



Stenvinkel, P. and Shiels, P. G. (2019) Long-lived animals with negligible senescence: clues for ageing research. *Biochemical Society Transactions*, 47(4), pp. 1157-1164. (doi: [10.1042/BST20190105](https://doi.org/10.1042/BST20190105))

The material cannot be used for any other purpose without further permission of the publisher and is for private use only.

There may be differences between this version and the published version. You are advised to consult the publisher's version if you wish to cite from it.

<http://eprints.gla.ac.uk/193157/>

Deposited on 11 September 2019

Enlighten – Research publications by members of the University of
Glasgow

<http://eprints.gla.ac.uk>

Long lived animals with negligible senescence - clues for ageing research

Stenvinkel P¹, Shiels PG²

¹Division of Renal Medicine, Department of Clinical Science, Intervention and Technology, Karolinska Institutet, Campus Flemingsberg, Stockholm, Sweden.

²Wolfson Wohl Translational Research Centre, University of Glasgow, Gartcube Estate, Switchback Road, Bearsden, Glasgow G61 1QH, UK

Corresponding author:

Peter Stenvinkel, M.D. PhD.

Dept of Renal Medicine M99

Karolinska University Hospital

141 86 Stockholm

Sweden

Abstract

Among several theories to explain the complicated process of human ageing, the mitochondrial oxidative stress hypothesis has received much attention. Considering that lifespan and ageing rates vary considerably across taxa, better understanding of factors that lead to negligible or extremely rapid senescence in mammals, may generate novel approaches to target human ageing. Several species, such as naked mole rats, ocean quahog, rockfish and Greenland shark, have been identified that exhibit negligible senescence and superior resistance to age-related diseases. Considering that the available literature suggests that their outstanding stress resistance is linked to maintenance of protein homeostasis and robust mitochondrial functions, treatments that target protein modification and upregulation of matrix antioxidants may have major implications for extending human health span.

Key words: senescence, long-lived species, naked mole rat, ocean quahog, mitochondrial function, protein homeostasis

Introduction

The principles of ageing and an inescapable advancement towards death have always captivated the interest of mankind. Today the hunt for the “fountain of youth” has been replaced by the search for geroprotectors; i.e. pharmacological or nutraceutical compounds that prevent ageing (1). Though life and health spans have been prolonged in many experimental animals, it has been difficult to conduct studies to test if these regimens also prolong life and health span in humans (2). As the main cause of accelerated mammalian ageing is energy excess, caloric restriction without malnutrition is currently the most effective non-genetic intervention to delay ageing phenotypes in a wide range of mammals from flies (3) to primates (4). A problem with this strategy is that caloric restriction may have adverse effects (4) and reduce quality of life in humans. An alternative approach to arrest ageing would be to use drugs that have caloric restriction mimetic effects (5). As sirtuins are believed to mediate the pro-longevity effect of calorie restriction, sirtuin activators have attracted interest within gerontology. As the human ageing process is slow and gradual, clinical trials of anti-senescence drugs (6), nuclear factor erythroid 2-related factor 2 (NRF2)-agonists (7), SIRT1 agonists (8) and/or dietary interventions that prolong life span in animals, such as caloric (4) and methionine (9) restriction, would need to be extremely long.

Considering that the global number of people >80 years old will triple by 2050, ageing is a considerable challenge for biomedical research that will have a dramatic influence on human morbidity and mortality (10). Diseases of ageing are influenced by mechanisms that show strong evolutionary conservation and ageing is the main risk factor for a number of burden of life style diseases, such as cancer, cardiovascular disease (CVD), obesity, type-2 diabetes, osteoporosis, Alzheimer and chronic kidney disease (CKD), that have become exceedingly prevalent in the modern society (7). Detailed studies of relatively large groups of patients displaying features of accelerated ageing, such as CKD, may be an opportunity to learn more about ageing processes (11). The majority of chronic diseases that accumulate with age share common underpinning mechanisms, such as constant inflammation, oxidative stress, mitochondrial dysfunction, metabolic imbalances and senescence (12). The role of cellular senescence in diseases that accumulate with age growing topic in gerontology, indicative of cellular senescence being integral to the physiology of ageing, longevity and diseases that accumulate with age, either by protein-protein interactions, allocation of common genes or prevailing signalling pathways (13). A major problem in modern health care is the growing gap between “health span” and “life span”; a gap that has major economic and societal implications. Thus, rather than focus on increasing life span, gerontology research should aim to increase health span. Strategies that target central ageing structures in the whole cluster of burden of life style diseases would likely reduce the gap between “health span” and “life span” (7).

Studies of longevity in the animal kingdom – could it provide clues for human ageing?

Studies of long-lived animals that stay healthy and capable of reproduction until death (i.e. long health span) could provide clues on how to target central ageing processes (7). Although the life-span trajectories in different model organisms depict remarkable similarities in the ageing process with humans (14), many wild animals may not have the time to develop features of senescence, as they die of disease, are predated or starve before they reach sufficiently advanced years (15, 16). In order to activate extra resources for survival during stressful periods of the animals life, such as hibernation, it has been speculated that ageing programs are dampened during such periods (17).

One of the biggest challenges in gerontology is to identify changes in cells that are related to senescence (cell cycle arrest with loss of a cells ability of division and growth), instead of just chronology (18). Senescence has been considered trivial in wild animals. However, current evidence implies that it is widespread in natural populations of mammals and birds, affecting survival and reproduction of older individuals. A problem with existing data on longevity in animals is that they often consist of maximum values for life-span (usually single observations) rather than sound and statistically robust estimates of their mean longevity (19). Delineating the physiological and biochemical causes of ageing process in animals is a developing area of research for several reasons, including the possible assets for human health, the relation of ageing to growth and reproduction and the responses of organisms to natural conditions and stress.

From our archaic past, when our common nematode-like ancestors aged exceptionally rapidly, mutations have expanded human lifespan more than 2000-fold (20). Given that protracted longevity in animals is associated with augmented stress resistance, longevity requires uninterrupted and large-scale alterations of organs to suspend senescence. Studies of animals with particularly long life spans would be an approach to better understand mechanisms linked to superior functionality and learn about ageing processes and find central targets for extending health span (21). In the following we discuss what we can learn from animal species that during evolution have adapted systems that render them extraordinary long-lived for their size (**Fig 1**).

Naked mole rats

Among long-lived animals, none has received as much recent attention as the naked mole rat (*Heterocephalus glaber*), a native of Eastern Africa that has emerged as a model organism for studies of ageing-related diseases and negligible senescence (22). In contrast to other mammals, naked mole rats do not conform to the Gompertzian law of age-related mortality risk (23); i.e. their risk of death does not increase with age. During evolution these rodents have adapted to severe constant hypoxia in underground burrows with limited gas exchange. They can survive up to 18 minutes of anoxia without apparent injury, due to fructose-driven glycolysis (24). It has been reported that HIF-1 α activation is also

an important mechanism for their remarkable hypoxia adaptation (25). Naked mole rats live >5 times longer than similarly sized rodents and >9 times longer than laboratory rats and show sustained good health during their lifetime. Notably, naked mole rats are subjected to high levels of oxidative stress and show increased levels of oxidative damage and less robust repair mechanisms than their short-lived rodent counter-parts, but still maintain a superior health span; this is not consistent with the free radical theory of ageing (26). Beside superior anoxia tolerance they are resistant to age-related diseases, such as neurodegeneration, CVD and cancer (21). Out of 1000's of necropsies only two cases of cancer have been reported in naked mole rats (27). In addition, naked mole rats experience no changes in body composition, or decline in genomic and proteomic integrity due to elevated proteasome quality control mechanisms (28). Their diet (well-off in tubers and roots) provide not only a rich source of anti-inflammatory and anti-oxidative polyphenols, but is also restricted in calories and methionine. It is likely that their diet in combination with low body temperature in a hypoxic and hypercapnic milieu has resulted in a series of metabolic adaptations that contribute to their exceptional resilience against age-associated diseases. Efficient protein stability and integrity with less urea-induced protein unfolding, less protein ubiquitination and higher proteasome activity (29), high levels of autophagy (30), high hyaluronan skin expression (31), a superior capacity for mitochondria to consume H₂O₂ (32) and overexpression of the transcription factor NRF2 (22) are reported factors liable for maintaining superior damage control mechanisms (**Fig 1**).

Secrets of longevity found in oceans

Fish, which are the most diverse class of vertebrates with >20,000 existing species, provide an enormous spectrum of longevity that could be used for gerontology research. Although some marine animals, such as sea turtles, sharks and sturgeons are reported to have a long life-span, they have not been extensively used for studies of ageing. In contrast to the common contention that turtles (*Chrysemys picta*) do not age, data spanning >20 years from a wild population of turtles has shown that survival declines among adults (33). Fish exhibit three types of senescence; rapid (such as eels, killifish and pacific salmon), gradual (such as guppy and platyfish) and negligible (such as sturgeons and rockfish). Given recent data indicating that senolytic drugs, such as quercetin and navitoclax, may be an option to arrest human ageing processes (34) comparative studies of fish with rapid or negligible senescence would be of the interest (35). Whereas lower environmental temperature and dietary restriction retards the ageing process in species showing gradual senescence, data on why only some species of fish exhibit negligible senescence is sparse (35). The existence of fish with negligible senescence allow extensive studies of cellular, genetic and biochemical characteristics of slow ageing. It has been reported that muscle extracts from fish with negligible senescence, such as sturgeon, have protective effects on senescence induced by oxidative stress (36). It would also be interest to find out if long-lived fish exhibit

an enhanced capacity to repair DNA damage, protect protein function or exhibit a continued turnover of tissue. The rockfish is a long-lived species with a documented age range of 100-200 years, that has attracted interest as a model system due to intermediate growth (i.e. it continues to increase in body size throughout life) and limited reproductive senescence. In contrast to other mammals, gamete production in female rockfish increases with age and size fecundity is positively correlated with age (37). Among long-lived fish, sharks also have received recent attention since they live much longer than realized previously (38). Radiocarbon studies of the eye lens in the Greenland shark (*Somniosus microcephalus*) have show that they may reach an age of almost 400 years; i.e. they are the longest-lived vertebrate documented (39) and surpass the maximum age of >200 years reported in bowhead whales (*Balaena mysticetus*) (40).

The most long-lived marine species are bivalve molluscs, such as the ocean quahog, which provide the widest range of lifespans available for interspecies comparisons. The maximal lifespan in different populations of bivalve molluscs (*Spisula solidissima* vs *Arctica Icelandica*) range between 36 to >500 years (age is counted by growth rings on the shell) despite similar size and living conditions (41). Thus, *Arctica Icelandica* is the longest-lived non-colonial metazoan known on the planet. As bivalves accommodate a protracted sphere of environmental conditions and express markedly contrasting metabolic phenotypes they constitute a valid model to study ageing processes (41). Their exceptional longevity appears independent of telomere length regulation (42). Moreover, whereas their longevity-related differences in peroxidation index seem resilient to environmental conditions, such as temperature, season, nutrition and age (43) they are resistant to multi-stress genotoxicity (44). To elucidate the underpinning features of successful cardiac ageing, age-related changes in proteasome activity, inflammatory markers, heat shock proteins and mitochondrial complexes were analysed in the heart of *A. Icelandica* (45). The results showed that maintenance of protein homeostasis preserves cardiac function and that levels of oxidatively damaged proteins did not change over the first 120 years of their life (45). Treaster et al (46) have also emphasized the relationship between proteostasis and longevity through estimation of function, unfolding and aggregation. As inhibition of Hsp90 protects from atherosclerosis via cytoprotective mechanisms that are NRF2-dependent (47) and SKN-1/NRF2 is essential for the health span gains provided by metformin treatment in *C. elegans* (48), a role for NRF2 as a pro-longevity signalling pathway across phyla is supported. Indeed, a transcriptomics-based screening recently identified that inhibition of Hsp90 deferred ageing by augmented protein homeostasis in *C. elegans* (1).

Free radical and mitochondrial theories of ageing

According to the free radical theory of ageing (reactive oxygen species [ROS] arise as bi-products of metabolism), long-lived species should be more resistant to oxidative damage and senescence than

short-lived species. As research using 18 different genetic manipulations of the antioxidant defense system showed no effect on life span in mice (except deletion of the *Sod1* gene), existing evidence argues against the hypothesis that oxidative damage plays a dominant role in ageing (49). Data from animal species other than mice have shown equivocal results. It was reported that augmented resistance to oxidative stress and maintenance of protein homeostasis preserves cardiac function in *A. Icelandica* (45) and may be a prerequisite for their extreme longevity (50). When differences in markers of oxidative stress were compared in mollusc species with a six-fold difference in life span, Gruber et al (50) reported correlations between DNA oxidation and biological, but not chronological age, implying that nucleic acid oxidation reflected intrinsic ageing mechanisms. On the other hand, since surrogate markers of oxidative damage in red blood cells and muscle did not correlate to lifespan in Greenland sharks, the free radical theory of ageing may not apply to all long-lived species (51). In the case of the Greenland shark, oxidative status may rather be shaped to the life in cold and deep arctic water (51).

It has been suggested that the cumulative effect of biochemical damage in mitochondria causing mitochondrial mutations plays a key role in many chronic diseases associated with premature ageing, such as CKD, type-2 diabetes and neurodegenerative diseases. The transcriptional coactivator peroxisome proliferator-activated receptor gamma co-activator 1 ($PGC1\alpha$), the master regulator of mitochondrial biogenesis, allow animals to conform their energy metabolism to their current state of nutrition and their environment. It is likely that the evolution of differences in the the distribution and density of mitochondria play a major role for longevity, aerobic capacity and protection in extreme conditions. In the animal kingdom, the levels of mitochondrial enzymes vary more than 100-fold, from fish white muscle to the flight muscles of birds (52). The Atlantic bluefin tuna (*Thunnus thynnus*) is a migratory fish with a remarkable swimming endurance and high metabolic demands, which stands out from other fish because they possess red muscle with an exceptionally high content of mitochondria. In a recent study, Dumesic et al (53) have demonstrated that the upstream open reading frame (uORF), which slows down production of $PGC1\alpha$, was completely absent in the Atlantic blue fish tuna. Their finding indicates that disruption of uORF may be an important evolutionary adaptation in the animal kingdom that allow high metabolic demands.

If the rate of mitochondrial mutation is negatively correlated with longevity, the accumulation of mitochondrial mutations should associate with the degree of senescence (13). Indeed, Blier et al (41) have reported lower ROS production and a lower mitochondrial membrane peroxidability index in the long-lived *A. Icelandica*, supporting the concept that mitochondrial dysfunction-mediated dysregulation of molecular and metabolic pathways boosts senescence. Considering that *A. Icelandica* produces markedly lower rates of H_2O_2 compared to short-lived species and carries mitochondria that are remarkably efficient in energy transduction, extreme longevity seems to be associated with robust and efficient mitochondria that produce low levels of H_2O_2 and ROS (41). Given the reported links between

mitochondrial dysfunction and cellular senescence (54), and recent data showing that NRF2 activation plays a role in mitochondrial quality control (55), it is not surprising that the NRF2 signalling pathway plays an important role in the fine tuning of gene expression in bivalve molluscs (56). In accordance, a superior antioxidant mitochondrial defence system (32) together with overexpression of NRF2 (22) characterizes the exceptionally long-lived naked mole rat. The evolution of kelch-like ECH-associated protein 1 (KEAP1) provided mammals with a way regulate NRF2 activity more precisely (57). Considering that studies on rockfish (*Sebastes*) show that their lifespan is negatively correlated with the rate of mitochondrial DNA mutations (37), this further support the concept that mitochondria play a main role for longevity in the animal kingdom. Data in birds and mammals corroborate these findings and show an inverse relationship between longevity and evolutionary rate of mitochondrial proteins (58). A comparative study in 21 bird species showed that red blood cells from long-lived birds produced less mitochondrial ROS than red blood cells of short-lived birds (59). However, although the majority of studies report that a reduced life span in mammals is related to an increase in somatic mitochondrial mutations (60) (61), Vermulst et al (62) have reported that mitochondrial point mutations do not limit the natural lifespan of mice. In accordance with the latter study, a study in *C. elegans* reported that mitochondrial DNA damage does not control lifespan (63).

What can we learn from species with accelerated aging?

Better understanding of senescence and life history strategies of remarkable short-lived species could also provide valuable clues about factors which favoured selection of species with a reduced life span. Among longevity minimalists, mayflies (*Ephemeroptera*) are exceptional, since these rapidly senescencing organisms have a life span of only 1-2 days (64). For the purpose of screening the potential of pharmaceutical agents with senescence-retarding effects, the extraordinary short-lived killifish (*Notobranchius furzeri*), living in shallow transitory pools on the Savannah in south-eastern Africa, is an interesting vertebrate in ageing research. This seasonal turquoise fish is the shortest-lived vertebrate that can be cultured in captivity and has a maximal life-span of 13 weeks. Their early sexual maturation is tied with accelerated development and expression of ageing biomarkers at a cellular level (65). Since killifish significantly resemble mammals (including humans) with respect to the molecular and cellular traits of ageing, they are of definite interest to gerontology research (66). It is notable that killifish do not display significant age-related decline in reproductive functions during their extremely short lifespan (67). Furthermore, considering their documented high degree of Klotho gene conservation they may serve as a model system for studies of molecular mechanisms and pathways that determine the anti-ageing effects of klotho (68). Baumgart et al (69) have performed a longitudinal study in killifish correlating variations in gene expression and found that mitochondrial respiratory chain complex 1 served as a hub for a group of genes whose expression correlated negatively with lifespan. This

observation further strengthens the hypothesis that the mitochondrion is central to ageing processes and is in keeping with the original mitochondrial, telomere ribosome biogenesis (MTR) postulate of Shiels & Davies (70). This espouses that interplay between these three intracellular features governs intracellular energy availability and thus determines the capacity of a cell for proteosynthesis in response to macromolecular damage and/or redox stress. Taken together, killifish seems to be a valid vertebrate model to identify pharmacological modulators and protein targets associated with aging. Indeed, supplementation with a sirtuin agonist (resveratrol) prolonged lifespan and retarded the expression of age-dependent attributes in killifish (65). The observation that recolonizing the gut of middle-age killifish with bacteria from younger killifish resulted in an extension of life span (71) support hypotheses suggesting links between gut microbiome and longevity in the animal kingdom.

Perspectives

- Ageing is a considerable challenge for biomedical research that will have a dramatic influence on human morbidity and mortality.
- An outstanding stress resistance is a common trait of long-lived mammals and recent literature position maintenance of protein homeostasis and robust mitochondrial function in the centre of preservation of long life in the animal kingdom (Figure 1).
- Better understanding of the biology of exceptionally long-lived animals with negligible senescence may contribute to better understanding of ageing processes and lead to novel interventions that extend human health span.

Acknowledgements

The Heart and Lung Foundation supported Peter Stenvinkel's research.

Legend to Figure:

Reported characteristics of four long-lived animals with negligible senescence.

References

1. Janssens GE, Lin XX, Millan-Ariño L, Kavšek A, Sen I, Seinstra RI, et al. Transcriptomics-Based Screening Identifies Pharmacological Inhibition of Hsp90 as a Means to Defer Aging. *Cell Rep.* 2019;27:467-80.
2. Kirkland JL. Translating the Science of Aging into Therapeutic Interventions. *Cold Spring Harb Perspect Med.* 2016;6:a025908.
3. Lian T, Wu Q, Hodge BA, Wilson KA, Yu G, Yang M. Drosophila Gut-A Nexus Between Dietary Restriction and Lifespan. *Int J Mol Sci.* 2018;19:pii: E3810.
4. Pifferi F, Terrien J, Perret M, Epelbaum J, Blanc S, Picq JL, et al. Promoting healthspan and lifespan with caloric restriction in primates. *Commun Biol.* 2019;2:107.
5. Handschin C. Caloric restriction and exercise "mimetics": Ready for prime time? *Pharmacol Res.* 2016;103:158-66.
6. Zhu Y, Tchkonina T, Pirtskhalava T, Gower AC, Ding H, Giorgadze N, et al. The Achilles' heel of senescent cells: from transcriptome to senolytic drugs. *Aging cell.* 2015;14:644-58.
7. Stenvinkel P, Meyer CJ, Block GA, Chertow GM, Shiels PG. Understanding the role of the cytoprotective transcription factor NRF2 - Lessons from evolution, the animal kingdom and rare progeroid syndromes. *Nephrol Dial Transpl.* 2019;In Press.
8. Lee SH, Lee JH, Lee HY, Min KJ. Sirtuin signaling in cellular senescence and aging. *BMB Rep.* 2019;52:24-34.
9. Lee BC, Kaya A, Gladyshev VN. Methionine restriction and life-span control. *Ann N Y Acad Sci.* 2016;1363:116-24.
10. Niccoli T, Partridge L. Ageing as a risk factor for disease. *Current Biology.* 2012;22:R741-R52.
11. Kooman JP, Kotanko P, Schols AM, Shiels PG, Stenvinkel P. Chronic kidney disease and premature ageing. *Nat Rev Nephrol.* 2014;10:732-42.
12. Fougère B, Boulanger E, Nourhashémi F, Guyonnet S, Cesari M. Chronic Inflammation: Accelerator of Biological Aging. *J Gerontol A Biol Sci Med Sci.* 2017;72:1218-25.
13. Tacutu R, Budovsky A, Yanai H, Fraifeld VE. Molecular links between cellular senescence, longevity and age-related diseases - a systems biology perspective. *Aging (Albany NY).* 2011;3:1178-91.
14. Mitchell SJ, Scheibye-Knudsen M, Longo DL, de Cabo R. Animal models of aging research: implications for human aging and age-related diseases. *Annu Rev Anim Biosci.* 2015;3:283-303.
15. Finch CE. Update on slow aging and negligible senescence - a mini review. *Gerontology.* 2009;55:307-11.
16. Buffenstein R. Negligible senescence in the longest living rodent, the naked mole-rat: insights from a successfully aging species. *J Comp Physiol B.* 2008;4:439-45.
17. Skulachev MV, Severin FF, Skulachev VP. Aging as an evolvability-increasing program which can be switched off by organism to mobilize additional resources for survival. *Curr Aging Sci.* 2015;8:95-109.
18. Jeyapalan JC, Sedivy JM. Cellular senescence and organismal aging. *Mech Ageing Dev.* 2008;129:467-74.
19. Che-Castaldo JP, Byrne A, Perišin K, Faust LJ. Sex-specific median life expectancies from ex situ populations for 330 animal species. *Sci Data.* 2019;6:190019.
20. Kenyon CJ. The genetics of aging. *Nature.* 2010;464:504-12.
21. Stenvinkel P, Painer J, Kuro-O M, Lanasa M, Arnold W, Ruf T, et al. Novel treatment strategies for chronic kidney disease: insights from the animal kingdom. *Nat Rev Nephrol.* 2018;14:265-84.
22. Lewis KN, Wason E, Edrey YH, Kristan DM, Nevo E, Buffenstein R. Regulation of Nrf2 signaling and longevity in naturally long-lived rodents. *Proc Natl Acad Sci U S A.* 2015;112:3722-7.
23. Ruby JG, Smith M, Buffenstein R. Naked Mole-Rat mortality rates defy gompertzian laws by not increasing with age. *Elife.* 2018;7.
24. Park TJ, Reznick J, Peterson BL, Blass G, Omerbasic D, Bennett NC, et al. Fructose-driven glycolysis supports anoxia resistance in the naked mole-rat. *Science.* 2017;356:307-11.
25. Xiao B, Wang S, Yang G, Sun X, Zhao S, Lin L, et al. HIF-1 α contributes to hypoxia adaptation of the naked mole rat. *Oncotarget.* 2017;8:109941-51.

26. Rodriguez KA, Wywiał E, Perez VI, Lambert AJ, Edrey YH, Lewis KN, et al. Walking the oxidative stress tightrope: a perspective from the naked mole-rat, the longest-living rodent. *Curr Pharm Des.* 2011;17:2290-307.
27. Delaney MA, Ward JM, Walsh TF, Chinnadurai SK, Kerns K, Kinsel MJ, et al. Initial Case Reports of Cancer in Naked Mole-rats (*Heterocephalus glaber*). *Vet Pathol.* 2016;53:691-6.
28. Lewis KN, Andziak B, Yang T, Buffenstein R. The naked mole-rat response to oxidative stress: just deal with it. *Antioxid Redox Signal* 2013;19:1388-99.
29. Pérez VI, Buffenstein R, Masamsetti V, Leonard S, Salmon AB, Mele J, et al. Protein stability and resistance to oxidative stress are determinants of longevity in the longest-living rodent, the naked mole-rat. *Proc Natl Acad Sci U S A* 2010;106:3059-64.
30. Zhao S, Lin L, Kan G, Xu C, Tang Q, Yu C, et al. High autophagy in the naked mole rat may play a significant role in maintaining good health. *Cell Physiol Biochem* 2014;33:321-32.
31. Tian X, Azpurua J, Hine C, Vaidya A, Myakishev-Rempel M, Ablueva J, et al. High-molecular-mass hyaluronan mediates the cancer resistance of the naked mole rat. *Nature.* 2013;499:346-9.
32. Munro D, Baldy C, Pamenter ME, Treberg JR. The exceptional longevity of the naked mole-rat may be explained by mitochondrial antioxidant defenses. *Aging Cell* 2019;18:e12916.
33. Warner DA, Miller DA, Bronikowski AM, Janzen FJ. Decades of field data reveal that turtles senesce in the wild. *Proc Natl Acad Sci U S A* 2016;113:6502-7.
34. Tchkonja T, Kirkland JL. Aging, Cell Senescence, and Chronic Disease: Emerging Therapeutic Strategies. *JAMA.* 2018;320:1319-20.
35. Patnaik BK, Mahapatro N, Jena BS. Ageing in fishes. *Gerontology.* 1994;40:113-32.
36. Mikhailova MV, Belyaeva NF, Kozlova NI, Zolotarev KV, Mikhailov AN, Berman AE, et al. Protective action of fish muscle extracts against cellular senescence induced by oxidative stress. *Biomed Khim.* 2017;63:351-5.
37. Hua X, Cowman P, Warren D, Bromham L. Longevity Is Linked to Mitochondrial Mutation Rates in Rockfish: A Test Using Poisson Regression. *Mol Biol Evol.* 2015;32:2633-45.
38. Cressey D. Sharks can live a lot longer than researchers realized. *Nature.* 2017;549:316-7.
39. Nielsen J, Hedeholm R, Heinemeier J, Bushnell PG, Christiansen JS, Olsen J, et al. Eye lens radiocarbon reveals centuries of longevity in the Greenland shark (*Somniosus microcephalus*). *Science.* 2016;353:702-4.
40. George JC, Bada J, Zeh J, Scott L, Brown SE, O'Hara T, et al. Age and growth estimates of bowhead whales (*Balaena mysticetus*) via aspartic acid racemization. *Can J Zool.* 1999;77:571-80.
41. Blier PU, Abele D, Munro D, Degletagne C, Rodriguez E, Hagen T. What modulates animal longevity? Fast and slow aging in bivalves as a model for the study of lifespan. *Semin Cell Dev Biol.* 2017;70:130-40.
42. Gruber H, Schaible R, Ridgway ID, Chow TT, Held C, Philipp EE. Telomere-independent ageing in the longest-lived non-colonial animal, *Arctica islandica*. *Exp Gerontol.* 2014;51:38-45.
43. Munro D, Blier PU. Age, diet, and season do not affect longevity-related differences in peroxidation index between *Spisula solidissima* and *Arctica islandica*. *J Gerontol A Biol Sci Med Sci* 2015;70:434-43.
44. Ungvari Z, Sosnowska D, Mason JB, Gruber H, Lee SW, Schwartz TS, et al. Resistance to genotoxic stresses in *Arctica islandica*, the longest living noncolonial animal: is extreme longevity associated with a multistress resistance phenotype? *J Gerontol A Biol Sci Med Sci* 2013;68:521-9.
45. Sosnowska D, Richardson C, Sonntag WE, Csiszar A, Ungvari Z, Ridgway I. A heart that beats for 500 years: age-related changes in cardiac proteasome activity, oxidative protein damage and expression of heat shock proteins, inflammatory factors, and mitochondrial complexes in *Arctica islandica*, the longest-living noncolonial animal. *J Gerontol A Biol Sci Med Sci.* 2014;69:1448-61.
46. Treaster SB, Ridgway ID, Richardson CA, Gaspar MB, Chaudhuri AR, Austad SN. Superior proteome stability in the longest lived animal. *Age (Dordr).* 2014;36:9597.
47. Lazaro I, Oguiza A, Recio C, Lopez-Sanz L, Bernal S, Egido J, et al. Interplay between HSP90 and Nrf2 pathways in diabetes-associated atherosclerosis. *Clin Investig Arterioscler.* 2017;29:51-9.
48. Onken B, Driscoll M. Metformin induces a dietary restriction-like state and the oxidative stress response to extend *C. elegans* Healthspan via AMPK, LKB1, and SKN-1. *PLoSOne.* 2010;5:e8758.

49. Pérez VI, Bokov A, Van Remmen H, Mele J, Ran Q, Ikeno Y, et al. Is the oxidative stress theory of aging dead? *Biochim Biophys Acta* 2009;1790:1005-14.
50. Gruber H, Wessels W, Bonyton P, Xu J, Wohlgemuth S, Leeuwenburgh C, et al. Age-related cellular changes in the long-lived bivalve *A. islandica*. *Age (Dordr)*. 2015;37:90.
51. Costantini D, Smith S, Killen SS, Nielsen J, Steffensen JF. The Greenland shark: A new challenge for the oxidative stress theory of ageing? *Comp Biochem Physiol A Mol Integr Physiol* 2017;203:227-32.
52. Moyers CD. Controlling muscle mitochondrial content. *J Exp Biol*. 2003;206:4385-91.
53. Dumesic PA, Egan DF, Gut P, Tran MT, Parisi A, Chatterjee N, et al. An Evolutionarily Conserved uORF Regulates PGC1 α and Oxidative Metabolism in Mice, Flies, and Bluefin Tuna. *Cell Metab* 2019;30:190-200.
54. Habiballa L, Salmonowicz H, Passos JF. Mitochondria and cellular senescence: Implications for musculoskeletal ageing. *Free Radic Biol Med*. 2019;132:3-10.
55. Kasai S, Yamazaki H, Tanji K, Engler MJ, Matsumiya T, Itoh K. Role of the ISR-ATF4 pathway and its cross talk with Nrf2 in mitochondrial quality control. *J Clin Biochem Nutr*. 2019;64:1-12.
56. Wang H, Pan L, Si L, Miao J. The role of Nrf2-Keap1 signaling pathway in the antioxidant defense response induced by PAHs in the calm *Ruditapes philippinarum*. *Fish Shellfish Immunol*. 2018;80:325-34.
57. Maher J, Yamamoto M. The rise of antioxidant signaling - the evolution and hormetic actions of Nrf2. *Toxicol Appl Pharmacol* 2010;244:4-15.
58. Galtier N, Blier PU, Nabholz B. Inverse relationship between longevity and evolutionary rate of mitochondrial proteins in mammals and birds. *Mitochondrion*. 2009;9:51-7.
59. Delhaye J, Salamin N, Roulin A, Criscuolo F, Bize P, Christe P. Interspecific correlation between red blood cell mitochondrial ROS production, cardiolipin content and longevity in birds. *Age (Dordr)*. 2016;38:433-43.
60. Trifunovic A, Wredenberg A, Falkenberg M, Spelbrink JN, Rovio AT, Bruder CE, et al. Premature ageing in mice expressing defective mitochondrial DNA polymerase. *Nature*. 2004;429:417-23.
61. Yang JN, Seluanov A, Gorbunova V. Mitochondrial inverted repeats strongly correlate with lifespan: mtDNA inversions and aging. *PLoS One*. 2013;8:e73318.
62. Vermulst M, Bielas JH, Kujoth GC, Ladiges WC, Rabinovitch PS, Prolla TA, et al. Mitochondrial point mutations do not limit the natural lifespan of mice. *Nat Genet*. 2006;39:540-3.
63. Ng L, Ng L, van Breugel M, Halliwell B, Gruber J. Mitochondrial DNA Damage Does Not Determine *C. elegans* Lifespan. *Front Genet*. 2019;10:311.
64. Carey JR. Longevity minimalists: life table studies of two species of northern Michigan adult mayflies. *Exp Geront*. 2002;37:567-70.
65. Valenzano DR, Terzibasi E, Genade T, Cattaneo A, Domenici L, Cellierino A. Resveratrol prolongs lifespan and retards the onset of age-related markers in a short-lived vertebrate. *Curr Biol*. 2006;16:296-300.
66. Platzer M, Englert C. *Nothobranchius furzeri*: A Model for Aging Research and More. *Trends Genet*. 2016;32:543-52.
67. Vrtílek M, Žák J, Blažek R, Polačik M, Cellierino A, Reichard M. Limited scope for reproductive senescence in wild populations of a short-lived fish. *Naturwissenschaften*. 2018;105:68.
68. Zupkovitz G, Kabiljo J, Martin D, Laffer S, Schöfer C, Pusch O. Phylogenetic analysis and expression profiling of the Klotho gene family in the short-lived African killifish *Nothobranchius furzeri*. *Dev Genes Evol*. 2018;228:255-65.
69. Baumgart M, Priebe S, Groth M, Hartmann N, Menzel U, Pandolfini L, et al. Longitudinal RNA-Seq Analysis of Vertebrate Aging Identifies Mitochondrial Complex I as a Small-Molecule-Sensitive Modifier of Lifespan. *Cell Syst*. 2016;2:122-32.
70. Shiels P, Davies RW. Ageing and death in neurones In: Davies RW, Morris BJ, editors. *Molecular Biology of the Neuron*. UK: Oxford University Press; 2004. p. 435-64.
71. Smith P, Willemsen D, Popkes M, Metge F, Gandiwa E, Reichard M, et al. Regulation of life span by the gut microbiota in the short-lived African turquoise killifish. *Elife*. 2017;6:e27014.