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1 **Improvement of the performance and microstructural development of**
2 **alkali-activated slag blends**

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16
17 **Abstract:** This study investigated the performance and microstructural
18 development of alkali-activated slag (AAS) blends containing hydromagnesite
19 seeds. AAS blends with and without seeds were assessed by isothermal
20 calorimetry, porosity, setting time and compressive strength measurements. The
21 formation of hydrate phases was further investigated via XRD, TGA-DSC, FTIR
22 and SEM analyses. AAS blends incorporating 2% hydromagnesite seeds revealed
23 the best performance in terms of hydration kinetics, strength and microstructural
24 development. The positive influence of seeds was highlighted via increased
25 hydration rates and degrees, shortened setting times, higher compressive
26 strengths and denser microstructures. These improvements were attributed to the
27 provision of additional nucleation sites in the presence of seeds that enabled the
28 increased formation of hydrate phases (e.g. C-(A)-S-H with more crosslinks
29 forming between adjacent silicate chains) within the pore structure.

30

31 **Keywords:** Alkali activated cement; kinetics; calcium-silicate-hydrate;
32 performance; microstructure

33 **1. Introduction**

34

35 Alkali-activated cements (AACs) are being investigated as an alternative to
36 ordinary Portland cement (OPC), due to the improvements they offer in terms of
37 long-term chemical resistance and lower environmental impact associated during
38 their production [1-3]. Recent studies looked into the use of AACs in foamed
39 products [4] and 3D printed composites [5, 6]. Ground granulated blast-furnace
40 slag (GGBS), produced by quenching molten iron slag from a blast furnace, has
41 been widely used since the emergence of AACs [1]. Different activators such as
42 sodium hydroxide and sodium silicate provide a high pH and thereby increase the
43 solubility and dissolution of slag. This activation process facilitates the formation
44 of hydrate phases such as C-(A)-S-H [7], resulting in a high mechanical
45 performance and chemical resistance [1]. However, some drawbacks of AACs are
46 still present, such as their susceptibility to drying shrinkage at room temperature,
47 whereas recent studies [8] reported a reduction in total drying shrinkage with the
48 inclusion of certain additives such as polyethylene glycol.

49

50 AACs can be considered under calcium- and silica-based and aluminosilicate-
51 based systems. The rate and degree of hydration and the morphology of phases
52 forming in AAS blends determine their mechanical properties due to the formation
53 of different phases [2, 9]. Similar to the hydration of OPC, the hydration of slag
54 under a high pH environment (e.g. > 12) involves the stages of dissolution,
55 induction, acceleration, deceleration and steady state diffusion [10]. However,
56 during this process, the initially formed hydrate phases could create a layer and
57 cover the surface of unreacted slag particles, which slows down the reaction and
58 leads to a lower overall hydration degree.

59

60 One possible solution to this limitation in hydration involves the use of seeds in the
61 initial mix design. These seeds can act as nuclei for the increased precipitation of
62 hydrate phases. Because of its influence in accelerating the hydration kinetics of
63 cementitious materials, nucleation seeding can be an effective approach in

64 enhancing the formation of hydrate phases at early ages [11]. A few studies
65 investigated the effect of seeding in cementitious systems. These involved the use
66 of synthesized C-S-H seeds as nucleation sites in OPC and pure tricalcium silicate
67 (C_3S) pastes [12], as well as AAS blends [13]. The obtained results revealed the
68 benefits of seeding in the acceleration of hydration kinetics via the increased
69 nucleation and growth of hydrate phases, which was previously identified as the
70 rate limiting step during the hydration process. This led to an improved strength
71 development at both early and later stages [12, 13].

72

73 Alternatively, recent studies [14] reported that while the synthesized C-S-H seeds
74 played a role in the further precipitation of C-(A)-S-H, they did not necessarily
75 facilitate the formation of the geopolymer gel. To obtain the C-S-H gel that is
76 introduced into the mix for seeding purposes, the main method followed in the
77 literature involves its synthesis through the reaction of sodium silicate and calcium
78 nitrate [15] or calcium hydroxide and nano-silica [16] in a laboratory environment.
79 This process is composed of several steps involving the washing, filtering and
80 drying of the synthesized material, which is time consuming and is prone to
81 carbonation when exposed to air [15]. Furthermore, differences in the Ca/Si ratio
82 of C-S-H can influence the formation and properties of hydrate phases in the mixes
83 it is introduced in [16], thereby limiting its use on a large scale in practice.

84

85 In addition to C-S-H, some nano particles were also involved as nucleation seeds
86 due to their large surface areas [11, 17]. For instance, the use of nano-silica in
87 OPC increased the hydration degree and rate, which was determined by the
88 measurement of total surface area and calcium concentration [17]. Another study
89 [18] revealed the catalytic effects of nano- Al_2O_3 particles in geopolymer mixes,
90 which enabled an increased amount of gel phase formation within a short period
91 of time due to their large surface areas. In addition to nano- Al_2O_3 , the nucleation
92 and filling effects of nano- $CaCO_3$ and nano- SiO_2 were also reported in PC-based
93 mixes that revealed a more compact and homogeneous structure [19]. Similarly,
94 the use of nano clay [20] and nano metakaolin [21] also contributed to the strength

95 development of PC mixes. A similar effect was found in samples containing
96 polycarboxylates, which can be an alternative admixture for improving the strength
97 of concrete mixes [22]. Further studies [23-25] indicated that seeding was also
98 effective in enhancing the hydration process in calcium-free binder systems.
99 However, nano materials can pose a threat to human health and have a very high
100 cost, presenting a challenge in their use in large scale applications [26, 27].

101

102 Other than nano materials, the use of hydromagnesite ($4\text{MgCO}_3 \cdot \text{Mg}(\text{OH})_2 \cdot 4\text{H}_2\text{O}$)
103 as an additive in cement-based mixes has also been investigated [23, 28]. Other
104 studies [29] looked into the influence of Mg^{2+} on the kinetics of calcite precipitation
105 and morphology. While it can be synthetically produced [30, 31], hydromagnesite
106 is a naturally occurring mineral and is related to other magnesium containing
107 minerals such as serpentine, dolomite and brucite [32]. Furthermore, its lower
108 commercial cost than many of the nano materials such as nano- SiO_2 , TiO_2 and
109 Al_2O_3 presents a benefit for the use of hydromagnesite. Unlike the hydration of
110 pure MgO that leads to the formation of brucite, which does not result in a
111 significant strength gain, a boost in the compressive strength of MgO mixes was
112 seen in the presence of hydromagnesite [33]. Within these MgO -hydromagnesite
113 blends, the formation of an amorphous phase was observed, which could
114 contribute to their cohesive binding [34]. A similar outcome was observed in AAS
115 mixes, where the combined used of MgO and hydromagnesite led to improved
116 performance via the adjustment of the pH to accelerate the dissolution of GGBS
117 [28].

118

119 Differing from the existing literature, this paper focuses on the incorporation of
120 various amounts of hydromagnesite in AAS blends with the goal of identifying its
121 effect on hydration kinetics and associated performance, which has not been
122 investigated before. To reveal the role of hydromagnesite seeds in the
123 improvement of reaction mechanisms, sample performance and microstructural
124 development, the seeded samples were compared with the precursor (i.e. slag
125 blend without any activator or seed) and unseeded AAS blends via the use of

126 different analysis techniques. The effect of nucleation seeding was evaluated by
127 isothermal calorimetry, setting time, porosity and compressive strength
128 measurements. X-ray diffraction (XRD), thermogravimetric analysis and
129 differential scanning calorimetry (TGA-DSC), Fourier transform infrared
130 spectroscopy (FTIR) with spectral subtraction and deconvolution, and scanning
131 electron microscopy (SEM) were used in the analysis of the hydration process and
132 the assessment of the amount and morphology of hydrate phases in both seeded
133 and unseeded samples.

134

135

136 **2. Materials and Methodology**

137

138 **2.1 Materials and sample preparation**

139

140 The GGBS used in this study, whose chemical composition (i.e. obtained from the
141 supplier, Engro Corporation Ltd. (Singapore)) is shown in Table 1, was grade
142 P8000, referring to a super-reactive ultrafine slag powder (average particle size: <
143 5 μm and fineness: 800-900 m^2/kg , which is 2-3 times higher than common GGBS).
144 In line with the findings of a previous study [13], the activator used was sodium
145 metasilicate pentahydrate ($\text{NaSiO}_3 \cdot 5\text{H}_2\text{O}$, > 97%), obtained from VWR Pte. Ltd.
146 (Singapore). The activator was included at 10% of the mass of the binder (slag).
147 In terms of seeds, hydromagnesite ($4\text{MgCO}_3 \cdot \text{Mg}(\text{OH})_2 \cdot 4\text{H}_2\text{O}$, specific surface area
148 (SSA): 43.5 m^2/g) was obtained from Fisher Scientific Ltd. (UK). Different amounts
149 of hydromagnesite, corresponding to 1, 2 and 5% of the mass of the binder
150 component, were used. The morphology of hydromagnesite presented in Fig. 1
151 demonstrated a rosette-like structure, which was in line with those reported in
152 previous studies [35-38].

153

154 The prepared fresh mixture was cast into 50×50×50 mm cubic molds, consolidated
155 by a vibrating table and finished by a trowel. All samples were demolded after 24

156 hours and cured under ambient temperature ($30\pm 1.5^{\circ}\text{C}$) and relative humidity
157 ($80\pm 5\%$).

158

159

160 **2.2 Methodology**

161

162 **2.2.1 Isothermal calorimetry**

163

164 The heat flow corresponding to the hydration of AAS blends was investigated at
165 30°C by an I-Cal 2000 high precision calorimeter according to ASTM C1702-17
166 [39]. To prepare the paste samples, hydromagnesite seeds were first dispersed in
167 the $\text{NaSiO}_3\cdot 5\text{H}_2\text{O}$ solution by an ultrasonic homogenizer (Q500 sonicator),
168 followed by the mixing of the slag into the solution, which was previously heated
169 to 30°C (i.e. room temperature). Once the prepared pastes were ready, they were
170 immediately placed into the calorimeter to record the heat released for up to 36
171 hours.

172

173

174 **2.2.2 Setting time**

175

176 The setting time of AAS blends was evaluated by the measurement of the
177 penetration depth of the Vicat needle into the prepared pastes in accordance with
178 ASTM C191-08 [40]. The initial setting time was calculated to the nearest 1 minute
179 according to Equation 1, where E represented the time (minutes) of the last
180 penetration greater than 25 mm, H represented the time (minutes) of the first
181 penetration less than 25 mm, C represented the penetration reading at time E and
182 D represented the penetration reading at time H. The final setting time, calculated
183 to the nearest 5 minutes, was determined by the time that elapsed between the
184 initial contact of water and cement and the time that the needle was unable to sink
185 visibly.

186

187 Initial setting time = $((H - E)/(C - D)) \cdot (C - 25) + E$ (1)

188

189

190 **2.2.3 Porosity**

191

192 X-ray computed tomography (μ -CT), performed with a SkyScan1183 scanner, a
193 camera, and a flat panel sensor, was employed to analyze the porosity of the
194 prepared blends, including the open and closed pores. Unlike other methods such
195 as Mercury Intrusion Porosimetry (MIP), where the pore distribution is highly
196 influenced by the intrusion pressure, μ -CT is capable of accurately revealing the
197 pore volume and size distribution without damaging the sample [41]. To facilitate
198 this, the specimens obtained from samples crushed during strength testing were
199 fixed on a micro-positioning stage and scanned using a source voltage and current
200 of 80 kV and 100 μ A, respectively. The analysis of porosity involved the process
201 of image acquisition, reconstruction, thresholding, pore labelling and quantification.
202 During image acquisition, a representative 2D projection of each sample (i.e. at a
203 diameter of 25 mm and spatial resolution of 7 μ m) was selected. The thresholding,
204 pore labelling and quantification of porosity were conducted with the CT-Analyzer
205 software.

206

207

208 **2.2.4 Compressive strength**

209

210 The compressive strength of AAS blends was tested at 3, 7 and 14 days. The test
211 was conducted using a compression machine (Toni Technik Baustoffprüfsysteme
212 GmbH), operated at a constant loading rate of 100 kN/min. The average and
213 standard deviation of three samples were reported for each data point.

214

215

216 **2.2.5 XRD, TGA-DSC, FTIR and SEM analyses**

217

218 Hardened paste specimens extracted from the cubic samples crushed after
219 strength testing were stored in acetone to stop hydration and dried under vacuum
220 for 24 hours to prepare for microstructural analysis. Prior to microstructural
221 analysis, the vacuum dried samples were ground down to pass through a 75 μm
222 sieve.

223

224 XRD was recorded on a Philips PW 1800 spectrometer using $\text{Cu K}\alpha$ radiation (40
225 kV, 30 mA) with a scanning rate of 2° 2θ /step from 5° to 80° (2θ). TGA-DSC was
226 conducted on a Perkin Elmer TGA 4000 equipment from 30 to 900°C with a heating
227 rate of $10^\circ\text{C}/\text{min}$ under nitrogen flow.

228

229 A Thermo Scientific Nicolet iS50 spectrometer with a built-in attenuated total
230 reflection (ATR) module was used to obtain Fourier transform infrared
231 spectroscopy (FTIR) spectra and identify the chemical components within the
232 samples. Prior to analysis in the transmittance mode, the preparation of a pellet
233 was conducted consisting of a mixture of 1 mg of powder from the samples with
234 300 mg of KBr. The mixture was pressed using 7 metric tons of hydraulic pressure
235 in a die. In the ATR mode, powder samples were placed directly on the diamond
236 ATR sampling station and pressed against the diamond crystal by a constant
237 compressive force of 267 N. The resolution was set at 4 cm^{-1} and a collection of
238 64 scans was obtained for each spectrum. Spectral subtraction and deconvolution
239 of FTIR were also performed in line with the procedure explained in [42].

240

241 Before SEM analysis, samples were vacuum dried for 24 hours and coated with
242 platinum under a 30 mA current, with a coating time of 30 seconds. The coated
243 samples were then studied via a field emission scanning electron microscope
244 (FESEM, 207 JOEL JSM-7600F). Backscattered electron (BSE) imaging was also
245 used in SEM. To facilitate this, the vacuum dried samples were mounted in epoxy
246 resin for 24 hours and polished to obtain smooth surfaces. Initial polishing was
247 performed with a sandpaper before final polishing by diamond slurry with sizes of
248 9, 3 and $0.3\ \mu\text{m}$. Dust and diamond particles generated during the polishing

249 process were removed by exposing the samples to an ultrasonic bath for 10
250 minutes, followed by vacuum drying for another 24 hours before the analysis.

251

252

253 **2.2.6 pH test**

254

255 The pH values of the activator and hydromagnesite seeds were measured via a
256 calibrated Mettler Toledo pH meter, in line with ASTM C25 [43]. Accordingly, 5
257 grams of powder was blended with 100 grams of distilled water for 30 min prior to
258 each test.

259

260

261 **3. Results**

262

263 **3.1 Isothermal calorimetry**

264

265 The heat flow curves showing the reaction of AAS blends containing different
266 amounts (i.e. 1-5% of the binder content by mass) of hydromagnesite during the
267 first 36 hours are presented in Fig. 2. The hydration of GGBS by itself did not
268 release any noticeable amount of heat, as indicated by the relatively flat heat curve.
269 This was attributed to the low pH of this system, limiting the dissolution of the
270 binder. Alternatively, when activated by the high pH provided by $\text{Na}_2\text{SiO}_3 \cdot 5\text{H}_2\text{O}$, all
271 AAS blends indicated notable hydration peaks, albeit at different times and
272 intensities. The common aspect of all AAS blends was the three stages of the
273 hydration process. The first stage was the pre-induction stage, associated with the
274 dissolution of slag [44]. The length and the timing of this stage depended on the
275 type of the seed. The second stage was the induction period, which was
276 significantly shortened due to the availability of a large number of nuclei provided
277 by the presence of the seeds. The third stage of hydration was the nucleation,
278 growth and precipitation of the hydrate phases. When compared to the unseeded
279 blends, the advances in the hydration of blends containing seeds were associated

280 with the provision of additional nucleation sites supplied by the seeds within the
281 pore space, away from the particle surfaces [12].

282

283 Increasing the amount of hydromagnesite seed from 1% to 2% led to an obvious
284 reduction in the induction period and enabled the acceleration of the hydration
285 reaction. A further increase in the seed content to 5% delayed the appearance of
286 the main hydration peak when compared to blends containing 2% seed. This
287 adverse effect could be associated with the agglomeration of the seed when its
288 quantity was increased, which was in line with the observations reported in
289 previous studies [24]. Accordingly, the high amount of seeds used in these blends
290 could have led to the trapping of the alkali solution within the solid particles, thereby
291 limiting the dissolution of the main binder phase. Therefore, hydromagnesite seed
292 included at 2% of the overall binder content was chosen in the preparation of
293 further mixes, whose details are shown in Table 2.

294

295

296 **3.2 Setting time**

297

298 The initial and final setting time of all samples are presented in Fig. 3. The setting
299 of all mixes generally took place during the acceleration period. In line with the
300 isothermal calorimetry results, the sample without any activator or seed (GA0H0)
301 demonstrated a very long setting time, which was associated with the initially low
302 alkalinity of the pore solution. The incorporation of 10% activator in sample
303 GA10H0 led to an obvious reduction in both the initial and the final setting times
304 due to the increase in the pH of the solution, which enabled the dissolution of the
305 binder and the precipitation of hydrate phases responsible for setting. The use of
306 seeds along with the activator in sample GA10H2 had a much more significant
307 effect on setting time, which was largely reduced. This notable reduction in the
308 setting time was associated with the provision of nucleation sites via the addition
309 of hydromagnesite. This led to an elimination of the induction period, thus reducing
310 the setting time. Furthermore, the presence of hydromagnesite, which acted as

311 nuclei within the pore solution, favored the formation of a higher amount of hydrate
312 phases, which led to the rapid hardening of sample GA10H2. These findings were
313 in line with those of previous studies [45], where it was indicated that the
314 incorporation of fine carbonate fillers led to a rapid increase in viscosity due to the
315 improvement of the packing density of cement-based systems, resulting in shorter
316 setting times in seeded samples.

317

318

319 **3.3 Porosity**

320

321 The typical 2D images of cross-sections (i.e. diameter = 25 mm; spatial resolution
322 = 7 μm) and corresponding porosities (i.e. shown in the lower left corner of each
323 image) of all samples after 14 days of curing are presented in Fig. 4. Since $\mu\text{-CT}$
324 analysis is known to enable the identification and quantification of both closed and
325 open pores within the samples [46], the obtained results were a clear indication of
326 the significant decline in porosity facilitated by the introduction of seeds in the
327 sample GA10H2, where the volume taken up by solid phases was notably
328 increased. This effect was attributed to the seeding effect of hydromagnesite
329 particles, which enabled the abundant formation of hydrate phases within the pore
330 solution. This could be further supported with the micro-filler effect of the seed itself,
331 whose physical presence reduced the void space between large particles [47],
332 albeit at a much smaller scale than the seeding effect. The micro-filler effect was
333 not identified as the major cause of porosity reduction as the porosity of sample
334 GA10H2 was 3.5 times smaller than that of sample GA10H0 (i.e. 16.9% vs. 59.7%),
335 in the presence of only 2% hydromagnesite seeds by weight.

336

337

338 **3.4 Compressive strength**

339

340 Fig. 5 presents the early strength development of all samples up to 14 days. All
341 samples revealed a stable increase in strength with curing age due to the

342 continuation of hydration. Activated (GA10H0) and seeded (GA10H2) samples
343 revealed higher 3-day strengths (17.1-19.1 MPa) than the control sample (GA0H0,
344 11.9 MPa). The notable strength of the control sample without the use of any
345 activator or seeds could be associated with the relatively high reactivity and
346 fineness of the GGBS used in this study. A similar trend was also observed at
347 longer curing periods, during which the effect of seeding became more prominent.
348 In line with the hydration kinetics and porosity results, the introduction of 10% of
349 activator in sample GA10H0 led to ~30% improvement in strength at 14 days due
350 to the increase in the dissolution of slag in the alkaline environment. While the 3-
351 day strengths of samples GA10H0 and GA10H2 were comparable, an increase in
352 the curing age resulted in an obvious improvement in the strength of the seeded
353 sample. Accordingly, sample GA10H2 revealed a 14-day strength of 38.3 MPa,
354 which was 71.7% higher than that of sample GA10H0 (22.3 MPa). This
355 improvement in the presence of seeds was associated with the formation of a
356 continuous network of hydrate phases around the solid particles within the pore
357 solution. Without the addition of seeds, the precipitation of hydrate phases was
358 restricted by the limited content of solid phases within the slag, whose surface area
359 gradually reduced as hydrate phases started growing around them. Alternatively,
360 the provision of additional nucleation sites enabled the further deposition of hydrate
361 phases on the nuclei as well, which was reflected as improved performance in
362 sample GA10H2 when compared to other samples. This could be further enhanced
363 with the filler effect of the seed, which contributed to the densification of sample
364 microstructure, and hence increased the strength in the long term.

365

366

367 **3.5 XRD**

368

369 Fig. 6 presents the XRD patterns of all samples after 14 days of curing. The broad
370 hump located at around $25-35^\circ 2\theta$ observed in sample GA0H0 was attributed to
371 the presence of amorphous phases [48], whereas with the introduction of activator
372 in samples GA10H0 and GA10H2 led to the appearance of C-(A)-S-H at $30-31^\circ 2\theta$

373 (PDF #00-033-0306) as the major hydrate phase, which was in line with the
374 findings of previous studies [49]. The XRD pattern of sample GA10H2 also
375 demonstrated the presence of hydrotalcite ($\text{Mg}_6\text{Al}_2(\text{OH})_{16}\text{CO}_3 \cdot 4\text{H}_2\text{O}$; PDF #01-
376 070-2151), however at a lower intensity when compared with C-(A)-S-H. The Mg^{2+}
377 source in hydrotalcite could be due to the dissolution of the slag. Alternatively, the
378 presence of calcite in samples GA10H0 and GA10H2 (CaCO_3 ; PDF #01-071-3699)
379 could be attributed to the reaction between Ca^{2+} (i.e. from the dissolution of slag)
380 and atmospheric CO_2 via natural carbonation (Equation 2), albeit small due to the
381 low concentration of CO_2 in the atmosphere (~0.04%). The absence of any obvious
382 hydromagnesite peaks in the XRD patterns was due to the low contents of
383 hydromagnesite (i.e. 2% of the binder content) present in the seeded samples, as
384 well as the precipitation of hydrate phases around the seed particles. Overall, the
385 XRD patterns confirmed that the formation of hydrate phases such as C-(A)-S-H
386 was associated with the strength development of the seeded samples.

387



389

390

391 **3.6 TGA-DSC**

392

393 The TGA-DSC results of all samples after 14 days curing are shown in Fig. 7. The
394 following three stages of thermal decomposition were observed:

395

396 (i) 30-220°C: Mass loss due to the dehydration of C-(A)-S-H, with an endothermic
397 peak at ~80°C [49].

398

399 (ii) 220-730°C: Mass loss due to the decomposition of hydrotalcite [49, 50].

400

401 (iii) 730-900°C: Mass loss due to the decarbonation of carbonates (e.g. calcite),
402 with an endothermic peak at ~870°C [24, 51].

403

404 The summary of mass loss values obtained at various stages of thermal
405 decomposition after 14 days of curing is shown in Table 3. The bound water
406 content calculated from the mass loss in C-(A)-S-H could be used as an
407 assessment of the hydration degree in cement blends [50, 52]. In line with the XRD
408 results, the introduction of seeds in sample GA10H2 revealed ~130% increase in
409 the mass loss when compared with unseeded samples. Furthermore, the sharper
410 endothermic peak observed at ~80°C in sample GA10H2 also hinted the presence
411 of a higher amount of C-(A)-S-H than in other samples. Similarly, a slightly higher
412 amount of mass loss corresponding to the decomposition of hydrotalcite was
413 observed in sample GA10H2. The overall decomposition patterns revealed by all
414 samples highlighted the higher contents of hydrate phases, especially C-(A)-S-H,
415 in samples involving the simultaneous use of activator and seeds. These outcomes
416 were in line with the XRD patterns, as well as the isothermal calorimetry, porosity
417 and strength measurements presented earlier.

418

419

420 **3.7 FTIR**

421

422 The FTIR spectra of the seed (hydromagnesite) and all samples after 14 days of
423 curing are shown in Fig. 8(a). The spectra of samples GA10H0 and GA10H2
424 revealed a peak at 800-1100 cm^{-1} , which differed in shape and location (i.e. peak
425 center) when compared with sample GA0H0. This observation was attributed to
426 the stretching vibrations of Si-O [53, 54] due to the formation of C-(A)-S-H gel.
427 Furthermore, a low-intensity band around 590-600 cm^{-1} was demonstrated by
428 samples GA10H0 and GA10H2, indicating the presence of C-A-S-H with the
429 incorporation of Al into C-S-H [55].

430

431 In order to identify the FTIR peaks of the hydrate phases in samples GA10H0 and
432 GA10H2, FTIR spectral subtraction [42] was conducted via subtracting the
433 spectrum of the control sample (GA0H0) from the spectra of GA10H0 and GA10H2,
434 respectively (Fig. 8(b)). As the band between 1080-1180 cm^{-1} was mainly due to

435 the presence of unhydrated binder [42], the presence of hydrate phases could be
436 conservatively defined through spectral subtraction. The spectra of hydrate phases
437 in samples GA10H0 and GA10H2 (i.e. with baseline correction) were further
438 investigated by spectral deconvolution [42] to reveal the individual bands hidden
439 in the broad peaks. Fig. 9 presents these deconvolution results, which were used
440 to indicate the Q values of Si-O. In terms of various spectra assignments, the peak
441 around 860 cm^{-1} was assigned to Q^1 [56], whereas bands at 900 and $950\text{-}980\text{ cm}^{-1}$
442 were unanimously assigned to stretching vibrations of Q^2 sites [57, 58]. In line
443 with the findings of previous studies [42, 59, 60], the peaks at 898 , 947 (945) and
444 985 cm^{-1} were assigned to Q^2 , as multiple Q^2 sites could coexist in C-(A)-S-H. The
445 deconvoluted spectra indicated that Q^2 was the major band in both spectra. In
446 addition to the Q^1 and Q^2 assignments, the peak at 1030 cm^{-1} also indicated the
447 presence of crosslinked Si-O, which was allocated to Q^3 [54] in C-A-S-H gels.
448 Along with the stretching vibrations of Si-O, the 872 cm^{-1} band at a position similar
449 to the bending band of CO_3^{2-} was observed, which usually occurred at around 875
450 cm^{-1} [58]. The noticeably higher peak intensities associated with C-(A)-S-H gel in
451 sample GA10H2, as revealed by Fig. 9, were an indication of a higher yield of C-
452 A-S-H when compared to those of GA10H0. This observation was in agreement
453 with the XRD and TGA findings, where an increased formation of hydrate phases
454 was revealed within sample GA10H2 involving the simultaneous use of activator
455 and seeds.

456

457 Table 4 shows the FTIR spectral deconvolution of the hydrate phases in samples
458 GA10H0 and GA10H2. While the total percentage of Q^2 in the C-(A)-S-H gel
459 observed within both samples were similar, a greater percentage of Q^3 sites in
460 sample GA10H2 was observed. This difference highlighted a higher degree of
461 polymerization of the hydrate phase, referring to the presence of a more-
462 crosslinked C-(A)-S-H gel (i.e. revealing an improved polymerization of the
463 aluminosilicate skeleton) in sample GA10H2 when compared to sample GA10H0.
464 Moving on from previous studies [61], where it was shown that the formation of
465 crosslinked C-(A)-S-H was accountable for high strength gain in AAS blends, this

466 finding could also contribute to the understanding of the improved reaction
467 mechanisms and associated performance of sample GA10H2.

468

469

470 **3.8 SEM**

471

472 Fig. 10 shows the microstructural images of selected samples after 14 days of
473 curing. When compared with the unseeded sample (GA10H0), the introduction of
474 hydromagnesite seeds in sample GA10H2 led to a denser microstructure, resulting
475 in a considerable strength improvement in sample GA10H2, which was in line with
476 the porosity results. The microstructural image of hydromagnesite, shown in Fig.
477 10(c), confirmed the growth of the hydrate phases (i.e. possibly C-(A)-S-H, as
478 indicated by XRD and FTIR results) around the rosette-like hydromagnesite
479 crystals, which grew and covered the surfaces of other particles. Previous studies
480 [62] reported that the adsorption force between Ca^{2+} ions and calcite (CaCO_3) was
481 a moderately strong acid-base (donor-acceptor) interaction, identified as an ionic-
482 covalent bond. This interaction was mainly related with the active surface sites
483 resulting from the presence of CO_3^{2-} in calcite, although its solubility was relatively
484 low [63]. Therefore, it can also be inferred that the interaction between Ca^{2+} ions
485 and hydromagnesite could also be based on an ionic-covalent bond due to the
486 presence of CO_3^{2-} in hydromagnesite ($4\text{MgCO}_3 \cdot \text{Mg}(\text{OH})_2 \cdot 4\text{H}_2\text{O}$). Accordingly,
487 Ca^{2+} ions were first chemically adsorbed via a strong force, which led to a reduction
488 in their mobility, resulting in the formation of “stable nuclei”. This was followed by
489 the precipitation of the C-(A)-S-H gel on the surface of these nuclei, during which
490 Ca^{2+} ions could establish a connection between the formed C-(A)-S-H gel and the
491 hydromagnesite particles by ionic-covalent bonds.

492

493

494 **3.9 pH**

495

496 In addition to its role as a nucleation seed, the dissolution of hydromagnesite may
497 affect the pH of the solution and hence influence the hydration process. To
498 eliminate any such concern, a comparison of the pH values of the activator solution
499 with and without the addition of 2% hydromagnesite seeds was shown in Fig. 11.
500 These measurements revealed stable pH values across the two solutions, indicating
501 that hydromagnesite particles mainly acted as nucleation sites. This was also
502 confirmed by the low pH of the pure hydromagnesite solution, showing that the
503 majority of hydromagnesite particles remained undissolved when incorporated in
504 the activator solution.

505

506

507 **4. Discussion**

508

509 Heterogeneous nucleation can occur through the seeding of the nuclei of new
510 phases into the existing phases, due to a lower free energy barrier [64, 65].
511 Accordingly, two types of heterogeneous nucleation and associated growth of
512 hydrate phases were expected to take place simultaneously within the seeded
513 AAS samples prepared in this study: (i) Hydration that started on the surfaces of
514 the original slag particles, and (ii) additional hydration that started on the surfaces
515 of hydromagnesite seeds distributed amongst the large pores in AAS blends. In
516 line with this expectation, Fig. 12 shows a schematic representation of the
517 hydration process in samples GA10H0 and GA10H2. After several days of curing,
518 unseeded samples (GA10H0) experienced a restriction of further nucleation and
519 growth via the initial formation of a thick dense layer of hydrate phases, leaving a
520 large quantity of capillary pores within the microstructure. This constraint led to a
521 large porosity and inferior strength in comparison to seeded samples (GA10H2),
522 in which the hydrate phases (e.g. C-(A)-S-H gel) nucleated both on the original
523 particle surfaces as well as the seeds. When compared to the unseeded samples,
524 the addition of seeds into AAS blends provided an increased amount of nuclei for
525 the precipitation of additional hydrate phases, thereby greatly reducing the
526 induction time for hydration. This advancement in the hydration process increased

527 the reaction kinetics, which was beneficial for strength development. The
528 nucleation of hydrate phases between the pores not only accelerated the hydration
529 rate and increased the overall hydration degree, but also led to denser
530 microstructures. These findings were in line with the seeding mechanisms
531 previously observed in OPC, C₃S and alkali-activated slag systems [12, 13].

532

533

534 **5. Conclusion**

535

536 This study demonstrated the effect of seeding in the acceleration of the hydration
537 kinetics of AAS blends by investigating the role of different amounts of
538 hydromagnesite as nucleation seeds. Within the seed content range (i.e. 1-5% of
539 the binder content by mass) used in this study, the introduction of 2%
540 hydromagnesite seeds proved to be effective in accelerating the hydration kinetics
541 and increasing the overall hydration degree of the prepared blends. This was
542 enabled via the continuation of hydration on the surface of the original binder as
543 well as the seeds. The improvement observed in the hydration process led to fast
544 setting within AAS blends, which translated into a dense microstructure via the
545 widespread precipitation of hydrate phases (e.g. C-(A)-S-H). When compared to
546 unseeded samples, the increase in the formation of crosslinked C-(A)-S-H was
547 responsible for the high strengths achieved by the seeded samples.

548

549 Overall, the findings of this study highlighted the increased potential of AAS blends
550 to be used in cement-based applications, considering their improved mechanical
551 performance in the presence of a small amount of seeds. Its straightforward
552 incorporation into the initial mix design, ease of application, lack of any safety
553 concerns and lower cost when compared with alternatives (e.g. synthesized C-S-
554 H and nano materials), have led to the identification of hydromagnesite as an ideal
555 choice for enhancing the performance of AAS blends.

556

557

558 **Acknowledgement**

559

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561 MOE Academic Research Fund Tier 1 (RG 95/16) for the completion of this
562 research project.

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List of Tables

Table 1 The chemical composition (wt.%) of GGBS.

	MgO	CaO	SiO ₂	Fe ₂ O ₃	Al ₂ O ₃	R ₂ O ₃	K ₂ O	TiO ₂
GGBS	9.3	37.8	32.3	0.5	14.6	-	-	-

Table 2 The mix designs used in this study.

Sample	Activator content (%)	Hydromagnesite seed content (%)
GA0H0	0	0
GA10H0	10	0
GA10H2	10	2

Table 3 Summary of mass loss obtained at various stages by TGA-DSC after 14 days of curing.

Sample	30-220°C	220-730°C	730-900°C	Total
	Dehydration of C-(A)-S-H	Decomposition of hydrotalcite	Decarbonation of carbonates	
GA0H0	4.2	3.7	1.3	9.2
GA10H0	5.4	5.1	1.0	11.5
GA10H2	12.4	7.0	0.7	20.1

Table 4 FTIR spectral deconvolution of the hydrate phase in samples GA10H0 and GA10H2, provided as wavenumber (cm⁻¹) and peak area percentage.

Band	1	2	3	4	5	6
Hydrate phase in sample GA10H0	840 (7.4%) ^a	-	898 (28.0%)	947 (35.4%)	985 (15.5%)	1030 (13.7%)
Hydrate phase in sample GA10H2	840 (4.3%)	872 (1.4%)	898 (25.1%)	945 (33.5%)	985 (18.8%)	1030 (17.0%)
Assignment ^b	Q ¹	CO ₃ ²⁻	Q ²	Q ²	Q ²	Q ³

^a value in parentheses is the ratio of the component peak area to the broad area.

^b Q denotes SiO₄ tetrahedron, whereas the superscripts indicate the number of other tetrahedron units attached to it.

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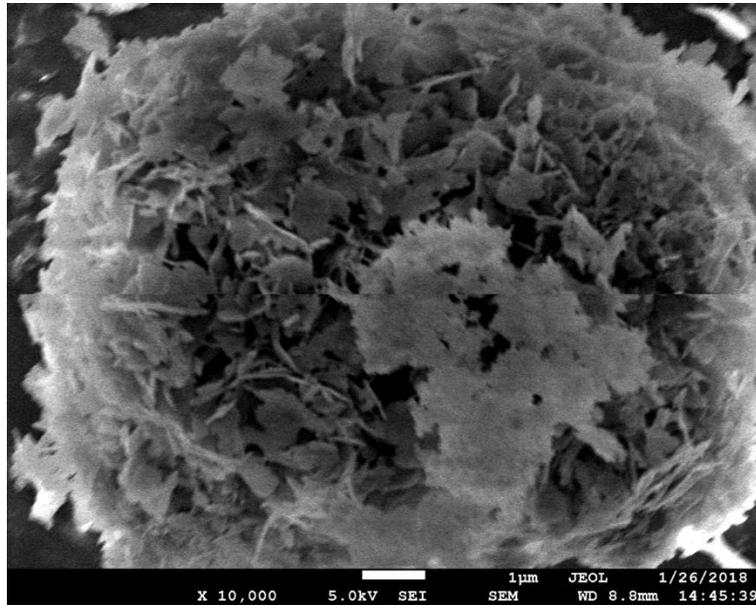


Fig. 1 SEM image of the hydromagnesite seed used in this study

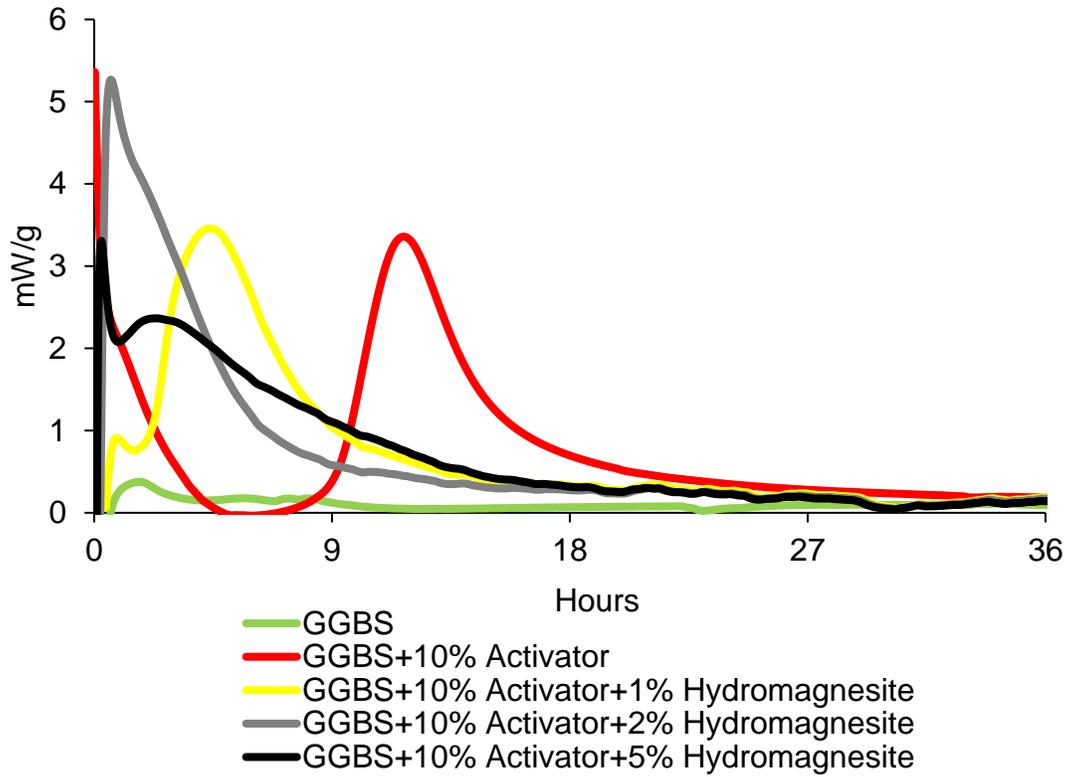


Fig. 2 Heat flow curves of AAS blends containing different amounts of hydromagnesite seed

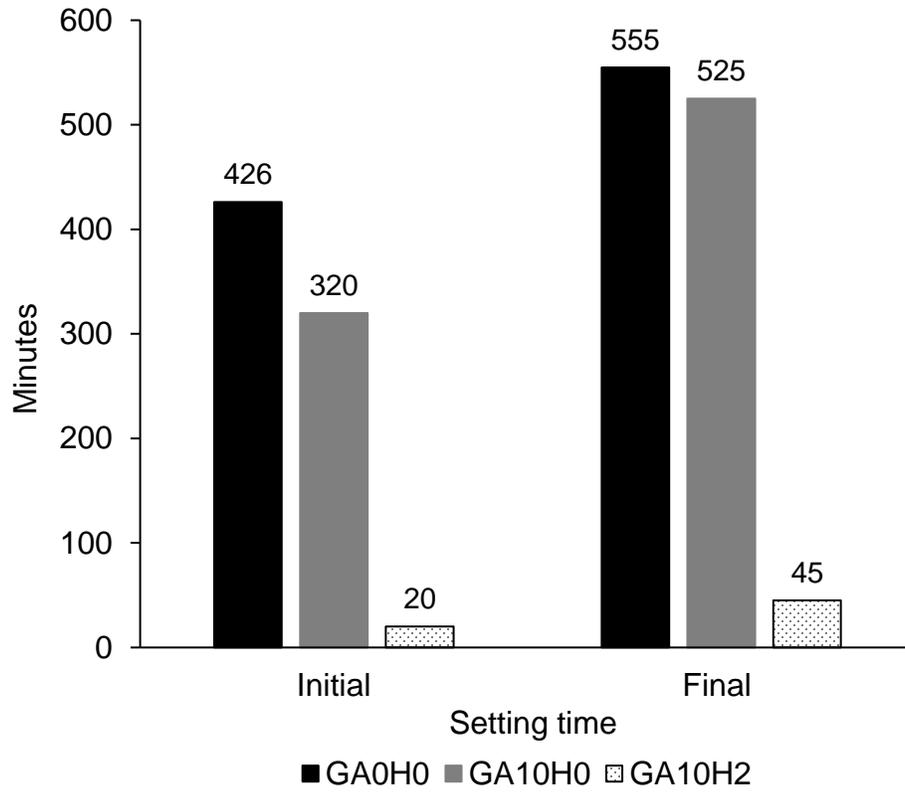


Fig. 3 The setting time of samples with varying amounts of activator and seeds

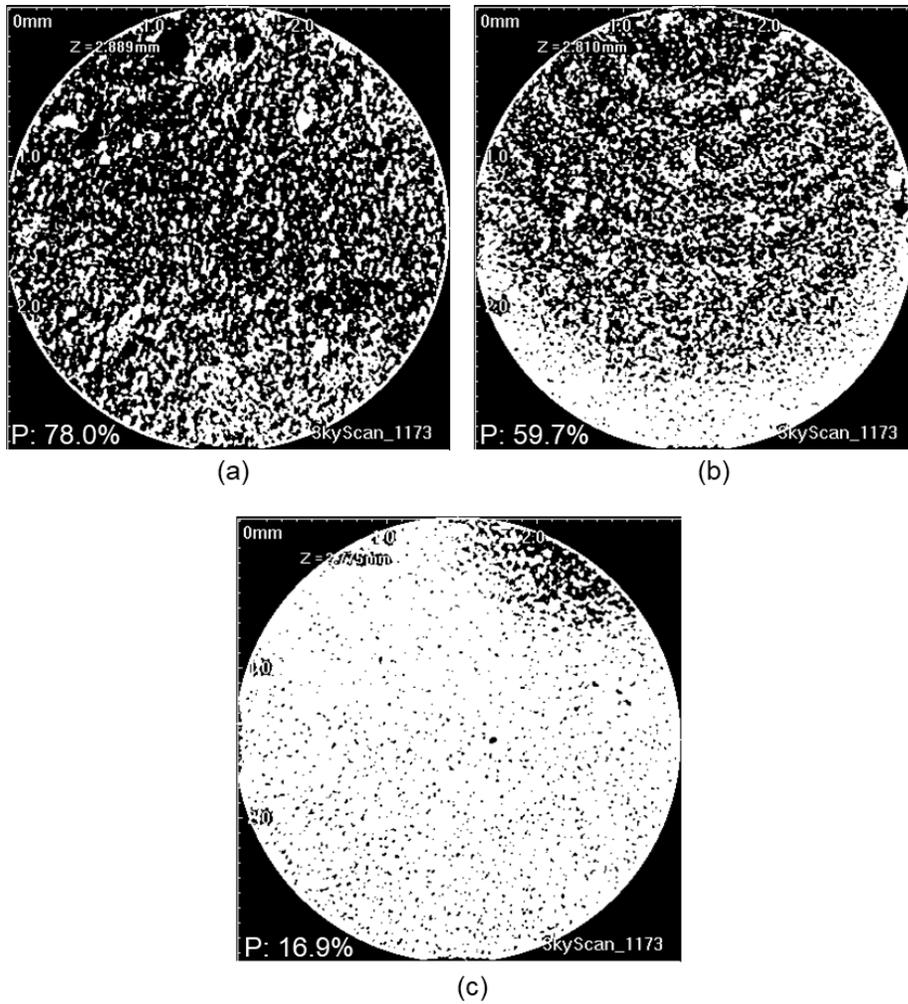


Fig. 4 Typical 2D images of cross-sections (i.e. diameter = 25 mm, spatial resolution = 7 μm) and corresponding porosities (i.e. shown in the lower left corner of each image) of samples (a) GA0H0, (b) GA10H0 and (c) GA10H2 after 14 days of curing

(white area: solid phases; black area: pores)

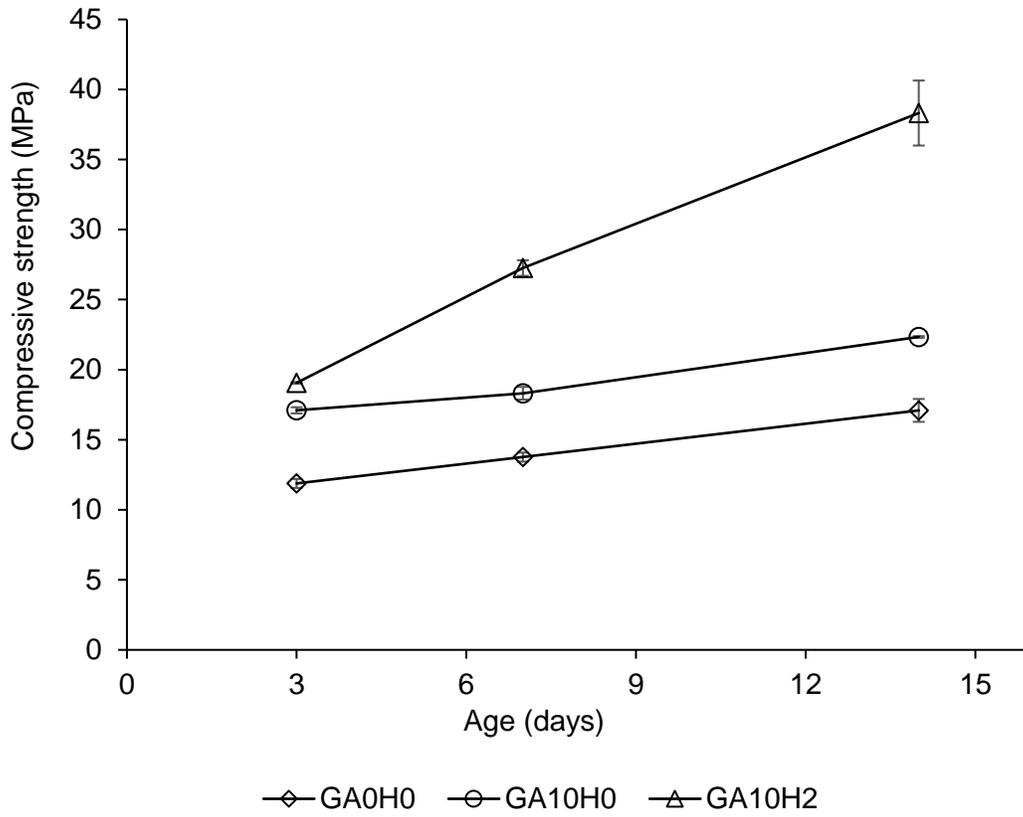


Fig. 5 Compressive strength of samples with varying amounts of activator and seeds during 14 days of curing

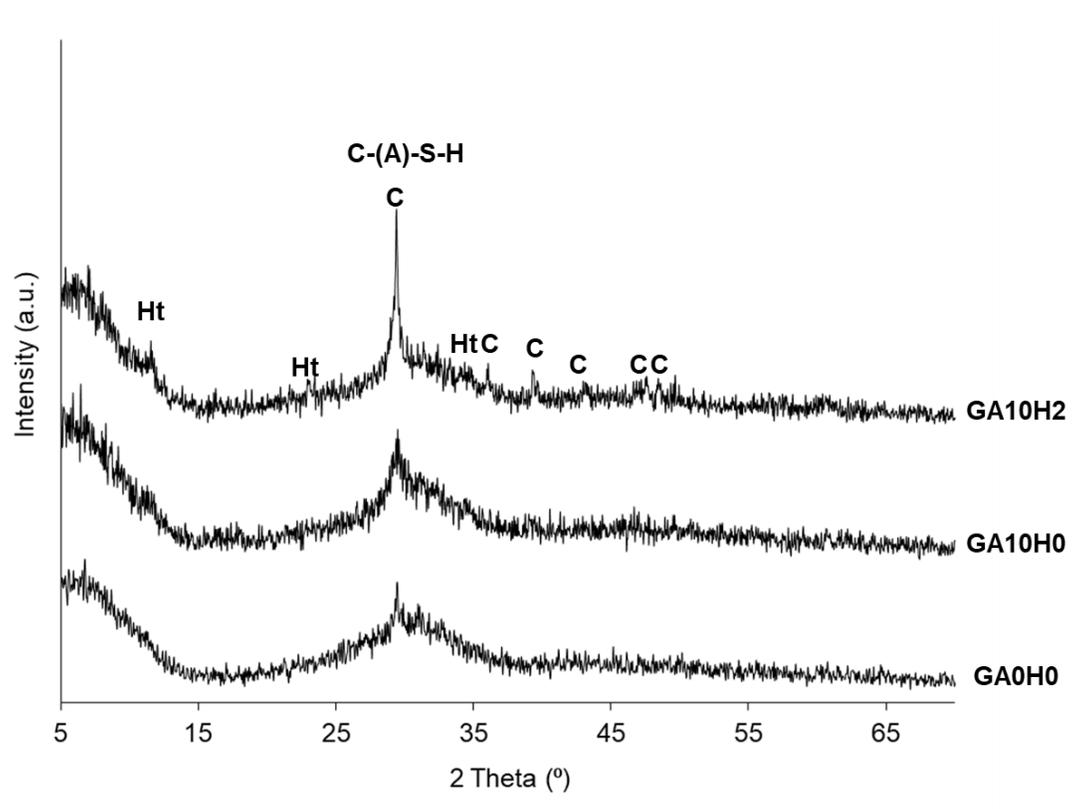


Fig. 6 XRD pattern of samples with varying amounts of activator and seeds at 14 days

(Ht: Hydrotalcite; C: Calcite; C-(A)-S-H: Calcium-(aluminate)-silicate-hydrate)

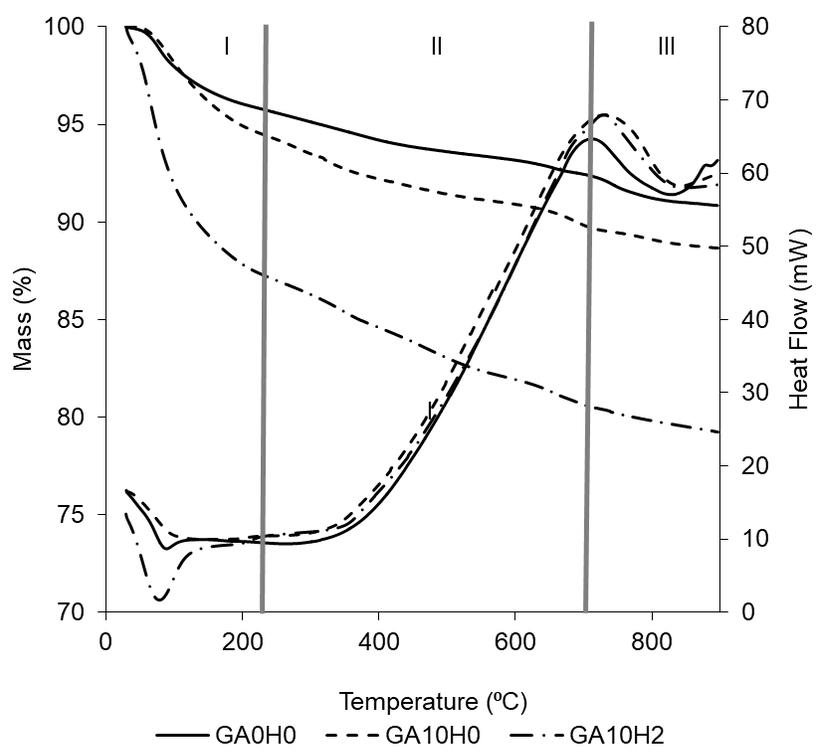
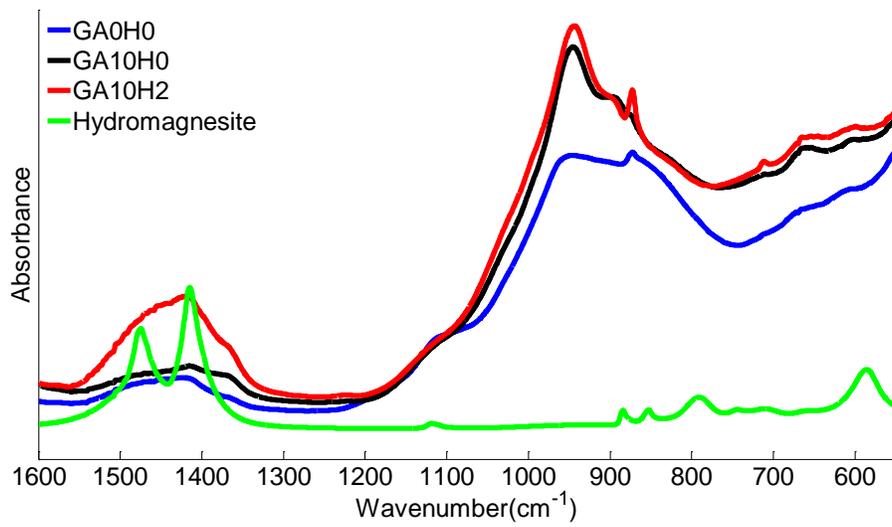
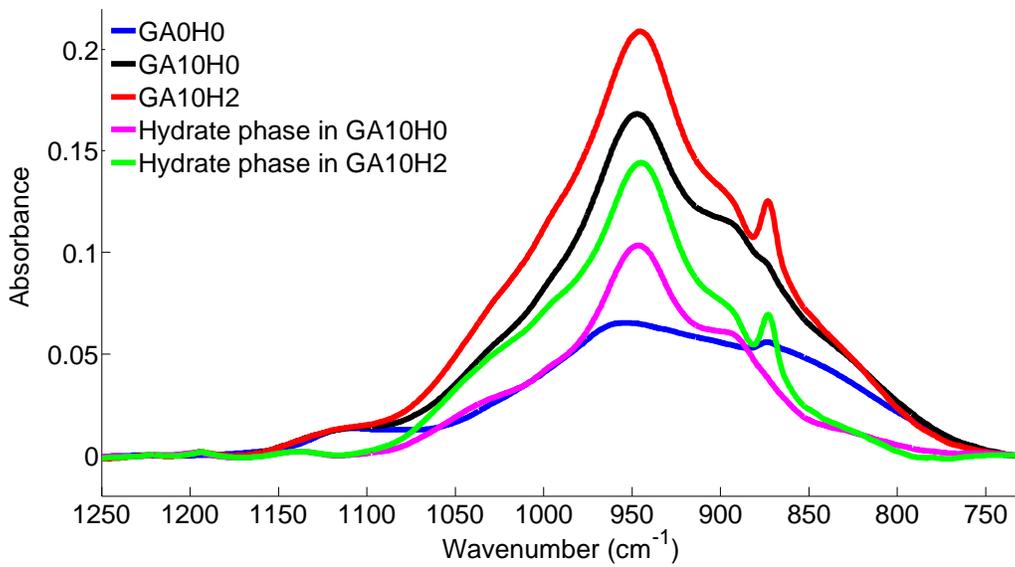


Fig. 7 TGA-DSC results of samples with varying amounts of activator and seeds at 14 days



(a)



(b)

Fig. 8 FTIR spectra of (a) hydromagnesite seed and samples with varying amounts of activator and seeds and (b) subtraction of the spectra of sample GA0H0 from samples GA10H0 and GA10H2 (i.e. after baseline correction) at 14 days

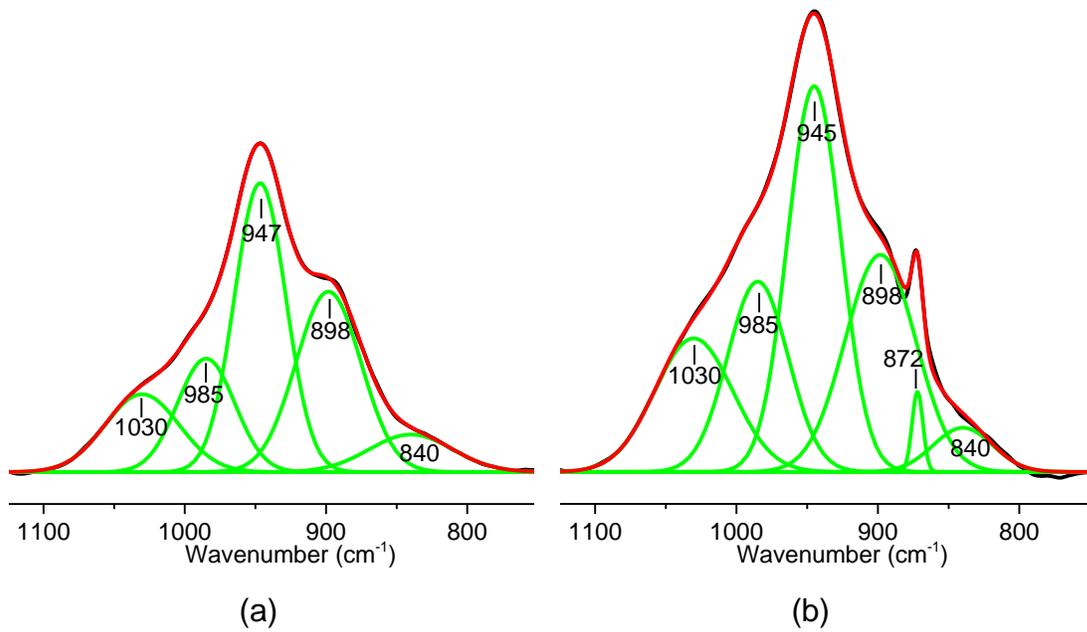
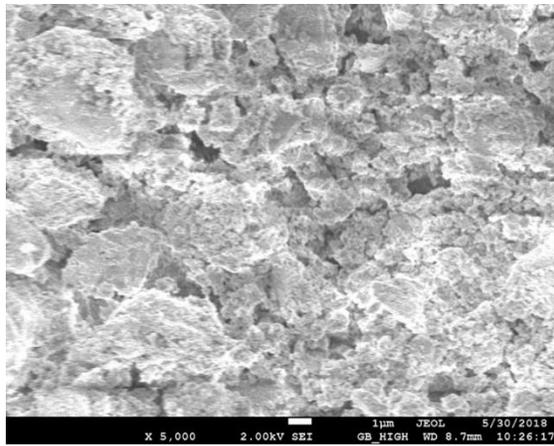
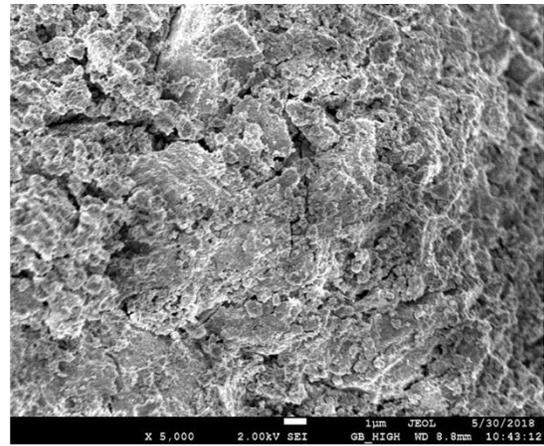


Fig. 9 Deconvoluted FTIR spectra of hydration phases in samples (a) GA10H0 and (b) GA10H2

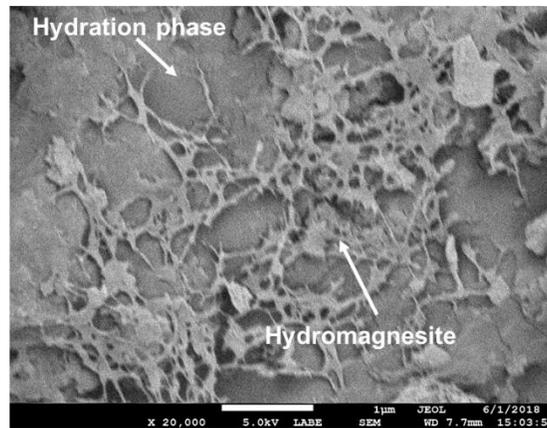
(black curve represents the original spectrum; green curves represent the bands associated with individual components; and red curve represents the sum of all the individual components)



(a)



(b)



(c)

Fig. 10 Microstructural images of (a) sample GA10H0, (b) sample GA10H2 and (c) the surface of a hydromagnesite particle at 14 days

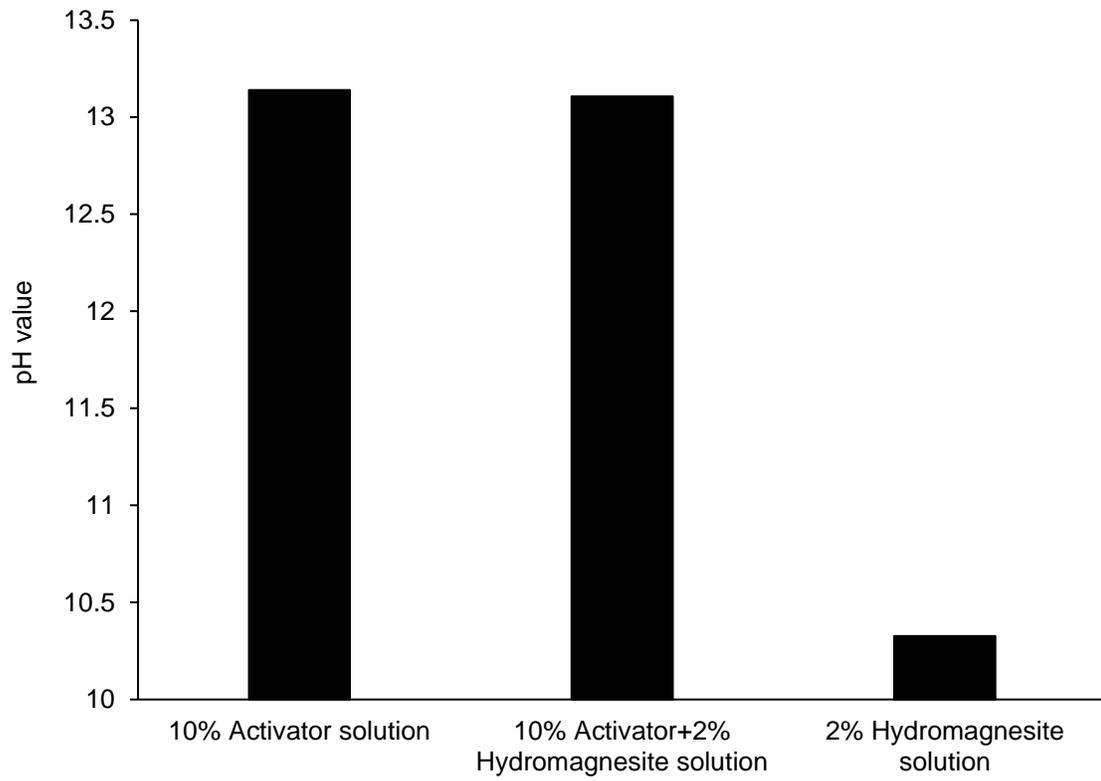


Fig. 11 pH values of different solutions used in this study

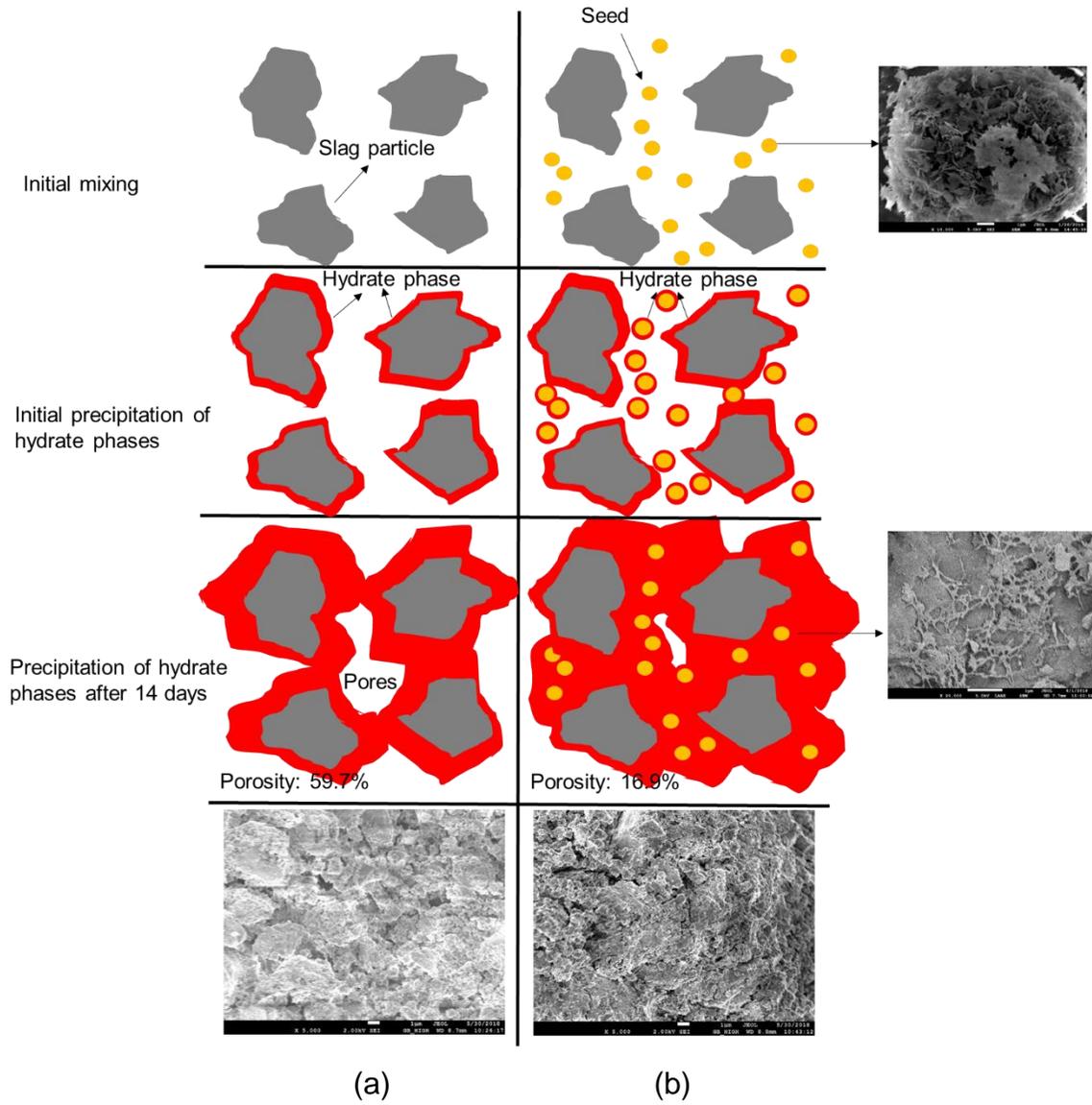


Fig. 12 Schematic representation of the hydration process in samples (a) GA10H0 and (b) GA10H2