proteomics (validation  
888 phase) approaches will make possible the identification of novel candidate biomarkers for  
889 better management of CVD. Of note, there are four biomarkers recommended for clinical  
890 use: 1) Cardiac Troponin T and Troponin I, 2) B-type natriuretic peptide (BNP), 3) C-reactive  
891 protein (CRP) and D-dimer, 4) Apolipoprotein A-I.

892

893 **Figure 4. Early diagnosis and/or prognosis of diseases improves chances for a better**  
894 **outcome for the patients.** The initiation of molecular processes that result in (chronic)  
895 diseases can be detected based on the decisive molecular changes, using proteomic  
896 technologies, substantially prior to advanced organ damage. In this early stage, molecular  
897 changes could be reverted, and onset of clinically relevant disease could be prevented. As a  
898 minimum, further progression can be slowed down by applying of appropriate therapeutic  
899 interventions and/or changes to the lifestyle. At a later stage, the disease becomes clinically  
900 evident, as a result of the organ damage. However, at this point in time curative treatment is  
901 not possible anymore, only disease progression can possibly be delayed to some extend  
902 (although with moderate success, as also evident by the multiple trials that failed). Reprinted  
903 with permission from [163].

904

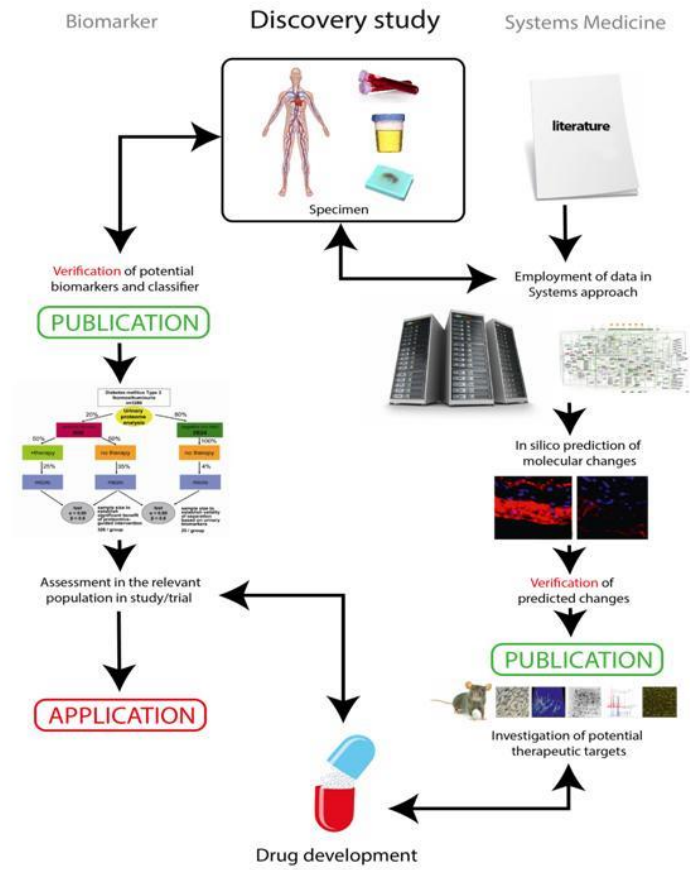




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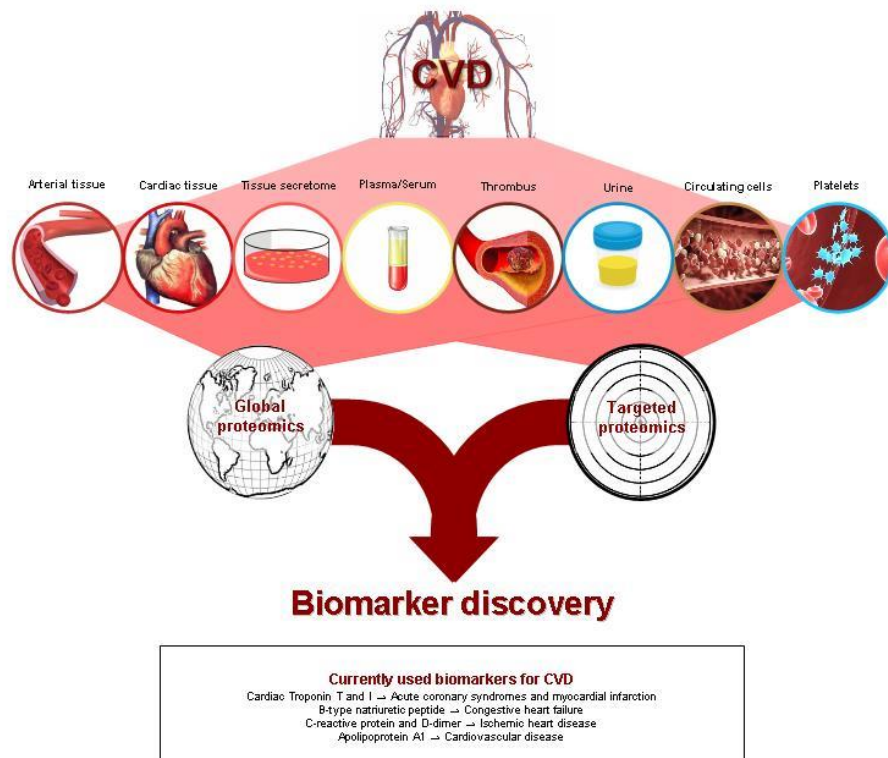
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Tissue

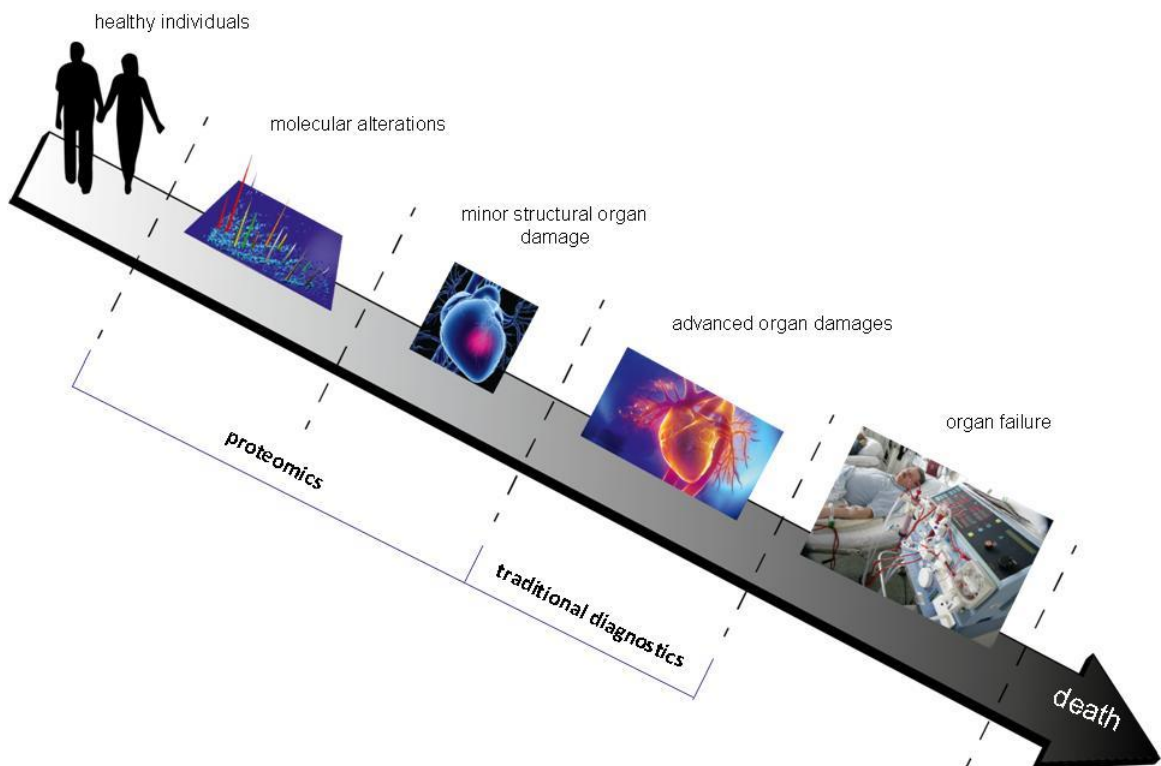


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911 **References**

912 **Reference annotations**

913 **\* Of interest**

914 **\*\* Of considerable interest**

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