**The dynamic response of IFI16 and PML Nuclear Body components to HSV-1 infection**.

Roger D. Everett

MRC - University of Glasgow Centre for Virus Research, University of Glasgow Scotland, U.K.

Running Head: Dynamic response of IFI16 to HSV-1 infection

Address correspondence to Roger Everett, roger.everett@glasgow.ac.uk

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**Abstract**

Intrinsic immunity is an aspect of antiviral defence that operates through diverse mechanisms at the intracellular level through a wide range of constitutively expressed cellular proteins. In the case of herpesviruses, intrinsic resistance involves the repression of viral gene expression during the very early stages of infection, a process that is normally overcome by viral tegument and/or immediate-early proteins. Thus the balance between cellular repressors and viral counteracting proteins determines whether or not a cell becomes productively infected. One aspect of intrinsic resistance to herpes simplex virus type 1 (HSV-1) is conferred by components of PML Nuclear Bodies, which respond to infection by accumulating at sites that are closely associated with the incoming parental HSV-1 genomes. Other cellular proteins also respond to viral genomes in this manner, including IFI16 which has been implicating in sensing pathogen DNA and initiating signalling pathways that lead to an interferon response. Here, studies of the dynamics of the response of PML NB components and IFI16 to invading HSV-1 genomes demonstrate that this response is extremely rapid, occuring within the first hour after addition of the virus, and that hDaxx and IFI16 respond more rapidly than PML. In the absence of HSV-1 regulatory protein ICP0, which counteracts the recruitment process, the newly formed, viral genome induced PML NB-like foci can fuse with existing PML NBs. These data are consistent with a model involving viral genome sequestration into such structures thereby contributing to the low probability of initiation of lytic infection in the absence of ICP0.

**Importance**

Herpesviruses have intimate interactions with their hosts, with infection leading either to the productive lytic cycle or to a quiescent infection in which viral gene expression is suppressed while the viral genome is maintained in the host cell nucleus. Whether a cell becomes lytically or quiescently infected can be determined through the competing activities of cellular repressors and viral activators, some of which counteract cell mediated repression. Therefore the events that occur within the earliest stages of infection can be of crucial importance. Using live cell microscopy, this paper describes the extremely rapid response to herpes simplex virus type 1 infection of the cellular protein IFI16, a sensor of pathogen DNA, and also the PML Nuclear Body proteins PML and hDaxx. The data imply that these proteins can accumulate on or close to the viral genomes in a sequential manner which may lead to their sequenstration and repression.

## INTRODUCTION

Whether or not a cell becomes productively infected with herpes simplex virus type 1 (HSV-1), as with other herpesviruses, depends on many factors that modulate the initial stages of infection. Amongst these are cellular proteins that respond in a restrictive manner to repress viral gene expression once the viral genomes have entered the nucleus, while the virus expresses proteins that counteract these repressive effects or stimulate viral gene expression more directly. Over the last decade, it has become clear that one class of restricting cellular factors comprises a number of components of promyelocytic leukaemia (PML) Nuclear Bodies (PML NBs, also known as ND10), including PML itself, Sp100, hDaxx and ATRX (reviewed in (1-3)). The HSV-1 Immediate-Early protein ICP0 is responsible for overcoming restriction mediated by these proteins through mechanisms that require its E3 ubiquitin ligase activity (reviewed in (1)). HSV-1 mutants that are unable to express active ICP0 have a very low probability of initiating lytic infection in restrictive cell types (4-6), but are able to replicate more efficiently in cells depleted of one or more of these PML NB proteins (7-10).

There is considerable evidence that the restrictive effects of PML NB components depend on their dynamic response to infection. PML, Sp100 and hDaxx are recruited to sites that are closely associated with HSV-1 genomes during the earliest stages of infection (11, 12) by mechanisms that involve sumoylation and/or their ability to interact with sumoylated proteins, and which are inhibited by ICP0 (7-9, 13). It is likely that other cellular proteins that accumulate on or near HSV-1 genomes in a SUMO pathway dependent manner will be identified in the future, and because ICP0 causes a wide ranging reduction in the levels of sumoylated species during infection (14-16) their recruitment may also be sensitive to ICP0. Interestingly, although PML is required for the assembly of PML NBs in uninfected cells (17, 18) it is not required for recruitment of either hDaxx or Sp100 to viral genomes, and indeed these proteins may be recruited independently (7-9). Recruitment defective mutants of PML and hDaxx, unlike their wild type counterparts, are unable to reverse the stimulatory effects on ICP0 null mutant HSV-1 replication of shRNA mediated knock-down of the endogenous proteins (9, 13, 19).

The signals that initiate the recruitment of PML NB components to HSV-1 genomes remain unknown. It is possible that it is related to a DNA repair response, and indeed several DNA repair proteins respond to infection in a similar manner (20), but recruitment of PML still occurs in DNA repair deficient cells (20). More recently, the cellular DNA sensor IFI16 (21, 22) has also been implicated in the initiation of pathways that are inhibitory to HSV-1, either through initiating events leading to an interferon response (22, 23) or more direct

effects on viral gene expression (24-27). ICP0 null mutant HSV-1 has an increased plaque forming potential in cells depleted of IFI16 (24, 26). IFI16 is degraded during HSV-1 infection by a mechanism that was originally thought to be ICP0-dependent (23), although later work showed that ICP0 was neither sufficient nor necessary for this degradation (24, 28). Indeed, cellular factors (and cell type) also contribute to the stability of IFI16 during HSV-1 infection (24, 25, 27). Nonetheless during a normal experimental infection IFI16 is degraded more rapidly during a wild type (wt) than an ICP0 null mutant infection, unless the latter is conducted at very high multiplicity. Like the PML NB components, IFI16 also localizes to HSV-1 genomes during the early stages of infection (23-29), and ICP0 inhibits this process (24).

In order to study the dynamic responses of this group of cellular proteins during the very earliest stages of HSV-1 infection, analyses were conducted in live cells expressing near endogenous levels of PML, hDaxx or IFI16, or combinations thereof, tagged with EYFP, ECFP or a dual fusion protein that expresses red fluorescence constitutively and in addition green fluorescence after photoactivation. These studies revealed the extremely rapid and in some cases transient responses of these proteins during HSV-1 infection. Within the first hour after addition of the virus, IFI16 was observed to form distinct but highly transient foci that are almost certainly associated with HSV-1 genomes. This occurred with both wt and ICP0-null mutant viruses, but in the latter case a second and more stable phase of recruitment of IFI16 was observed. Recruitment of IFI16 to HSV-1 genomes was observed with equal efficiency in PML-depleted cells. Efficient recruitment of hDaxx to these foci also occurred very rapidly, but that of PML took place more slowly. These events occur in both wt and ICP0 null mutant infections, but were very transient in the former, presumably because of the effects of ICP0. Photoactivation experiments illustrated that hDaxx molecules in one part of the cell nucleus could be recruited to viral genomes in the opposite half of the nucleus within seconds. These studies reveal highly dynamic and in some cases sequential or transient events of cellular protein recruitment to HSV-1 genomes that would not be amenable to study by fixed cell methods. Importantly, the results reveal that the cell responds to the entry of viral genomes into the nucleus certainly within minutes and probably within seconds, long before a cell can be detected as being infected through the production of viral proteins.

## MATERIALS AND METHODS

### Viruses and cells

HSV-1 strain 17+ was the wt strain used, from which the ICP0-null mutant *dl*1403 was derived (5). Virus *dl*0C4, a derivative of *dl*1403 that expresses ECFP-linked ICP4, was constructed as described (4). All viruses were grown in BHK cells and titrated in U2OS cells, in which ICP0 is not required for efficient HSV-1 replication (6). Human diploid fibroblasts (HFs), PML-depleted HFs (7), telomerase immortalized HFs (HFTs, a gift from Chris Boutell), U2OS and HEK-293T cells were grown in Dulbecco's Modified Eagles' Medium supplemented with 10% fetal calf serum (FCS). BHK cells were grown in Glasgow Modified Eagles' Medium supplemented with 10% new born calf serum and 10% tryptose phosphate broth. HepaRG cells were grown in William's Medium E supplemented with 10% fetal bovine serum Gold (PAA Laboratories Ltd), 2 mM glutamine, 5 μg/ml insulin and 500 nM hydrocortisone. All cell growth media were supplemented with 100 units/ml penicillin and 0.1 mg/ml streptomycin. Lentivirus transduced cells were maintained with continuous antibiotic selection, as appropriate.

### Lentiviral vectors

Lentiviral vectors expressing EYFP-linked PML isoform I or hDaxx from the weak HSV-1 glycoprotein D promoter and including G418 resistance have been described previously (9, 13, 19). Derivatives of these were constructed in which the G418 resistance marker was replaced by puromycin resistance, while versions were also constructed in which ECFP was used in place of EYFP. The same backbone was used to express EYFP-linked IFI16 using a cDNA purchased from Origene. IFI16 mutant m3 (S27A/L28A/D50A; (30)) was constructed by replacement of the wt cDNA with the complete m3 cDNA made by PCR from a plasmid supplied by Jungsan Sohn. Derivative ΔHIN2 which lacks IFI16 codons 518 – 729 was constructed by PCR splicing and replacement of the wt sequence. Lentiviral vectors expressing PML isoform I or hDaxx linked to a fusion of photoactivatable EGFP and constitutively fluorescent mCherry were constructed by replacement of the EYFP coding region of vectors pLNGY-PML.I and pLNGY-hDaxx with the dual fluorescent (GAPC) cassette (31). Lentivirus transduction, selection of transduced cells and maintenance of cell lines were as described previously (7). Sequential transduction was used to prepare cells expressing EYFP-IFI16 with either ECFP-hDaxx or ECFP-PML, or EYFP-hDaxx with ECFP-PML. Selection during routine culture used G418 at 0.5 mg/ml and/or puromycin at 500 ng/ml as relevant. The antibiotic was omitted from cells seeded for and during experimentation.

**Antibodies**

The following antibodies were used: anti-actin mAb AC-40 (Sigma-Aldrich), anti-PML rabbit polyclonal (rAb) ABD030 (Jena Bioscience) or mAb 5E10 (32), anti-IFI16 mAb ab55328 (Abcam), anti-hDaxx rAb 07-471 (Upstate), anti-ICP4 mAb 58S (33) and anti-EGFP rAb ab290 (Abcam).

**Immunofluorescence**

Cells on 13 mm glass coverslips were fixed with formaldehyde and prepared for immunofluorescence using standard methods. The secondary antibodies used were FITC-conjugated sheep anti-mouse IgG (Sigma), Alexa 555 conjugated goat anti-mouse or anti-rabbit IgG and Alexa 633 conjugated goat anti-rabbit IgG (Invitrogen). The samples were examined using a Zeiss LSM 710 confocal microscope, with 488 nm, 561 nm and 633 nm laser lines, scanning each channel separately under image capture conditions that eliminated channel overlap. The images were exported as tif files, minimally adjusted using Photoshop, then assembled into the figures using Illustrator.

**Live cell microscopy**

Cells were seeded into Nunc Lab-Tec chambered coverglass live cell chambers at 1 x $10^5$ cells per well (4 chambered unit) then infected or not as relevant the following day. Immediately prior to infection the cells were washed with DMEM without phenol red, and after virus adsorption they were overlayed with the same medium containing 2% FCS and antibiotics as above. For high multiplicity of infection (MOI) experiments, a range of MOI between 25 and 100 was used. The nature of the results was not essentially influenced by choice of MOI, except that events occurred more commonly and were thus easier to detect as the MOI was increased. If plaques were later to be examined, the medium in the overlay also included 1% human serum. If the cells were to be examined immediately, the virus absorption period was 15 minutes, then the cells were placed into the microscope incubation chamber (pre-heated to 37 $^{o}$C) and imaging was commenced as soon as possible. In this latter case, times stated in the figures relate to time after addition of the virus. The microscope was a Zeiss Axio Observer Z1 equipped with definite focus control to maintain the correct focal plane during a time course. The S1 environmental system was used to maintain $CO_2$ at 5% and temperature and humidity. Filter sets 47 HE and 46 HE were used for detecting ECFP and EYFP respectively, using illumination from an HXP 120V unit. The samples were examined using either x40 NA 1.3 or x63 NA 1.4 oil immersion lenses. Image capture conditions generally utilised 70% or 50% HXP power with 2x gain and 4x4 binning, with exposure times of the order of 100 – 200 ms for both channels. The time intervals between

images are noted in each figure as relevant. Imaging conditions were adjusted so that the cells did not suffer detectably from light poisoning and so that any photobleaching was minimized during the course of the experiment. Generally this was not a problem for EYFP-IFI16 or ECFP-PML, but ECFP-hDaxx gave weaker fluorescence and was subject to photobleaching if image capture frequency or illumination intensity was too high. Cells were selected for imaging on the basis of signal intensities to ensure sufficient image quality. For IFI16 this was fairly uninform, but cells expressing sufficient ECFP-hDaxx were in a minority. Relevant segments of the image series were cropped and exported as avi files (for the Supplemental movies) or saved separately for excision of individual frames for presentation in the figures, after adjustment of minimum and maximum thresholds for each channel for ease of visualization. Due to space constraints, only a small number of examples of each phenomenon can be presented in the figures. In some cases, additional events of the same nature can be seen in the accompanying Supplemental Movies (which may cover a longer time frame). In all cases, the data presented are representative of several independent experiments.

For the photoactivation experiments, cells were examined in a live cell adapted Zeiss LSM 510 META microscope with incubation at 37$^{o}$C and 5% $CO_2$. Selected cells were first imaged for mCherry, then regions of interest were illuminated with the 405 nm laser at 50% power for 10 reiterations using the bleach programme in the LSM 510 software. Subsequent imaging was performed at timed intervals thereafter for both EGFP and mCherry using the 488 nm and 543 nm lasers respectively. For GAPC-hDaxx, images were acquired at 2 second intervals after bleaching, while 15 second intervals were used for GAPC-PML.I.

## RESULTS

### Description of the basic experimental system and underlying assumptions and extrapolations

Gerd Maul was first to observe that the genomes of many DNA viruses, particularly HSV-1 and HCMV, could be observed to be in association with PML NBs during the early stages of infection (34, 35). These studies used the difficult technique of fluorescence in situ hybridisation (FISH) to detect the viral genomes, which although providing direct evidence, is limited because antibody staining techniques to detect viral or cellular proteins are not always compatible with the required harshness of the hybridisation procedure. Later it was discovered that in cells at the edges of developing plaques, viral genomes could be detected frequently in large numbers of foci that formed characteristic arcs just inside the nuclear

envelope. These genomes could be detected by FISH, but also importantly by simple fluorescence detection of the viral transcriptional activator protein ICP4 (11). This occurs because ICP4 binds avidly to viral DNA. Thus in such cells an asymmetric pattern of ICP4 foci near the nuclear periphery could be taken to identify viral genome sites, without the need for FISH. Because this highly asymmetric staining pattern is completely distinct from the normal situation in uninfected cells, it can be safely deduced that PML NB protein foci in this arrangement are also associated with the mutant viral genomes, even if neither FISH nor ICP4 staining are included in the protocol. Several experiments in this report are dependent on this deduction. Analogous recruitment can also be observed in wt HSV-1 infections, but this is difficult to detect because the effects of ICP0 render it weak and transient (11). Having established these facts, it becomes possible to infer that small novel foci of relevant cellular proteins that can be detected only after a cell is infected are also highly likely to be associated with viral genomes. This is an important extrapolation in a number of the following experiments, but it is also supported by additional data where possible. These reasonable deductions and extrapolations are necessary because it is not possible to utilise FISH in live cells, and it is sometimes difficult to combine an experimental protocol with the use of a virus expressing fluorescently tagged ICP4. As examples of these phenomena, various phenotypes of PML and hDaxx recruited to ICP4 foci in cells at the edge of a developing ICP0 null mutant plaque are presented in Fig. 1, and for IFI16 in Fig. 2.

**The pyrin domain of IFI16 is required for its recruitment to HSV-1 genomes**

We and others have observed that IFI16 is also recruited efficiently to HSV-1 genomes at the early stages of infection (23-29). In order to study this in more detail, we used a lentiviral vector system for expressing EYFP-tagged IFI16 (Fig. 2A). Transduced human fibroblasts were selected with G418 then EYFP-positive cells were enriched by FACS. Analysis of the resultant cell line by western blotting showed that the cells expressed EYFP-linked IFI16 at levels of the same order as the three endogenous isoforms (Fig. 2B). We used the same system to express two mutant forms of IFI16, one with three point mutations in the pyrin domain (mutant m3) (30) and one a deletion of the second DNA binding HIN domain (ΔHIN2) (Fig. 2C). The EYFP-IFI16 proteins expressed in these cell lines were distributed similarly in the nucleus, with a general diffuse signal throughout the nucleoplasm and a concentration in the nucleoli (Fig. 2D). This is very similar to endogenous IFI16 in HFs (24, 26). As with the endogenous protein (24), wt EYFP-IFI16 was clearly recruited to sites associated with HSV-1 genomes (detected by ICP4) in cells at the edge of developing ICP0 null mutant plaques (Fig. 2E, upper row). This recruitment is not detectable in this way in wt

infected cells because of ICP0 activity (24). The recruitment of EYFP-IFI16 also occurred normally in the case of the ΔHIN2 mutant (Fig. 2E, bottom row), showing that one DNA binding HIN domain is sufficient for this activity, but the PYD mutant m3 was not efficiently recruited (Fig. 2E, middle row). The pyrin domain is required for oligomerization of IFI16 when bound to DNA (30), and therefore the recruitment of IFI16 probably represents assembly of IFI16 oligomers on the viral DNA at this stage of infection, soon after it is released into the nucleus in an unchromatinised form. It was not possible to test whether deletion of both HIN domains also inhibited recruitment as further deletion upstream of HIN2 caused loss of expression in this system.

**Depletion of PML does not compromise recruitment of IFI16 to HSV-1 genomes**

It was of interest to determine whether or not PML is involved in the recruitment of IFI16 to HSV-1 genomes. HFs depleted of PML using a lentiviral shRNA vector (7) were infected with ICP0 null mutant HSV-1 and the distributions of hDaxx and IFI16 in cells at the edges of developing plaques were investigated. Infected cells could be identified by the characteristic redistribution of hDaxx (see Fig. 1), and as expected IFI16 was similarly re-distributed in such cells (Fig. 2F, left). The same distributions of hDaxx and IFI16 were observed in infected PML depleted cells, while in uninfected cells hDaxx was dispersed throughout the nucleoplasm (Fig. 2F, right). These data demonstrate that while PML is required for hDaxx to be present in PML NB foci in uninfected cells (as reported previously; (17, 18)), PML depletion compromises recruitment of neither hDaxx (in agreement with previous data; (8)) nor IFI16. Thus recruitment of IFI16 to HSV-1 genomes is independent of the events that lead to the transient co-localization of IFI16 with PML during the early stages of wt HSV-1 infection (23, 24).

**Rapid response of IFI16 to HSV-1 genomes as they enter the nucleus**

Having developed a cell line expressing EYFP-linked wt IFI16, it was now possible to examine the behaviour of the protein in live infected cells. An unexpected observation was that high multiplicity ICP0 null mutant HSV-1 infection induced the formation of small, discrete novel IFI16 foci, often just inside the nuclear periphery. These were highly transient, in that they could appear and then disappear within 5 minutes, with others subsequently appearing similarly in a different part of the cell (Fig. 3A and Supplemental Movie 1). Such events also occurred during wt HSV-1 infection (Fig. 3B and Supplemental Movie 2). The appearance of these foci was completely dependent on infection, and generally occurred within the first hour or two after addition of the virus to the cell monolayer. Given their size and location, it seemed possible that they represent transient recruitment of IFI16 onto HSV-1

genomes at the time and point that they are released into the nucleus. Support for this explanation came from study of cells at the edge of a developing ICP0 null mutant plaque. Such cells will be infected at much higher multiplicity than can be achieved by adding a virus inoculum to the cell monolayer, and would thus be expected to display more of such foci. This was indeed the case, with many examples of such transient foci being plainly visible in Fig. 3C and Supplemental Movie 3.

Further support for this hypothesis came from infection of HFs expressing EYFP-IFI16 with dl0C4, an ICP0 null mutant HSV-1 expressing ECFP-linked ICP4 (4). The compilation in Fig. 3D and Supplemental Movie 4 shows a cell at the edge of a developing plaque from an image sequence that began before any IFI16 foci were detected. The displayed images illustrate the appearance of transient IFI16 foci at various locations within the first 42 minutes, mostly in the lower left quadrant of the cell but also elsewhere. As time continues, the foci in this region become more stable and prominent, and develop into an arc-like mass. The ECFP signal indicates that these apparently more stable IFI16 accumulations are very close to where ICP4 foci begin to accumulate, marking the positions of viral genomes. These data are consistent with the interpretation that this cell is being infected by genomes arriving mostly in the lower left quadrant of the nucleus which initially induce transient foci of IFI16. As infection develops, and ICP4 expression becomes detectable (and likely DNA replication begins, as judged from the expansion of the ICP4 compartments), the accumulations of IFI16 become more marked and longer lasting. From previous results, it can also be deduced that this second phase is inhibited by ICP0, but the first phase takes place too rapidly for it to be inhibited as it probably occurs before ICP0 is expressed in sufficient quantities.

**Rapid recruitment of hDaxx from the nucleoplasm to HSV-1 genomes**

Previous work using fluorescence recovery after photobleaching (FRAP) had demonstrated that hDaxx is highly mobile in the nucleus, with an exchange rate between PML NBs and the general nucleoplasm measured in terms of seconds, while that for PML was slower but still in the low numbers of minutes (11, 36-38). It had previously been suggested that this dynamics behaviour would enable the formation of novel PML NB-like foci in association with HSV-1 genomes without the need for movement of pre-existing PML NBs (11). To address this point further, lentiviral vectors were constructed that express either hDaxx or PML linked to a fusion protein that includes mCherry and a photoactivatable version of EGFP (31). Thus the expressed proteins are constitutively red fluorescent, but also are green fluorescent after photoactivation by pulses of a 405 nm laser. In this way, the movement of molecules from

activated to unactivated parts of the nucleus could be monitored. This technique was therefore adopted to examine the dynamics of recruitment of both hDaxx and PML from one region of the nucleus to another in which viral genome foci were present. Experiments in uninfected cells illustrated that photoactivation of both the PML and hDaxx fusion proteins occurred as expected, and subsequently the migration of activated molecules to the unactivated part of the nucleus could be observed. In the case of PML, this took place over a period of minutes, while with hDaxx the timescale was in seconds, to the extent that some transfer into the unactivated part of the nucleus was seen as soon as an image could be captured after photoactivation (Fig. 4A). These results are consistent with the previous FRAP experiments (11, 36-38). By applying this technique to cells at the edge of developing ICP0 null mutant HSV-1 plaques, characteristic recruited patterns of hDaxx and PML could be seen via the mCherry fluorescence. After photoactivation of molecules in the opposite part of the nucleus, the presence of activated hDaxx in the viral genome associated foci could be detected at the earliest possible time point, becoming more pronounced within 15 seconds (Fig. 4B, upper row). The same occurred with PML over a longer time course (Fig. 4B, lower row). Therefore recruitment of hDaxx in particular to viral genome associated foci is extremely rapid.

**IFI16 and hDaxx respond to HSV-1 genomes more rapidly than PML**

Given that both IFI16 and hDaxx can be recruited very rapidly to sites associated with HSV-1 genomes, it was of interest to determine whether a temporal order of IFI16 and PML NB components could be determined. Therefore cells were constructed that express EYFP-IFI16 and either ECFP linked hDaxx or PML. The EYFP-IFI16/ECFP-hDaxx cells were infected at high multiplicity with ICP0 null mutant HSV-1 then an image sequence was captured during the very early stages of infection (Fig. 5). As before, in a proportion of cells transient virus-induced small foci of IFI16 appeared, to which hDaxx was also recruited *de novo* either at the same time point or very shortly thereafter. Therefore these proteins respond with similar kinetics to HSV-1 infection. That hDaxx is also recruited to these novel, virus-induced IFI16 foci is further evidence that they are associated with viral genomes because it has been established that hDaxx responds in this manner. As time proceeds, the IFI16 signal is lost from these foci but they remain hDaxx positive.

A similar experiment was performed using EYFP-IFI16/ECFP-PML cells infected with ICP0 null mutant HSV-1. Fig. 6 shows a cell infected at high MOI in which an example of an event that commonly occurs in such cells was observed. First, a small accumulation of IFI16 appears near the nuclear periphery which at this point contains no detectable PML (Fig.

6, 3 min time point). As before IFI16 appears only transiently, but at the 7 min time point this dot also contains PML. At later time points, IFI16 is no longer detectable in this dot, while PML remains in this novel focus. Over the following minutes the new PML dot becomes closer to a pre-exiting PML NB and then appears to merge with it. The top three rows of Fig. 6 show the IFI16, PML and merged signals of the cell in question over an 11 minute period, while the lower three panels show enlargements of these images covering the event in question. Examination of similar times over a longer period indicated that these events were quite common, although the novel foci did not always merge with pre-existing PML NBs (Supplemental movie 6). A possible interpretation of the these observations will be presented in the Discussion, but for now it can be concluded that these events are most easily explained by a local and transient accumulation of IFI16 at viral genomes after their entry into the nucleus, which is followed by a more stable recruitment of PML.

**Recruitment of hDaxx is detectable before that of PML**

The results of Figs. 5 and 6 suggest that hDaxx responds more quickly than PML to virus infection, because there is a delay before the latter is detectable at the novel IFI16 foci, while the former appears commonly within the same time frame. To test this directly, the relative dynamics of recruitment of hDaxx and PML to HSV-1 induced foci were compared in HFs constructed to express ECFP-hDaxx and EYFP-PML. A common phenomenon observed in such cells near the periphery of developing ICP0-null mutant HSV-1 plaques was the appearance of novel foci that first included hDaxx, then at a slightly later time PML was also present. Many such examples are visible in the images of a time course presented in Fig. 7. A longer period of the time course of this cell is shown in Supplemental movie 7. A detail of one such example (boxed in the merged image of the first time point) is shown in the lowest row of Fig 7. At the first time point, a dot which is green only is present, which at 15 minutes is less intense (probably because it has moved slightly out of the focal plane) but by this time it also includes PML. At the 35 min time point another new green dot has appeared at the lower left of the detail (also clearly seen in the image of the whole cell nucleus; this also later accumulates PML) while one of the pre-existing PML NBs has moved out of this segment, leaving the dot that was green only at the start in the centre of detail. By the 45 min time point this appears to merge with its nearest neighboring PML NB. Detailed examination of the complete time course (see Supplemental movie 7) illustrates that these events are very common in such cells; novel foci including only hDaxx appear, which later include both hDaxx and PML, and which sometimes, but not always, merge with neighboring PML NBs.

Such events can only be detected by live cell analysis, which reveals these transient and subtle events that occur in cells close to developing plaques. Previous examination of such cells using fixed cell methods has concentrated on those in which marked viral genome associated foci in arcs near the edge of the nucleus have been formed, because in these cases it was clear that these novel foci were virus induced. The live cell analysis extends these findings to reveal virus induced events in many cells that do not or have not yet formed the characteristic peri-nuclear arcs of foci, and which could not be detected at such time points by co-staining for ICP4.

## DISCUSSION

The results presented in this paper allow an extension of previous models of the interactions between HSV-1 genomes and PML NB proteins during the earliest stages of infection (summarised in Fig. 8). At a very early stage of infection, probably within seconds and certainly within minutes of the release of the viral genome through the nuclear pore into the nucleoplasm, IFI16 can be seen to accumulate in distinct foci that are very likely to be closely associated with the viral genomes. This event is PML independent, but it requires the pyrin domain of IFI16 which is involved in its oligomerization on naked DNA (30). Equally rapidly, hDaxx also accumulates in these foci, co-localizing with IFI16 transiently while after a few minutes the IFI16 signal is dispersed. As the IFI16 signal weakens, PML then also accumulates in the foci, co-localizing with hDaxx. All these events can be observed in both wt and ICP0 null mutant infections, but in the latter the recruitment of hDaxx and PML is much more difficult to detect because of the activity of ICP0 (9, 11, 13). In the absence of ICP0, at high multiplicity and in cells which enter the lytic cycle, hDaxx and PML remain associated at sites close to the viral genomes as replication compartments develop, and a second and more stable phase of IFI16 recruitment occurs. At low multiplicity however, it is known that the majority of ICP0 null mutant HSV-1 genomes are repressed. The evidence here suggests that these genomes may remain stably associated with hDaxx and PML, perhaps sequestered within these modified PML NB-like structures, which in some cases appear to fuse with pre-existing PML NBs. This hypothesis is consistent with the observation that quiescent HSV-1 genomes can be detected within enlarged PML NBs in fibroblasts (39) (and in similar structures in latently infected mouse neurones; (40)). Unlike in the case of active replication in which IFI16 more stably accumulates close to the viral genomes (Fig. 2), IFI16 was not detected within PML NBs in quiescently infected cells (data not shown). Therefore recruitment of IFI16 appears to be a transient event.

In previous studies on the recruitment of PML NB proteins to incoming HSV-1 genomes, the analysis has concentrated on cells in which there are abundant numbers of ICP4 foci close to the edge of the nucleus. This was because the asymmetric pattern of PML NB protein foci could be clearly distinguished from their normal distribution in uninfected cells, and therefore clear unequivocal conclusions could be made. The observation of virus induced IFI16 foci in this study has allowed a more detailed analysis, because IFI16 never forms small punctate foci in uninfected cells. The use of dual labelled cells, combining EYFP-IFI16 with either ECFP-hDaxx or ECFP-PML, also increases the compelling nature of the observations. Because of these factors, the virus induced foci could be studied in cells in which they are neither abundant nor routinely close to the nuclear envelop, within cells at the early stages of a normal infection as well as of cells at the edges of developing plaques, and also within a background of pre-existing PML NBs. The live cell sequences thus clearly show the formation of novel virus-induced PML NB-like structures, that in some cases later fuse with pre-existing PML NBs, events that could underlie the detection of multiple quiescent viral genomes within enlarged PML NBs (39).

The mechanism of the recruitment of IFI16 to HSV-1 genomes involves several factors. One important aspect is likely to be that at the point of delivery into the nucleoplasm the viral DNA is not chromatinized, and this naked configuration is likely to underlie its recognition by IFI16 (30). ChIP assays performed by others have confirmed a direct association between IFI16 and HSV-1 DNA (25). IFI16 binds to DNA through its HIN domains, and it can form oligomers on DNA through pyrin domain interactions (30). A single HIN domain was sufficient IFI16 to accumulate on HSV-1 genomes, and the pyrin domain was essential for this abundant accumulation (Fig. 2). It is possible that the pyrin domain mutant protein still binds to viral DNA, but lack of oligomerization may reduce its accumulation on viral genomes to below a level detectable by microscopy. The recruitment of several PML NB proteins to HSV-1 genomes occurs through SUMO-related pathways, but whether sumoylation is also involved in the recruitment of IFI16 remains to be determined. IFI16 has been identified as a substrate for sumoylation in proteomic screens (16, 41), but an abundantly sumoylated form is not evident on western blots. Whatever the mechanisms involved, they are clearly inhibited by ICP0. As abundant recruitment of IFI16 does not occur in wt HSV-1 infected cells even when it remains easily detectable by fluorescence (24), this inhibition is not a simple consequence of the degradation of IFI16 that occurs at later times of wt HSV-1 infection (23, 24).

The transient nature of IFI16 accumulation in the virus induced foci is intriguing. It is not a simple case of the entity in question moving out of focus (although this can occur), because the hDaxx or PML signal remains. Clearly the recruitment of IFI16 is a very early event, and it is possible that as the viral genome is assembled into more complex nucleoprotein complexes, including nucleosomal structures, such that the parental viral DNA ceases to be in a naked form required for IFI16 accumulation. The more stable accumulation of IFI16 that occurs as replication compartments develop in ICP0 null mutant infections could reflect the production of newly replicated naked DNA in these structures. However, IFI16 does not co-localize with viral replication compartments in the same way as ICP8 and ICP4 even in ICP0 null mutant infections, but instead it often forms tangled filament-like structures that are associated with the replication compartments (24). These thread-like structures are likely to represent oligomers of IFI16, but how they relate to the viral DNA remains to be determined.

A fundamental question that is beyond the scope of this current study is whether or not IFI16 plays an important role in the recruitment of other proteins to the HSV-1 genome. This hypothesis would provide an interesting aspect to the role of IFI16 as a sensor of pathogen DNA. Previously it was shown that in cells depleted of IFI16, the recruitment of hDaxx appeared to be delayed, in that it was much more difficult to detect hDaxx in the asymmetric staining pattern characteristic of infected cells prior to detectable expression of ICP4 (in other words, cells of the phenotype of cell d in Fig. 1B were very rare in IFI16 depleted cells) (24). However, recruitment of both PML and hDaxx was readily detectable in the majority of IFI16 depleted cells once ICP4 was expressed (24). Although depletion of IFI16 was efficient in these cells, it is possible that trace remaining levels were sufficient for a hypothetical function in which recruitment of IFI16 is a necessary step prior to recruitment of the other proteins. Alternatively, the recruitment of IFI16 may be unlinked that that of PML NB proteins, and is more involved in regulation of chromatin assembly on the viral genome, as proposed by others (25, 26). Whatever the functions of the events described here, it is clear that detailed examination in live cells reveals striking and unexpected aspects of the dynamic nature of the interaction between cellular proteins and the invading HSV-1 genomes.

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**FIGURE LEGENDS**

**Figure 1**

Examples of recruitment of PML NB proteins to sites associated with HSV-1 genomes, as detected by staining for ICP4. The panels show views of cells close to developing ICP0 null mutant HSV-1 plaques in HFs, stained for ICP4 (green) and PML (red) (A) or ICP4 (green) and hDaxx (red) (B). Cells indicated by 'a' show the pattern of PML or hDaxx expected of uninfected cells. Cells labelled 'b' show various typical asymmetric patterns of ICP4 and PML NB protein staining showing recruitment of PML or hDaxx to sites associated with HSV-1 DNA. The cell labelled 'c' in (B) exhibits some faint foci of ICP4, each of which is associated with hDaxx staining. The cell labelled 'd' in (B) shows a less common phenotype, in which hDaxx foci are distributed in a highly asymmetric pattern, very likely associated with parental HSV-1 genomes, but before ICP4 expression has reached a detectable level.

**Figure 2**

The PYD domain is required for the recruitment of IFI16 to HSV-1 genomes. A. A map of a lentivirus vector that expresses EYFP-linked IFI16. B. Western blot analysis of cell extracts from a cell line transduced with the vector, analysed for IFI16 (left) and EGFP (right). There are three endogenous IFI16 isoforms, which differ in the number of 'S' regions in the hinge region between Hin-200A and Hin-200B. Only a single EYFP-linked isoform is expressed in these cells. C. A map of the coding sequence of IFI16, indicating the PYD and the two HIN domains, with two linker sequences (S1, S2). Also marked are the locations of the triple point mutations in mutant m3, and the region deleted in the mutant ΔHIN2. D. The nuclear distribution of wild type (wt) and mutant m3 and ΔHIN2 mutant forms of IFI16, detected by autofluorescence in transduced cell lines. E. The distributions of wt and mutant m3 and

ΔHIN2 mutant forms of IFI16 in cells at the edge of developing ICP0 null mutant plaques, indicating prominent recruitment of wt and ΔHIN2 IFI16, but not the m3 mutant, to sites that are closely associated with HSV-1 genomes (detected by staining for ICP4). F. Depletion of PML does not compromise efficient recruitment of IFI16 to HSV-1 genomes. The panels show either uninfected HFs or examples of cells at the edge of ICP0 null mutant HSV-1 plaques stained for hDaxx and IFI16, in control and PML depleted cells (left- and right-hand pairs of images, respectively).

**Figure 3**

Detection of the rapid response of IFI16 to HSV-1 infection. A. HFs expressing EYFP-IFI16 were infected with ICP0 null mutant HSV-1 (MOI 50) then images were captured at the indicated times after addition of the virus. B. A similar experiment was conducted using wt HSV-1 infection (MOI 20). C. Images from a sequence of a cell close to the edge of an ICP0 null mutant HSV-1 plaque. D. HFs expressing EYFP-IFI16 were infected with virus dl0C4 and images from a sequence of a cell at the edge of a developing plaque are presented, showing the IFI16, ICP4 and merged signals as indicated.

**Figure 4**

Dynamics of PML.I and hDaxx in uninfected cells and their rapid recruitment to sites associated with HSV-1 genomes. (A). Uninfected HepaRG cells transduced to express either GAPC-PML.I or GAPC-hDaxx were imaged before and after photoactivation of the PA-EGFP moiety within the red boxed area. Detection of PA-EGFP at intervals after photoactivation reveals migration of activated molecules from the activated region to the rest of the nucleoplasm. (B). As for A, but cells at the edge of developing ICP0 mutant HSV-1 plaques with characteristic asymmetrically distributed foci were examined under the same activation and time course conditions. Activated hDaxx can be detected in the asymmetric foci at the first time point after bleaching (about 2 seconds), becoming more prominent as time progresses. Activated PML accumulates in the foci at the nuclear periphery more slowly.

**Figure 5**

IFI16 and hDaxx respond to HSV-1 infection with similar kinetics. HFs expressing EYFP-IFI16 and ECFP-hDaxx were infected with ICP0 null mutant HSV-1 (MOI 100). After a 15 minute absorption period, the cells were examined by live cell microscopy, with images

captured every 90 sec. A selection of images is shown, with the times after adding the virus indicated in the top row. The upper three rows show the IFI16, hDaxx and merged channels, with the numbered arrows pointing to IFI16 foci that transiently appear during the sequence. The corresponding foci are also indicated on the hDaxx and merged panels. The sets of smaller images show details from each time point of the boxed areas marked in the 48 min sample. The left- and right-hand columns of these images for each time point show the images corresponding to the lower and upper boxes respectively, using the same numbering system. In this sequence covering just 9 minutes, four IFI16 foci appear simultaneously with an hDaxx signal, although in dot 3 the latter is weak at the 55.5 min time point. The presence of IFI16 is transient in dots 1 and 2 in this sequence, while hDaxx is more stable. A longer sequence from the same time course is shown in Supplemental movie 5, in which several other examples of the same phenomena can be seen.

**Figure 6**

IFI16 responds to HSV-1 infection more rapidly than PML. HFs expressing EYFP-IFI16 and ECFP-PML were infected with ICP0 null mutant HSV-1 (MOI 25). After a 20 minute absorption period, the cells were examined by live cell microscopy, with images captured every minute, starting 35 min after addition of the virus. A selection of images from the time course are presented, showing a cell in which a novel focus of IFI16 appears, which later becomes PML positive before merging with a pre-exiting PML NB structure. The lower set of three rows presents expansions for each time point of the boxed are in the merged image of the 82 min time point. Further details are provided in the text.

**Figure 7**

hDaxx responds more rapidly than PML to HSV-1 infection. HFs expressing both ECFP-hDaxx and EYFP-PML were infected with ICP0-null mutant HSV-1 and cells at the edge of a developing plaque were examined. A selection of images from an image sequence are presented, indicating the time (arbitrary) after the first image shown (which is image 54 of the original sequence, frame 24 as presented in Supplemental movie 7). The lowermost row shows a magnified view of the region that is boxed in the merged image of the leftmost column. Further details are provided in the text.

**Figure 8**

A model for the dynamics of IFI16 and PML NB components to HSV-1 genomes. A full explanation is provided in the text. The hexagon on the left represents a full capsid bound to the outer side of a nuclear pore through which the naked viral genome (tangled line) is transferred into the nucleoplasm.

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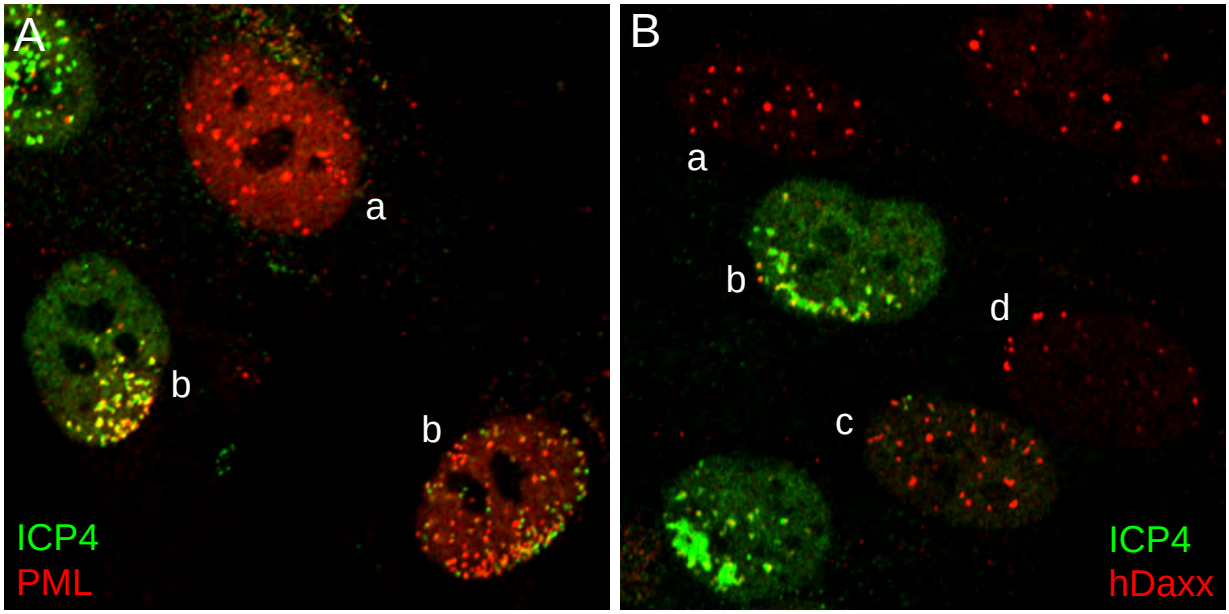
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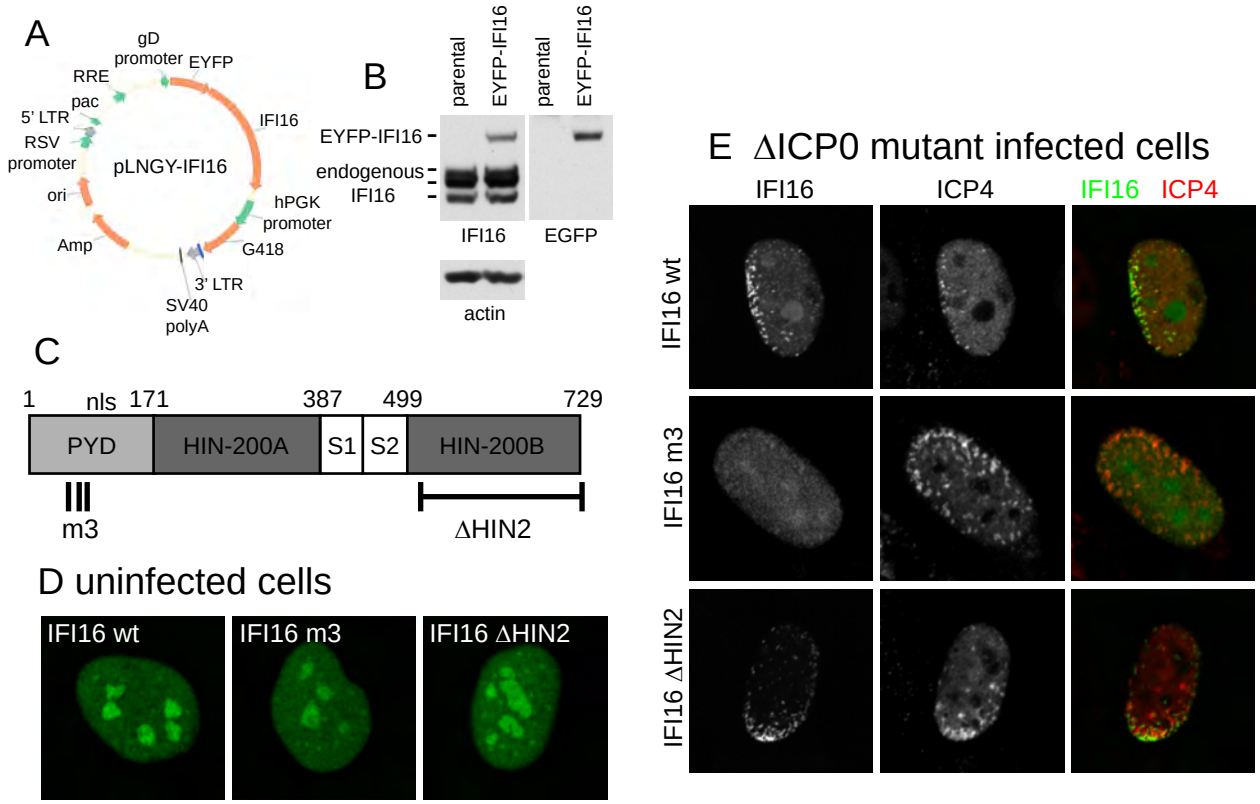
a
b
b
ICP4
PML
B
a
b
d
c
ICP4
hDaxx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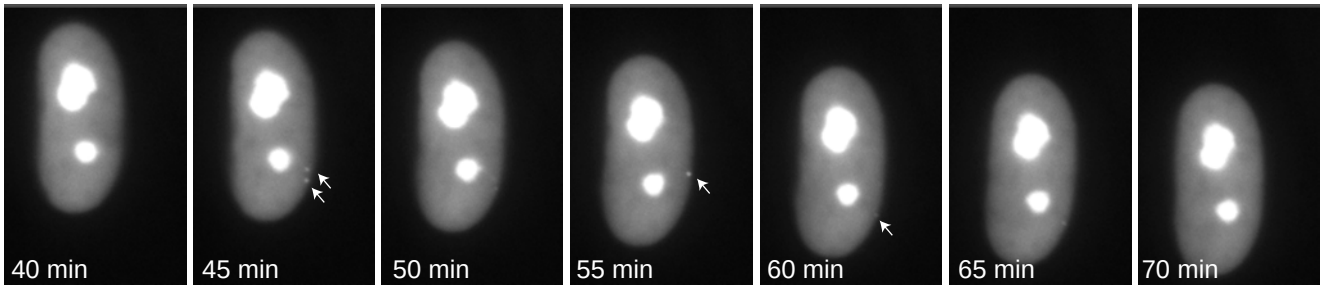
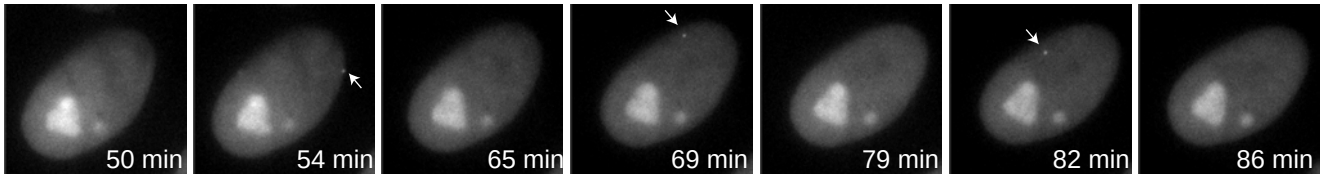
EYFP
RRE
pac
5' LTR
RSV promoter
IFI16
pLNGY-IFI16
ori
hPGK promoter
Amp
G418
3' LTR
SV40 polyA
B
parental
EYFP-IFI16
parental
EYFP-IFI16
EYFP-IFI16
endogenous IFI16
IFI16
EGFP
actin
C
1
nls
171
387
499
729
PYD
HIN-200A
S1
S2
HIN-200B
m3
ΔHIN2
D uninfected cells
IFI16 wt
IFI16 m3
IFI16 ΔHIN2
E ΔICP0 mutant infected cells
IFI16
ICP4
IFI16 ICP4
IFI16 wt
IFI16 m3
IFI16 ΔHIN2
F
HF control cells
uninfected
ΔICP0 HSV-1 infected
HF shPML cells
uninfected
ΔICP0 HSV-1 infected
hDaxx
IFI16
hDaxx
IFI16
hDaxx IFI16
hDaxx
IFI16

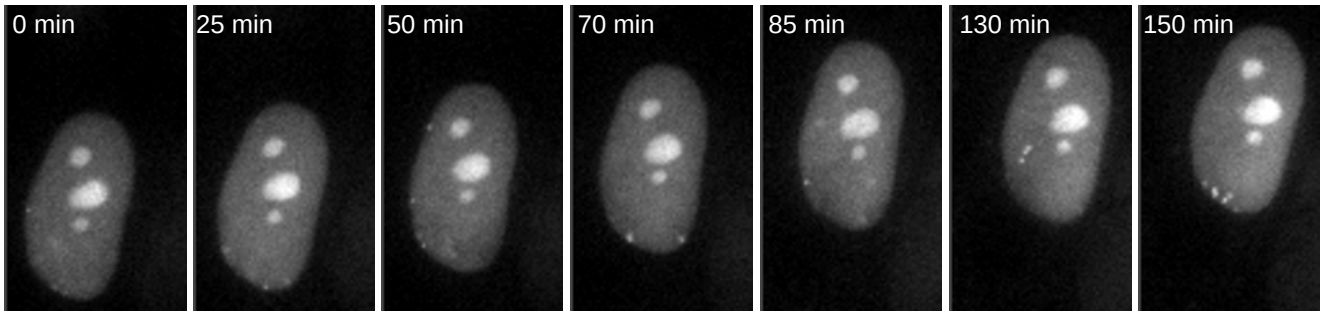
A: HFs, EYFP-IFI16, infected with ΔICP0 HSV-1, times after addition of virus



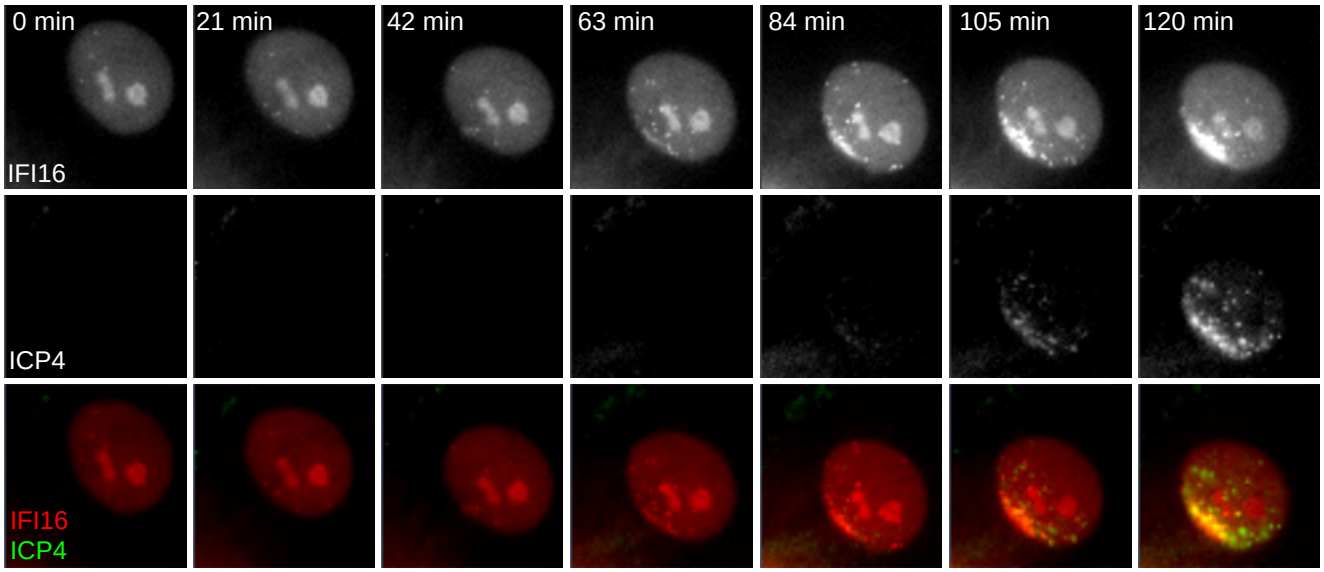

B: HFs, EYFP-IFI16, infected with wt HSV-1, times after addition of virus

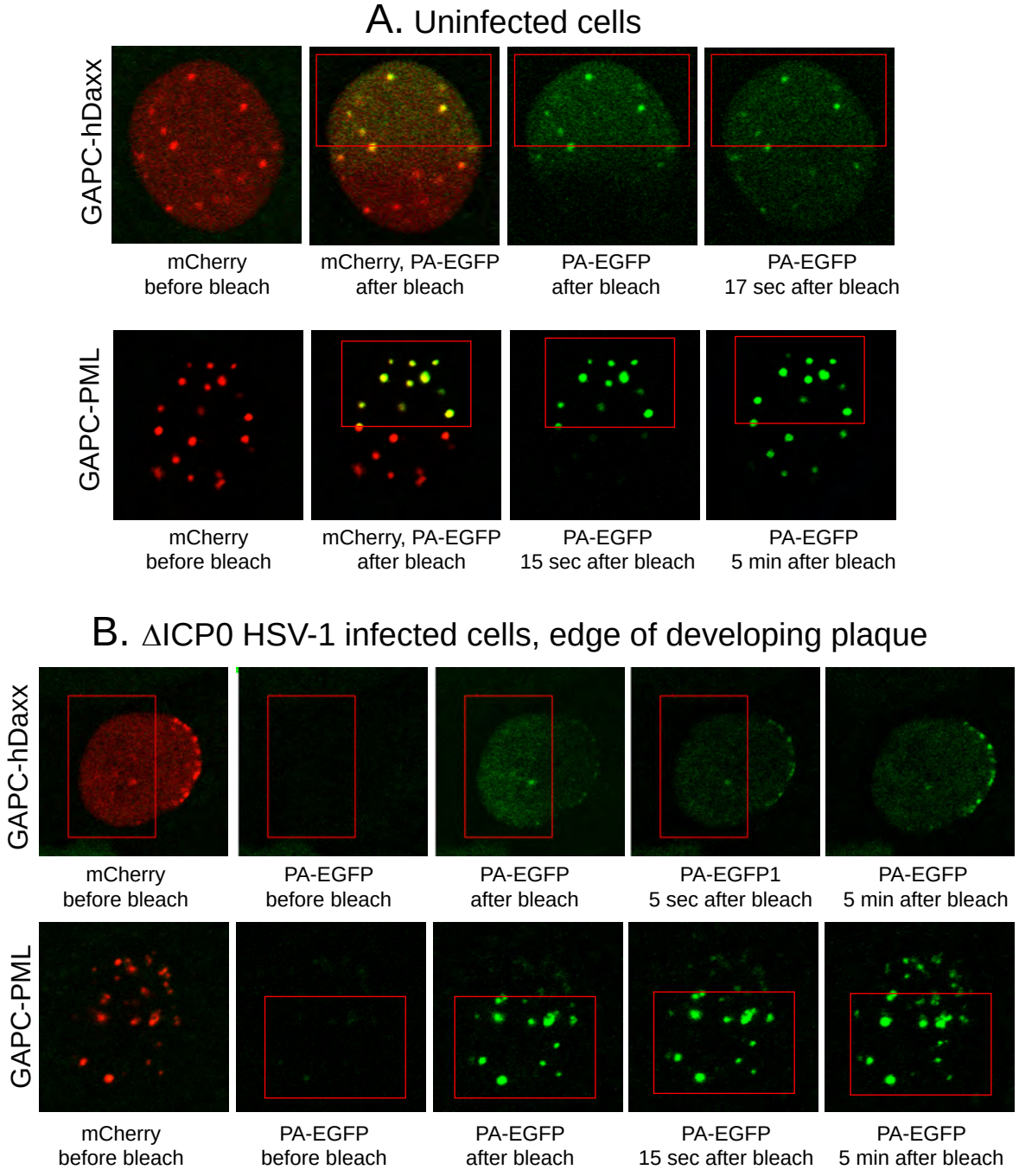



C: HFs, EYFP-IFI16, infected with ΔICP0 HSV-1, cell at edge of plaque, times after first image

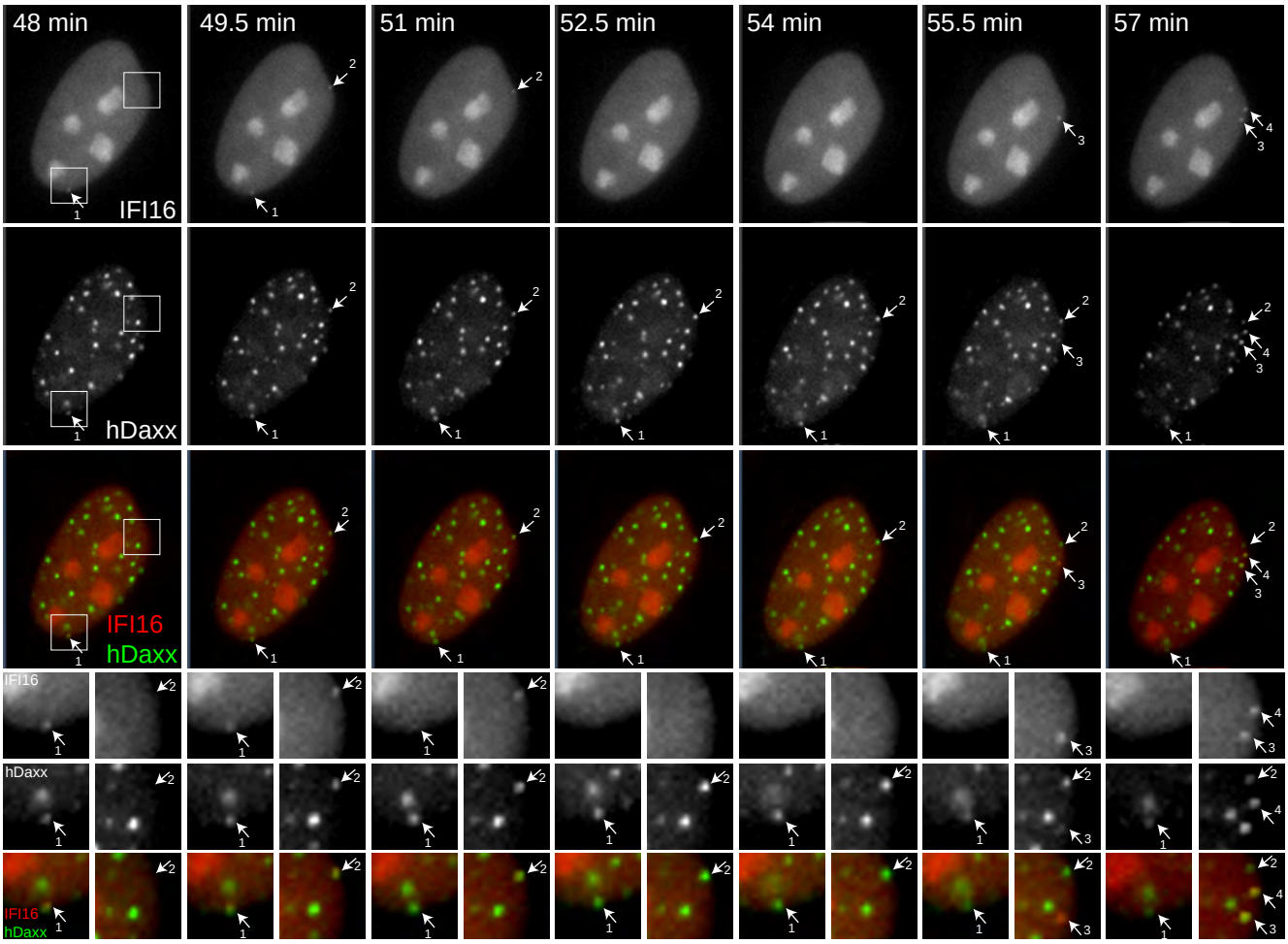



D: HFs, EYFP-IFI16, infected with dl0C4, cell at edge of plaque, times after first image

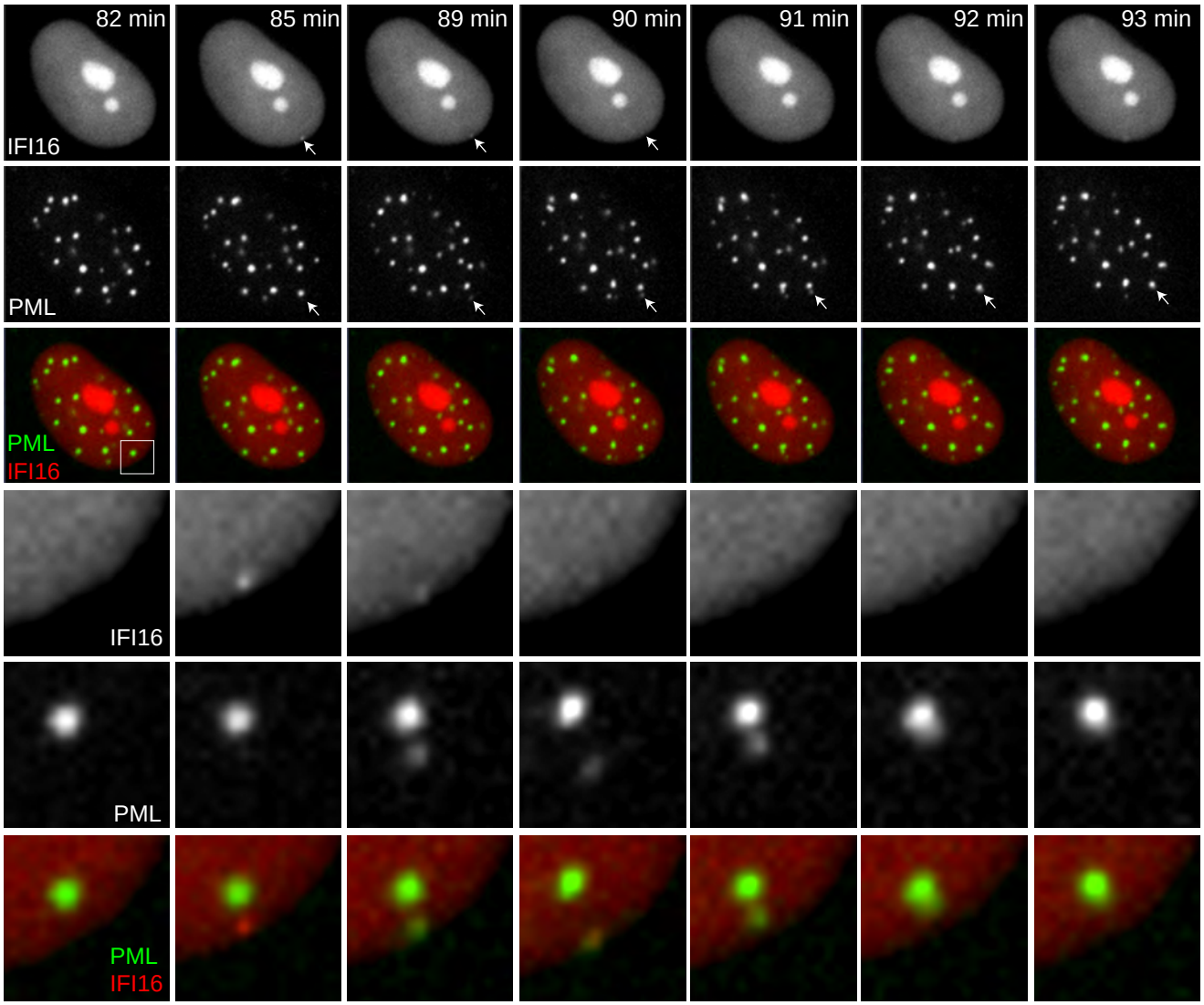



A. Uninfected cells
GAPC-hDaxx
mCherry
before bleach
mCherry, PA-EGFP
after bleach
PA-EGFP
after bleach
PA-EGFP
17 sec after bleach
GAPC-PML
mCherry
before bleach
mCherry, PA-EGFP
after bleach
PA-EGFP
15 sec after bleach
PA-EGFP
5 min after bleach
B. ΔICP0 HSV-1 infected cells, edge of developing plaque
GAPC-hDaxx
mCherry
before bleach
PA-EGFP
before bleach
PA-EGFP
after bleach
PA-EGFP1
5 sec after bleach
PA-EGFP
5 min after bleach
GAPC-PML
mCherry
before bleach
PA-EGFP
before bleach
PA-EGFP
after bleach
PA-EGFP
15 sec after bleach
PA-EGFP
5 min after bleach

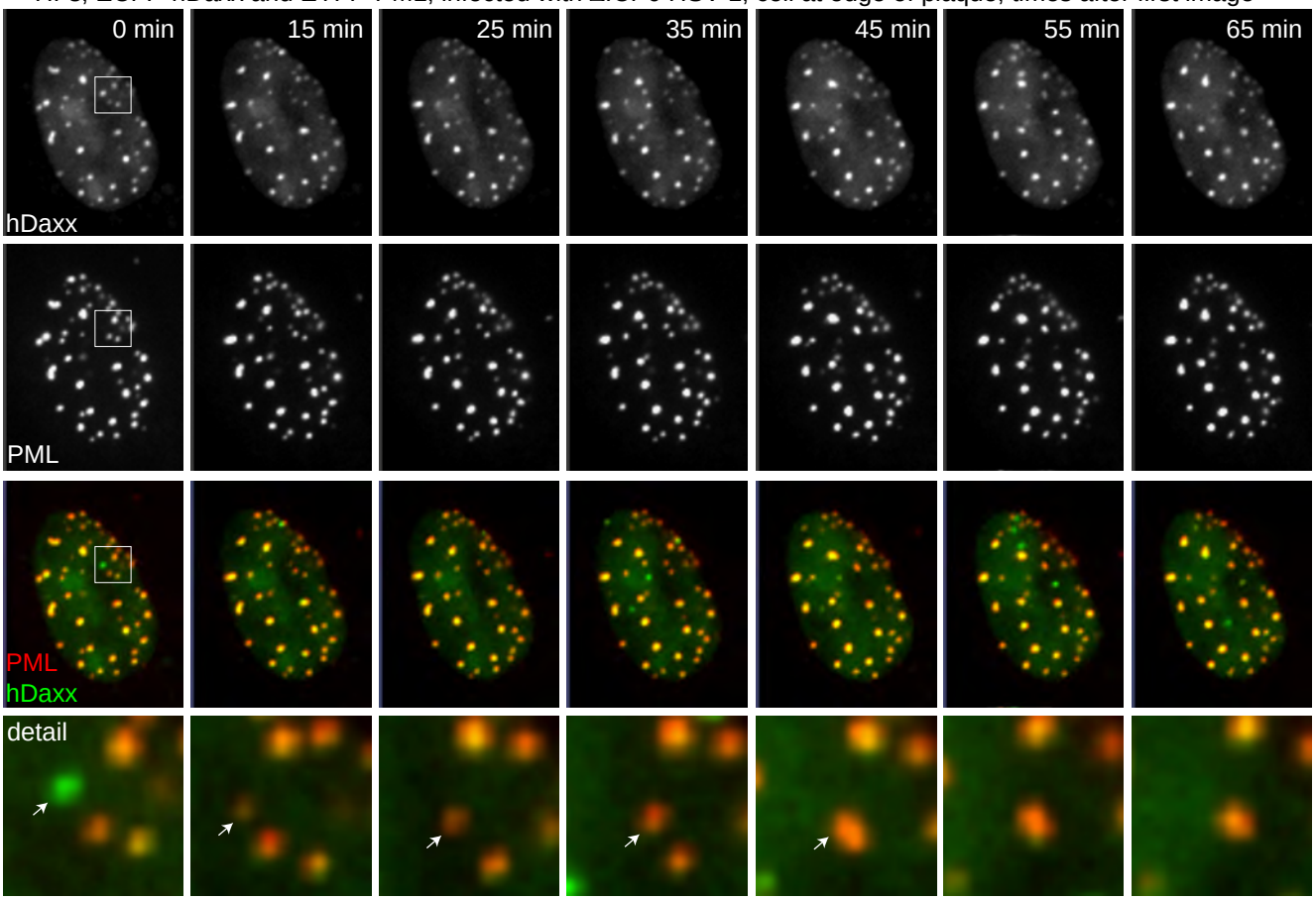
HFs, EYFP-IFI16 and ECFP-hDaxx, infected with ΔICP0 HSV-1 (MOI 100), times after addition of virus

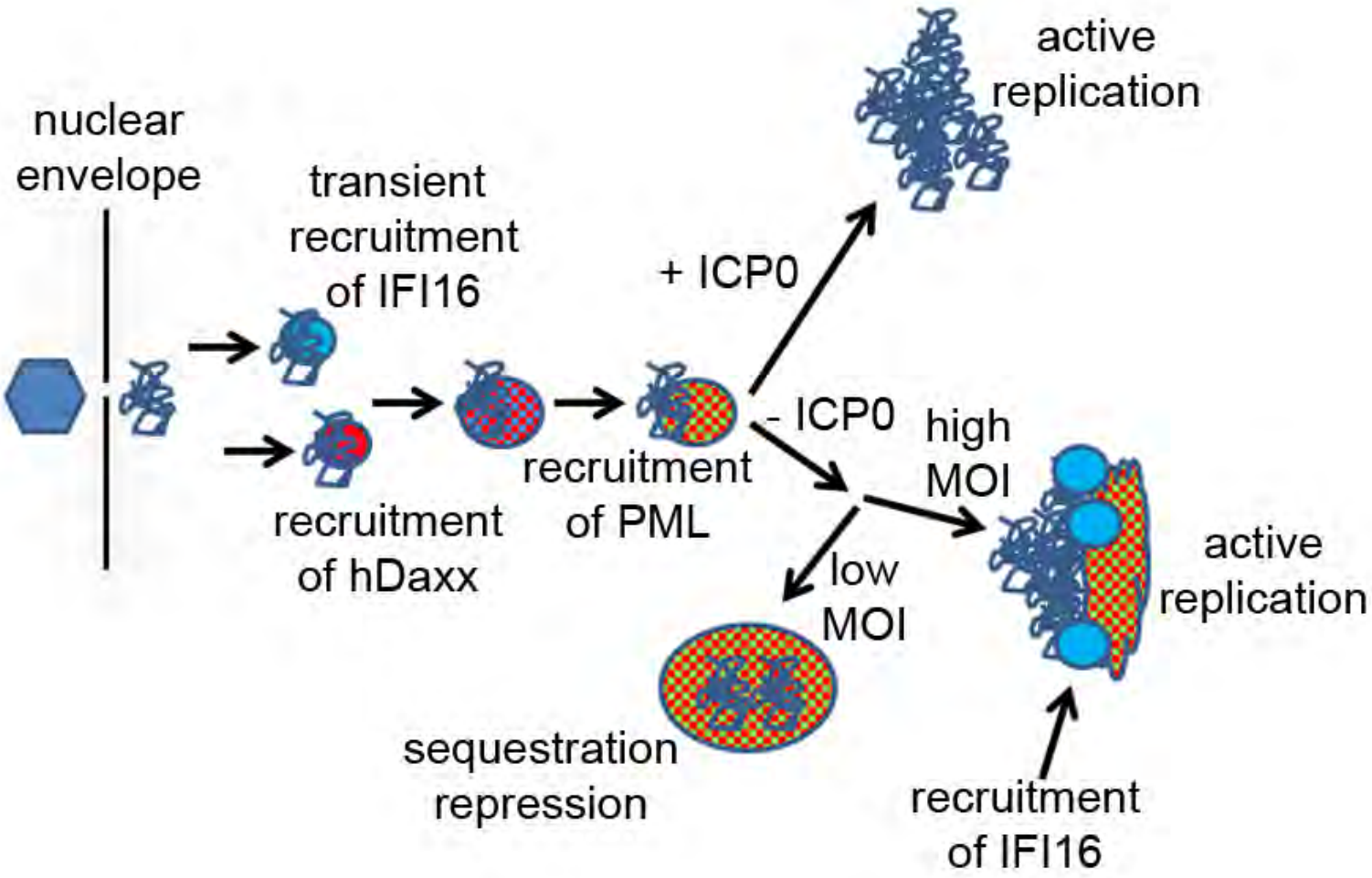


HFs, EYFP-IFI16 and ECFP-PML, infected with ΔICP0 HSV-1 (MOI 25), times after addition of virus



HFs, ECFP-hDaxx and EYFP-PML, infected with ΔICP0 HSV-1, cell at edge of plaque, times after first image




nuclear
envelope
transient
recruitment
of IFI16
recruitment
of hDaxx
recruitment
of PML
+ ICP0
active
replication
- ICP0
high
MOI
low
MOI
sequestration
repression
active
replication
recruitment
of IFI16