



Mansouri, D., McSorley, S. T., Park, J. H., Orange, C., Horgan, P. G., McMillan, D. C. and Edwards, J. (2021) The inflammatory microenvironment in screen-detected premalignant adenomatous polyps: early results from the integrated technologies for improved polyp surveillance (INCISE) project. *European Journal of Gastroenterology and Hepatology*, 33(7), pp. 983-989.

(doi: [10.1097/MEG.0000000000002202](https://doi.org/10.1097/MEG.0000000000002202))

This is the Author Accepted Manuscript.

There may be differences between this version and the published version. You are advised to consult the publisher's version if you wish to cite from it.

<http://eprints.gla.ac.uk/246927/>

Deposited on: 20 July 2021

## Original Article

### THE INFLAMMATORY MICROENVIRONMENT IN SCREEN-DETECTED PREMALIGNANT ADENOMATOUS POLYPS: EARLY RESULTS FROM THE INTEGRATED TECHNOLOGIES FOR IMPROVED POLYP SURVEILLANCE (INCISE) PROJECT

David Mansouri, Stephen T McSorley, James H Park, Clare Orange, Paul G Horgan, Donald C McMillan, Joanne Edwards, on behalf of the Integrated Technologies for Improved Polyp Surveillance (INCISE) collaborators

1. Academic Unit of Surgery, University of Glasgow, Glasgow Royal Infirmary, Glasgow, UK
2. NHS Greater Glasgow and Clyde Biorepository, Queen Elizabeth University Hospital, Glasgow, UK
3. Institute of Cancer Sciences, University of Glasgow, Wolfson Wohl Cancer Research Centre, Glasgow, UK

Corresponding Author

Mr David Mansouri, Consultant Colorectal Surgeon

Glasgow Royal Infirmary, Glasgow, UK, G31 2ER

Email: [David.mansouri@glasgow.ac.uk](mailto:David.mansouri@glasgow.ac.uk)

Tel: 0141 201 8675

Disclosures: none

Funding: The Cunningham Trust, SCO13499, Ref ACC/KWF/CT12/21

Word count: 2,649

Key words: INCISE, polyp, surveillance, inflammation, adenoma, colorectal

## **Abstract**

### Introduction

Around 40% of patients who attend for colonoscopy following a positive stool screening test have adenomatous polyps. Identifying which patients have a higher propensity for malignant transformation is currently poorly understood. The aim of the present study was to assess whether the type and intensity of inflammatory infiltrate differs between high-grade (HGD) and low-grade dysplastic (LGD) screen detected adenomas.

### Methods

A representative sample of 207 polyps from 134 individuals were included from a database of all patients with adenomas detected through the first round of the Scottish Bowel Screening Programme (SBoSP) in NHS GG&C (April 2009 to April 2011).

Inflammatory cell phenotype infiltrate was assessed by immunohistochemistry for CD3+, CD8+, CD45+ and CD68+ in a semi-quantitative manner at 20x resolution. Immune-cell infiltrate was graded as absent, weak, moderate or strong.

Patient and polyp characteristics and inflammatory infiltrate were then compared between HGD and LGD polyps.

### Results

CD3+ infiltrate was significantly higher in HGD polyps compared to LGD polyps (74% vs 69%,  $p < 0.05$ ). CD8+ infiltrate was significantly higher in HGD polyps compared to LGD

polyps (36% vs 13%,  $p < 0.001$ ) whereas CD45+ infiltrate was not significantly different (69% vs 64%,  $p = 0.401$ ). There was no significant difference in CD68+ infiltrate ( $p = 0.540$ ) or total inflammatory cell infiltrate (calculated from CD3+ and CD68+) ( $p = 0.226$ ).

## Conclusions

This study reports an increase in CD3+ and CD8+ infiltrate in HGD colonic adenomas when compared to LGD adenomas. It may therefore have a use in the prognostic stratification and treatment of dysplastic polyps.

## Introduction

Screening for colorectal cancer utilising either the guaiac-based faecal occult blood test (gFOBT) or the faecal immunochemical test (FIT) has been shown to reduce cancer specific mortality through the detection of early stage disease [1-3]. However, the majority of individuals who attend for colonoscopy following a positive screening test do not have cancer detected, but a large proportion do have adenomatous polyps. There is good evidence that colorectal cancer develops through the adenoma-carcinoma sequence and it has been estimated that approximately 25% of polyps greater than 1cm will develop into cancer over 20 years [4]. There is some evidence from gFOBT screening that in the context of a high positivity rate the incidence of cancer in a given population can be reduced by removal of these polyps [5].

There is now a wealth of evidence that progression and outcome of colorectal cancer is related to a complex interaction between tumour and host [6]. In particular, those with a more pronounced peri-tumoural inflammatory reaction have better cancer specific outcomes [7]. However, it is not clear whether such a relationship is relevant to malignant adenomatous polyps. Previous studies examining this phenomenon have been limited by both numbers and a focus on early invasive cancer and have failed to examine the host inflammatory response across the spectrum of dysplasia. Therefore, our understanding of the natural history of such polyps and in particular the role of the inflammatory infiltrate is limited [8-10].

Despite such insights, determining which patients with polyps have a higher propensity for malignant transformation remains a challenge. Current UK guidelines advise repeat or surveillance colonoscopies depending on the number, size and grade of polyp detected at index procedure [11]. Nevertheless, there is limited evidence that this impacts on patient outcome, especially in those of low to intermediate risk [12]. In addition, a recent population

study has suggested that colorectal cancer mortality may actually be higher in those patients who have a high risk polyp removed compared with the general population [13].

Therefore, the goal of the Integrated Technologies for Improved Polyp Surveillance (INCISE) project is to combine polyp and tissue characteristics, in particular genomic, transcriptomic, and local inflammatory features which are associated with a higher likelihood of future polyps. This will allow for post-polypectomy surveillance with greater precision, allowing fewer unnecessary procedures for those at low risk, and concentrating valuable resource on those at high risk.

As part of this overall project, the aim of the present study was to assess the role of the local inflammatory response in screen-detected dysplastic adenomas and to assess whether the type and intensity of inflammatory infiltrate differs between high-grade and low-grade dysplasia.

## Materials and Methods

### Patients

A database of all patients with adenomas detected through the first round of the Scottish Bowel Screening Programme (SBoSP) in NHS GG&C (April 2009 to April 2011) had previously been created. The screening algorithm and derivation of this cohort has been reported previously [14]. All colonoscopists have to comply with strict quality control measures and require to have Joint Advisory Group (JAG) accreditation. In addition, all polyps identified are removed at colonoscopy. Only sporadic adenomatous polyps found at screening were included, with serrated lesions, serrated adenomas, hyperplastic lesions, hamartomas, juvenile and inflammatory polyps excluded, along with polyps from any patients with a known polyposis/cancer syndrome or inflammatory bowel disease.

A representative sample of 207 polyps from 134 individuals was chosen for inclusion in the study. All samples were processed in a single pathology department (Glasgow Royal Infirmary). All polyps were greater than 10mm in size at histological analysis. Details on site and macroscopic morphological appearance of the polyp were obtained from endoscopic reports. Details on post-fixation size, grade of dysplasia and microscopic appearance were obtained from histopathology reports. All such histopathology reports were produced in accordance with Royal College of Pathologists (RCPATH) and Healthcare Improvement Scotland (HIS) Bowel Screening standards. Patient details included age, sex, and socioeconomic deprivation status. The Scottish Index of Multiple Deprivation (2009) was used as a measure of deprivation as has been described previously (SIMD 2009)[15]. Details on patient usage of Aspirin was obtained from pre-assessment documentation. Follow-up details on any further colonoscopies and details of recurrent or metachronous neoplasia

(colorectal cancer or dysplastic polyp), were obtained from patient medical records on a case-by-case basis.

Ethical approval for use of this tissue was obtained from the West of Scotland Research Ethics Committee (12/WS/0152).

### Immunohistochemistry

Assessment of inflammatory cell phenotype infiltrate was carried out by immunohistochemistry. Representative archival formalin fixed paraffin embedded tissue blocks were retrieved from archive and 2.5µm sections cut. Sections were then dewaxed rehydrated through graded alcohol. An autostainer (ThermoFisher, Autostainer 480s) was used to perform staining. Antigen retrieval was carried out in a PT module (ThermoFisher) using ThermoFisher dewax/retrieve solution pH9. Primary antibody was applied for 20 minutes at RT following antigen retrieval. Signal was amplified and visualised using the ThermoFisher Quanto kit and the diaminobenzidine (DAB) colour developer. Cell surface antigens were evaluated for T-lymphocytes (CD3+) dilution 1:300 (ThermoFisher), cytotoxic T-lymphocytes (CD8+) dilution 1:100 (ThermoFisher), helper T-lymphocytes (CD45+) dilution 1:500 (ThermoFisher) and macrophages (CD68+) dilution 1:5000 (ThermoFisher).

### Assessment of inflammatory infiltrate

All stained slides were converted to electronic format using a high resolution digital scanner (Hamamatsu NanoZoomer, Hamamatsu, Welwyn Garden City) and images viewed and assessed using Slidepath Digital Image Hub and Image Analysis module (Leica Microsystems, Wetzlar, Germany). The whole slide was then analysed in a semi-quantitative manner to assess intra-epithelial cell infiltrate at a resolution of 20x [16]. Immune-cell infiltrate was graded on a four-point scale as absent, weak, moderate or strong (Supplementary Figures 1 to 4). Following initial scoring, this was further dichotomised into low (absent and weak) and high (moderate and strong) for the purpose of analysis. A total of 30 slides for each stain were scored independently by two observers to confirm consistency of scoring (DM and JHP). The remainder of the slides were then scored by a single observer (DM). The inter-observer intraclass coefficients for each subtype were: CD3+ = 0.66, CD8+ = 0.66, CD45+ 0.69 and CD68+ = 0.79. A kappa value above 0.6 indicates good concordance.

A combined CD3+/CD8+ score was derived for each polyp to reflect the intra-epithelial densities of these two stains in manner similar to the recognised 'Immunoscore' for established tumours [17]. This was defined as such: high combined CD3+/CD8+ score polyps were CD3+ high and CD8+ high, low combined CD3+/CD8+ score polyps were CD3+ low and CD8+ low, medium combined CD3+/CD8+ score were all other polyps.

Furthermore, the total inflammatory infiltrate was then derived for each polyp based on a combination of lymphocyte (CD3+) and macrophage (CD68+) scores. This was defined as such: high total inflammatory infiltrate polyps were CD3+ high and CD68+ high, low total inflammatory infiltrate polyps were CD3+ low and CD68+ low, medium total inflammatory infiltrate were all other polyps.

## Statistical Methods

Associations between categorical variables were examined using the  $\chi^2$  test. For ordered variables with multiple categories the  $\chi^2$  test for linear trend was used. Wilcoxon signed-rank test was used for analysis of paired variables. Binary logistic regression analysis was used to assess risk of neoplasia recurrence. A value of  $p < 0.05$  was considered statistically significant. Statistical analysis was performed using SPSS software (SPSS Inc., Chicago, IL, USA)

## Results

### Per polyp analysis

A total of 207 polyps from 134 patients were included. 107 were high grade (HGD) and 100 were low grade (LGD). The median age of patients was 65 years and 33 (25%) were female. The majority of polyps were left sided, pedunculated and were between 10mm and 20mm in size. Only 23 (11%) of patients reported aspirin use (Table 1). Comparing HGD and LGD polyps there were more older, female and less deprived patients in the HGD group (both  $p < 0.05$ ). HGD polyps were more likely to be larger and have a villous component (both  $p < 0.05$ ) (Table 1). Examining the inflammatory infiltrate in polyps, high levels of CD3+, CD8+, CD45+ and CD68+ were observed in 67%, 25%, 67% and 72% of cases respectively. CD3+ infiltrate was significantly higher in HGD polyps compared to LGD polyps (74% vs 69%,  $p < 0.05$ ). CD8+ infiltrate was significantly higher in HGD polyps compared to LGD polyps (36% vs 13%,  $p < 0.001$ ) whereas CD45+ infiltrate was not significantly different (69% vs 64%,  $p = 0.401$ ). The combined CD3+/CD8+ score was significantly higher in HGD polyps than LGD polyps (35% vs 11%,  $p < 0.001$ ). There was no significant difference in CD68+ infiltrate (74% vs 70%,  $p = 0.540$ ) (Table 2) or total inflammatory cell infiltrate (calculated from CD3+ and CD68+) ( $p = 0.226$ ).

Both patient and polyp related factors were then examined to identify features associated with altered inflammatory infiltrates. There was no difference in the degree of CD3+, CD8+, CD45+ or CD68+ inflammatory infiltrate with regards to patient factors such as age, sex or deprivation. Aspirin exposure was associated with a significantly higher level of CD45+ infiltrate ( $p = 0.007$ ). With regards to polyp factors, larger polyps were associated with a significantly higher level of CD8+ infiltrate ( $p = 0.004$ ) and polyps with a villous component

had significantly higher levels of CD8+ infiltrate ( $p=0.021$ ) Larger polyps ( $p=0.003$ ) and polyps with a villous component ( $p=0.043$ ) had higher combined CD3+/CD8+ scores. Total inflammatory infiltrate as calculated from CD3+ and CD68+ was not related to patient or polyp factors (Table 3).

### Per patient analysis

In order to examine whether alterations in the microenvironment were related to altered host response, a per patient paired analysis was then carried out of those patients with multiple polyps ( $n=46$  patients) (Figure 1). This included those with multiple low-grade dysplastic polyps ( $n=24$  patients) and those with multiple mixed low and high-grade dysplastic polyps ( $n=20$  patients). Due to low numbers, analysis of the 2 patients with multiple high-grade polyps was not carried out. On paired testing using Wilcoxon signed-rank analysis there was an increase in CD3+ ( $p=0.059$ ), CD68+ ( $p=0.046$ ) and total inflammatory infiltrate ( $p=0.021$ ) in high-grade polyps of those who had both low and high-grade dysplasia. There was no change in CD8+ ( $p=0.705$ ), CD45+ ( $p=0.605$ ), or combined CD3+/CD8+ score ( $p=0.218$ ). No significant changes in inflammatory infiltrate were seen between polyps in those patients with low grade dysplasia only (CD3+,  $p=0.317$ ; CD8+,  $p=0.083$ ; CD45+,  $p=0.206$ ; CD68+,  $p=0.705$ ; combined CD3+/CD8+ score,  $p=0.109$ ; total inflammatory infiltrate,  $p=0.617$ ) (Figures 2a & b).

## Risk of future / metachronous neoplasia

In those patients with a single polyp who had been included in the analysis (n=88 patients) outcome was assessed with regard to risk of metachronous neoplasia. Of the 88 patients, 39 (44%) patients were excluded as they had multiple polyps at index colonoscopy that had not been assessed in this analysis. On follow up, with a minimum of 4 years, 34 patients had undergone at least 1 colonoscopy whereby 14 had evidence of further neoplasia (Figure 1). There was no significant association between any of the measured local inflammatory infiltrate parameters and metachronous polyp (Table 4).

## Discussion

The present study shows that, within the context of a colorectal cancer screening programme, there was an increase in inflammatory cell infiltrate in high-grade dysplastic polyps when compared to those adenomas with low-grade dysplasia. This was evident whether analysed within or between patients. Therefore, it appears that there is a specific host response to dysplastic changes in colorectal adenomas. Given the prognostic value of the tumour inflammatory cell infiltrate in established cancer [7], it may be that the inflammatory cell infiltrate will inform the likely outcome in patients with dysplastic polyps.

The results of the present study are consistent with the observation that there is a specific interaction between adenomatous cells and the microenvironment [8-10]. A full understanding of the local inflammatory microenvironment of colorectal adenomas is essential if we are to learn about why some adenomas progress. The immunoediting hypothesis suggests that for neoplasia to develop there is an immune profile shift from immunosurveillance to immunosuppression [18]. Based on this theory, adenomatous polyps represent a neoplastic lesion in the equilibrium phase and hence identifying which polyps are appear more likely to escape would be of considerable benefit [19]. Indeed, the findings of the present study are in keeping with this model, and an increasing lymphocytic infiltrate as a should be considered further as an advanced feature, and possibly as a marker of potential to escape. If such results were to be confirmed, then modulation of this local host inflammatory microenvironment to prevent escape could become of interest.

It is interesting to consider the finding of the present study with regards to patient outcome. Currently, follow-up of patients with adenomatous polyps is based on size, grade and number of polyps as previous studies have shown that patients with larger, high-grade and multiple polyps are at a higher risk of recurrence and of malignant transformation [11]. This

surveillance paradigm has been in use in the UK for around two decades [20-21]. However, this current risk stratification technique is far from ideal and a recent population based study has suggested that there is little benefit in terms of cancer incidence reduction in following these guidelines [13]. With just under half of all those attending for a colonoscopy following a positive bowel screening test having adenomatous polyps detected, accurate prognostic stratification is vital if we are to avoid unnecessary follow-up colonoscopy in a large proportion of the population [22]. If such a link were to be proven, then it would represent a potential immunomodulatory target to help prevent the progression of adenomas in a pre-malignant phase.

The present study was predominantly cross-sectional in nature and lacked numbers to study outcome with sufficient power. However, a pilot group of 34 patients who had solitary polyps excised were followed up and a high rate of recurrent or metachronous polyps were noted (over 40% within 4 years). This should inform planning for future studies in larger numbers to explore this further and could also include those with multiple polyps. However, care should be taken with such analysis as it would be prone to potential confounding factors such as the heterogeneity of inflammatory infiltrate between polyps within the same person as has been demonstrated within the present study. Therefore, it may be useful to examine other polyp characteristics including gene expression and transcription patterns along with inflammatory infiltrate to identify additional factors associated with advanced adenomas and indeed metachronous polyp formation. Indeed, there is some existing evidence that such omics techniques can be applied to excised polyps in such a manner [23-25].

The strengths of the present study are that it is, to date, the largest study examining the local inflammatory response in colorectal adenomas. It has examined a variety of inflammatory cells using robust immunochemical techniques. In including a per patient analysis, changes in inflammatory infiltrate between different lesions within the same colon is achieved and adds

to the robustness of the findings. Consensus to the grading of dysplasia can be variable, however all polyps were reported in line with RCPATH and national bowel screening standards in a single department, with centralised national slide review used when required. A potential additional weakness is the use of CD68+ as a marker for macrophage infiltration. When considering the immune microenvironment, macrophage subtypes associated with either an increase in the adaptive or the innate response can be of interest and CD68+ staining does not account for that. Further studies examining macrophage subtypes to explore this concept are planned in this cohort. One further potential weakness may be our use of a derived total inflammatory infiltrate rather than a true observed one. For example, in colorectal cancer the Klintrup-Makinen score, assessing inflammatory cell infiltrate at the invasive margin on routinely stained H&E slides, has been widely validated as means of observing a total inflammatory cell infiltrate and has been correlated with patient outcome [26]. However, given the nature of endoscopic polyp resections, where there is no clear margin in a large proportion of cases, means that inflammatory cell infiltrate can only be examined within the adenoma itself and not at the margin. Therefore, the Klintrup-Makinen score cannot accurately be assessed, however recent findings in colorectal cancer suggest that the lymphocyte subtype and location of the infiltrate is less prognostically important than the density [27-28]. The lack of margin assessment also limited the use of the well validated Immunoscore as described by Galon et al. [17]. However, the use of a combined intra-epithelial CD3+/CD8+ score has been proposed here as a method of compensating for this and further work in larger numbers is planned utilising this adaptation. Moreover, studies have shown that it is in fact density, rather than location that is the most important aspect of inflammatory infiltrate related to outcome when examined in established disease [27].

In conclusion, the present study has shown an increase in inflammatory infiltrate when adenomas with high-grade dysplasia were compared to those with low-grade dysplasia. This

might suggest a specific response to early disease progression, confirming increased host immunosurveillance. Such a finding may have a use in the prognostic stratification and treatment of dysplastic polyps, and as a mechanism of predicting the likelihood of future polyps given the known associations between advanced features and metachronous adenomas.

## **Acknowledgements**

This work was funded by The Cunningham Trust, St Andrews, Scotland (Scottish Charity number SCO13499, Ref ACC/KWF/CT12/21, awarded 23/11/2012).

The authors report no conflict of interest.

This manuscript was uploaded to the medRxiv server as a pre-print, prior to peer-review, and can be found at doi: <https://doi.org/10.1101/2020.08.16.20175935>

## References

1. Mandel JS, Bond JH, Church TR, Snover DC, Bradley GM, Schuman LM, Ederer F. "Reducing mortality from colorectal cancer by screening for fecal occult blood. Minnesota Colon Cancer Control Study." *N Engl J Med* 1993;328(19): 1365-1371.
2. Hardcastle JD, Chamberlain JO, Robinson MH, Moss SM, Amar SS, Balfour TW, James PD, Mangham CM. "Randomised controlled trial of faecal-occult-blood screening for colorectal cancer." *Lancet* 1996;348(9040): 1472-1477.
3. Kronborg O, Fenger C, Olsen J, Jorgensen OD, Sondergaard O. "Randomised study of screening for colorectal cancer with faecal-occult-blood test." *Lancet* 1996;348(9040): 1467-1471.
4. Stryker SJ, Wolff BG, Culp CE, Libbe SD, Ilstrup DM, MacCarty RL. "Natural history of untreated colonic polyps." *Gastroenterology* 1987;93(5): 1009-1013.
5. Mandel JS, Church TR, Bond JH, Ederer F, Geisser MS, Mongin SJ, Snover DC, Schuman LM. "The effect of fecal occult-blood screening on the incidence of colorectal cancer." *N Engl J Med* 2000;343(22): 1603-1607.
6. Hanahan D, Weinberg RA. "Hallmarks of cancer: the next generation." *Cell* 2011;144(5): 646-674.
7. Roxburgh CS, McMillan DC. "The role of the in situ local inflammatory response in predicting recurrence and survival in patients with primary operable colorectal cancer." *Cancer Treat Rev* 2012;38(5): 451-466.
8. Cui G, Yuan A, Vonen B, Florholmen J. "Progressive cellular response in the lamina propria of the colorectal adenoma-carcinoma sequence." *Histopathology* 2009;54(5): 550-560.

9. Cui G, Shi Y, Cui J, Tang F, Florholmen J. "Immune microenvironmental shift along human colorectal adenoma-carcinoma sequence: is it relevant to tumor development, biomarkers and biotherapeutic targets?" *Scand J Gastroenterol* 2012;47(4): 367-377.
10. McLean MH, Murray GI, Stewart KN, Norrie G, Mayer C, Hold GL, Thomson J, Fyfe N, Hope M, Mowat NA, Drew JE, El-Omar EM. "The inflammatory microenvironment in colorectal neoplasia." *PLoS One* 2011;6(1): e15366.
11. Rutter MD, East J, Rees CJ, et al. British Society of Gastroenterology/Association of Coloproctology of Great Britain and Ireland/Public Health England post-polypectomy and post-colorectal cancer resection surveillance guidelines. *Gut* 2020;69:201–223
12. Cross AJ, Robbins EC, Pack K, Stenson I, Kirby PL, Patel B, Rutter MD, Veitch AM, Saunders PB, Duffy SW, Wooldrage K. Long-term colorectal cancer incidence after adenoma removal and the effects of surveillance on incidence: a multicentre, retrospective, cohort study. *Gut* 2020;Published Online First: 17 January 2020. doi: 10.1136/gutjnl-2019-320036
13. Loberg M, Kalager M, Holme O, Hoff G, Adami HO, Bretthauer M (2014). "Long-term colorectal-cancer mortality after adenoma removal." *N Engl J Med* 371(9): 799-807.
14. Mansouri D, McMillan DC, Grant Y, Crighton EM, Horgan PG. The impact of age, sex and socioeconomic deprivation on outcomes in a colorectal cancer screening programme. *PLoS One* 2013;8(6):e66063
15. Scottish Index of Multiple Deprivation 2009. Available at <http://www.scotland.gov.uk/Topics/Statistics/SIMD>.

16. Gujam FJA, McMillan DC, Edwards J. The relationship between total and phosphorylated STAT1 and STAT3 tumour cell expression, components of tumour microenvironment and survival in patients with invasive ductal breast cancer. *Oncotarget* 2016;7:77607-77621
17. Galon J, Mlecnik B, Bindea G, Angell HK et al. "Towards the introduction of the 'Immunoscore' in the classification of malignant tumours." *J Pathol* 2014;232(2):199-209
18. Koebel CM, Vermi W, Swann JB, Zerafa N, Rodig SJ, Old LJ, Smyth MJ, Schreiber RD. "Adaptive immunity maintains occult cancer in an equilibrium state." *Nature* 2007;450(7171): 903-907.
19. Dunn GP, Old LJ, Schreiber RD. "The immunobiology of cancer immunosurveillance and immunoediting." *Immunity* 2004;21(2): 137-148.
20. Cairns S, Scholefield JH. Guidelines for colorectal cancer screening in high risk groups. *Gut* 2002;(SupplV):v1e2
21. Cairns SR, Scholefield JH, Steele RJ, Dunlop MG et al. Guidelines for colorectal cancer screening and surveillance in moderate and high risk groups (update from 2002). *Gut* 2010;59:666-690
22. Logan RF, Patnick J, Nickerson C, Coleman L, Rutter MD, von Wagner C. "Outcomes of the Bowel Cancer Screening Programme (BCSP) in England after the first 1 million tests." *Gut* 2012;61(10): 1439-1446.
23. Berger AW et al. Genetic Biopsy for Prediction of Surveillance Intervals after Endoscopic Resection of Colonic Polyps: Results of the GENESIS Study. *United European Gastroenterol J.* 2018 Mar;6(2):290-299.

24. Druliner BR et al. Molecular characterization of colorectal adenomas with and without malignancy reveals distinguishing genome, transcriptome and methylome alterations. *Sci Rep*. 2018 Feb 16;8(1):3161
25. van Lanschot MCJ, et al. Molecular profiling of longitudinally observed small colorectal polyps: A cohort study. *EBioMedicine*. 2019 Jan;39:292-300.
26. Klintrup K, Makinen JM, Kauppila S, Vare PO, Melkko J, Tuominen H, Tuppurainen K, Makela J, Karttunen TJ, Makinen MJ. "Inflammation and prognosis in colorectal cancer." *Eur J Cancer* 2005;41(17): 2645-2654.
27. Alexander PG, McMillan DC, Park JH. The local inflammatory response in colorectal cancer - Type, location or density? A systematic review and meta-analysis. *Cancer Treat Rev*. 2020 Feb;83:101949.
28. Haruki K, Kosumi K, Li P, Arima K, Väyrynen JP, Lau MC, Twombly TS, Hamada T, Glickman JN, Fujiyoshi K, Chen Y, Du C, Guo C, Väyrynen SA, Dias Costa A, Song M, Chan AT, Meyerhardt JA, Nishihara R, Fuchs CS, Liu L, Zhang X, Wu K, Giannakis M, Nowak JA, Ogino S. An integrated analysis of lymphocytic reaction, tumour molecular characteristics and patient survival in colorectal cancer. *Br J Cancer*. 2020 doi: 10.1038/s41416-020-0780-3 [epub ahead of print]

## **Tables and footnotes**

Table 1: Baseline characteristics of adenomatous polyps by grade of dysplasia (per polyp analysis)

Table 2: Comparison of inflammatory infiltrate by grade of adenomatous polyp (per polyp analysis)

Table 3: Associations between adenomatous polyp and host variables and degree of inflammatory infiltrate (per polyp analysis)

Table 4: Associations between inflammatory infiltrate and future / metachronous neoplasia in those with solitary polyps

## Figures and legends

Figure 1: Outline of patient cohort and explanation of per patient analysis in those with multiple polyps, and follow up in those with solitary polyps (shaded box), (HGD = High-grade dysplasia, LGD = Low-grade dysplasia)

Figure 2a: Per patient paired analysis of CD3+, CD8+, CD45+ T-lymphocyte cell infiltrate changes between polyps in patients with multiple polyps (mixed high/low grade dysplasia or low grade dysplasia only)

Figure 2b: Per patient paired analysis of CD68+ macrophage, combined CD3+/CD8+ score and total inflammatory cell infiltrate changes between polyps in patients with multiple polyps (mixed high/low grade dysplasia or low grade dysplasia only)

## **Supplementary Data**

Supplementary Figure 1: Assessment of CD3+ T-lymphocyte inflammatory cell infiltrate

Supplementary Figure 2: Assessment of CD8+ cytotoxic T-lymphocyte inflammatory cell infiltrate

Supplementary Figure 3: Assessment of CD45+ helper T-lymphocyte inflammatory cell infiltrate

Supplementary Figure 4: Assessment of CD68+ macrophage inflammatory cell infiltrate