



Welden, N. A. , Abylkhani, B. and Howarth, L. M. (2018) The effects of trophic transfer and environmental factors on microplastic uptake by plaice, *Pleuronectes platessa*, and spider crab, *Maja squinado*. *Environmental Pollution*, 239, 351 - 358. (doi:[10.1016/j.envpol.2018.03.110](https://doi.org/10.1016/j.envpol.2018.03.110))

This is the author's final accepted version.

There may be differences between this version and the published version. You are advised to consult the publisher's version if you wish to cite from it.

<http://eprints.gla.ac.uk/166743/>

Deposited on: 20 September 2018

Enlighten – Research publications by members of the University of Glasgow
<http://eprints.gla.ac.uk>

1 The effects of trophic transfer and environmental factors on microplastic 2 uptake by Plaice, *Pleuronectes platessa*, and Spider Crab, *Maja Squinado*

3 Natalie Ann Welden^{1*}, Bexultan Abylkhani² and Leigh Michael Howarth³

4 1. School of Biological Sciences, University of Portsmouth, Portsmouth, England

5 2. School of Engineering, Nazarbayev University, 53, Kabanbay batyr ave. Astana, 010000, Kazakhstan

6 3. Sheffield University, Animal and Plant Sciences Western bank, Sheffield, S10 2TN

7 *Corresponding author: natalie.welden@port.ac.uk

8

9 Highlights

- 10 • *M. squinado* (42.5%) and *P. platessa* (50%) sampled from the Celtic Sea Contained microplastic
- 11 • The proportion of contaminated individuals varied between site and species
- 12 • Microplastic abundance in spider crab and plaice was not linked to local fishing intensity
- 13 • Observations of microplastic in ingested sand eels demonstrate ongoing trophic transfer

14

15 Abstract

16 Microplastic pollution is apparent throughout the marine environment from deep ocean sediments to coastal
17 habitats. Most of this is believed to originate on land, although marine activities, such as fishing and shipping, also
18 contribute to the release and redistribution of microplastic. The relative importance of these maritime plastic
19 sources, the manner by which they are distributed in the environment, and their effect on uptake by marine
20 organisms are yet to be fully quantified. In this study, the relative impact of fishing activities on microplastic uptake
21 by demersal fish and crustaceans was explored. Local fishing intensity, proximity to land and mean water velocity
22 are compared to microplastic uptake in plaice, *Pleuronectes platessa*, and spider crab, *Maja squinado*, from the
23 Celtic Sea. Observations were also made of microplastic contamination in ingested sand eels, *Ammodytes tobianus*,
24 to establish a potential route of trophic transfer. This study is the first to identify microplastic contamination in

25 spider crab and to document trophic transfer in the wild. Individuals were sampled from sites of varied fishing
26 intensity in the Celtic Sea, and their stomach contents examined for the presence of microplastic. Contamination
27 was observed in 50% of *P. platessa*, 42.4% of *M. squinado*, and 44.4% of *A. tobianus*. Locations of highest plastic
28 abundance varied between *P. platessa* and *M. squinado*, indicating that different factors influence the uptake of
29 microplastic in these two taxa. No significant link was observed between fishing effort and microplastic abundance;
30 however, proximity to land was linked to increased abundance in *M. squinado* and Observations of whole prey
31 demonstrate ongoing trophic transfer from *A. tobianus* to *P. platessa*. The lack of significant difference in
32 microplastic abundance between predator and prey suggests that microplastic is not retained by *P. platessa*.

33

34 Keywords: fishing; pollution; plastic; particles; food web; sand eel

35

36 Capsule: Observations of microplastic uptake by plaice and spider crab in UK waters reveals ongoing trophic
37 transfer from sand eels and compares the relative importance of fisheries and land based sources

38 Introduction

39 Microplastics (plastic particles measuring below 5mm) can be found throughout the marine environment. From
40 planktonic organisms (Cole et al., 2013; Desforges et al., 2015) to top predators (Alomar and Deudero, 2017),
41 microplastic uptake has been recorded in a variety of marine taxa representing all trophic levels and feeding modes
42 (Cole et al., 2011). Despite the diversity of organisms seen to consume plastic, the route by which it enters the food
43 chain is still uncertain. Whilst studies have shown a number of species are unable to distinguish between
44 microplastics and prey items (Bern, 1990; Hämer et al., 2014) , it is unclear as to whether microplastic uptake is
45 predominantly direct (from sea water or sediment) or indirect (for example from contaminated prey).

46 Both laboratory and field studies have previously been employed to explore the uptake of microplastic (Lusher et
47 al., 2017a). Crustaceans including filtering planktonic copepods and larvae (Cole et al., 2013), isopods (*Idotea*
48 *emarginata*) (Hämer et al., 2014), and decapods (Brennecke et al., 2015; Farrell and Nelson, 2013; Watts et al.,
49 2014) have each been shown to ingest and aggregate microplastics. Observations of ingested particles have
50 indicated that potential for aggregation in the foregut (Welden and Cowie, 2016b) and translocation into tissues
51 (Farrell and Nelson, 2013). Microplastics may also be taken in during ventilation of the gills (Watts et al., 2014). The
52 tendency for crustaceans to take in plastic is supported by a number of observations in wild caught animals. Whilst
53 the number of studies is comparatively low, the uptake of microplastics in wild crustaceans has been seen to be
54 highly heterogeneous, varying with location (Devriese et al., 2015; Welden and Cowie, 2016a). This may be partially
55 due to variation in environmental levels of microplastic, however, retention in *Nephrops norvegicus* has also been
56 linked to size, sex and moult stage (Welden and Cowie, 2016a).

57 Fewer laboratory studies have analysed the uptake of microplastics by fish, however, ingestion has been seen in a
58 number of species (Batel et al., 2016; Lusher et al., 2017b; Mazurais et al., 2015). Wild-caught fish are more widely
59 studied, and microplastic contamination has been reported in species from both benthic and pelagic habitats.
60 Carnivorous pelagic fishes (Foekema et al., 2013; Lusher et al., 2013; Romeo et al., 2015; Rummel et al., 2016),
61 demersal feeders (Lusher et al., 2013; Rummel et al., 2016), and secondary consumers have all been seen to

62 consume microplastic. Some, such as the Japanese anchovy (*Engraulis japonicus*) may represent a trophic link to
63 predatory species (Tanaka and Takada, 2016).

64 The uptake of microplastics has been shown to have negative impacts on both invertebrates and fish. Previous
65 observations of the impact of microplastic uptake have included translocation from the stomach and gills to other
66 tissues (Batel et al., 2016; Farrell and Nelson, 2013) and reduced nutrient uptake resulting from false satiation and
67 nutrient dilution (Welden and Cowie, 2016b). Secondary effects include reduced reproductive success and chemical
68 transfer (Rochman et al., 2013), and histological changes in the intestine (Pedà et al., 2016) and liver (Lu et al.,
69 2016). In reducing the size, health and fecundity of individuals, these impacts can negatively affect the profitability
70 and sustainability fisheries (Froese, 2004; Howarth et al., 2014).

71 Whether or not microplastics are transferred to humans through their food, and the potential health effects of this
72 transfer, has also recently become a key research priority (Galloway, 2015; Rochman et al., 2015; Van
73 Cauwenberghe and Janssen, 2014). Microplastics have been observed in several commercially harvested species of
74 fish and shellfish including cod, haddock, mackerel (Foekema et al., 2013; Murphy et al., 2017), langoustine (Welden
75 and Cowie, 2016a), oysters (Green, 2016), and mussels (Li et al., 2016), as well as other species from fish markets
76 around the world (Miranda and de Carvalho-Souza, 2016; Rochman et al., 2015). If microplastics do generate
77 negative health effects in humans, it is highly likely that these effects will increase in relation with the abundance
78 of microplastic in our food.

79 The uptake of microplastic by wild caught organisms has previously been related to the scale and proximity of
80 sources, local bathymetry and transfer from prey. As the weathering of in-use and abandoned, lost and discarded
81 fishing gear can be a source of microplastics in the marine environment, it has been hypothesised that areas of high
82 fishing activity will demonstrate in locally elevated levels of microplastic contamination. For the reasons above, we
83 investigated levels of microplastic contamination in two commercially important species in the Celtic Sea to test if:
84 (1) these organisms contained traces of microplastics; (2) whether microplastic contamination increased with levels
85 of fishing effort or other environmental drivers; and (3) whether these organisms had become contaminated
86 through trophic transfer. To observe the difference in plastic uptake by invertebrates and fish, this study compares

87 two species of commercial interest; plaice (*Pleuronectes platessa*) and spider crab (*Maja squinado*). Additional
88 observations of microplastics ingested by the sand-eel (*Ammodytes tobianus*) were made to establish the potential
89 importance of this species as a route of trophic transfer.

90 Of the two focal species, *P. platessa* is a predatory demersal flat fish which feeds on worms, molluscs and small
91 crustaceans (Millner et al., 2005). Thanks to their wide distribution, they are targeted by European otter and beam
92 trawlers throughout the North East Atlantic (Dunn and Pawson, 2002). In contrast, *M. squinado* is an omnivorous
93 crustacean which can feed opportunistically on a range of food items including seaweed, detritus, invertebrates
94 and carrion (Bernárdez et al., 2000). Due to their life history, they are only seasonally targeted by potting vessels
95 (González-Gurriarán et al., 2002). Whilst both species routinely feed in benthic habitats and should be exposed to
96 a similar level of environmental microplastics, crabs and lobsters, including *M. squinado*, have a complex filter system
97 in the gut including a hardened gastric mill which could lead to increased microplastic retention (Welden and Cowie,
98 2016a).

99

100 Methods

101 Study Site

102 The Celtic Sea is the region of coastal shelf bordered by the Irish Sea, English Channel and Atlantic Ocean. It reaches
103 depths of up to 200m and contains a combination of sandy and muddy sediments. Fisheries operating in the area
104 target a range of species including gadoids, flatfish and crustaceans. Potential sources of terrestrial plastic include
105 the catchment of the River Severn, and industrial activities around Cardiff, Newport and Bristol. Net transport of
106 this plastic is expected to be offshore, moving east to west.

107 *Pleuronectes platessa* and *M. squinado* were sampled from six sites in the Celtic Sea (Figure 1), chosen for similar
108 benthic substrate and variable fishing intensity. Fishing intensity was evaluated as the swept-area ratio per year.
109 The ratio is calculated as the mean number of times a 1.8 km² square is affected by trawling gear. For this study,
110 the mean swept area was derived from data recorded by Eigaard et al (2016). In addition, the mean horizontal

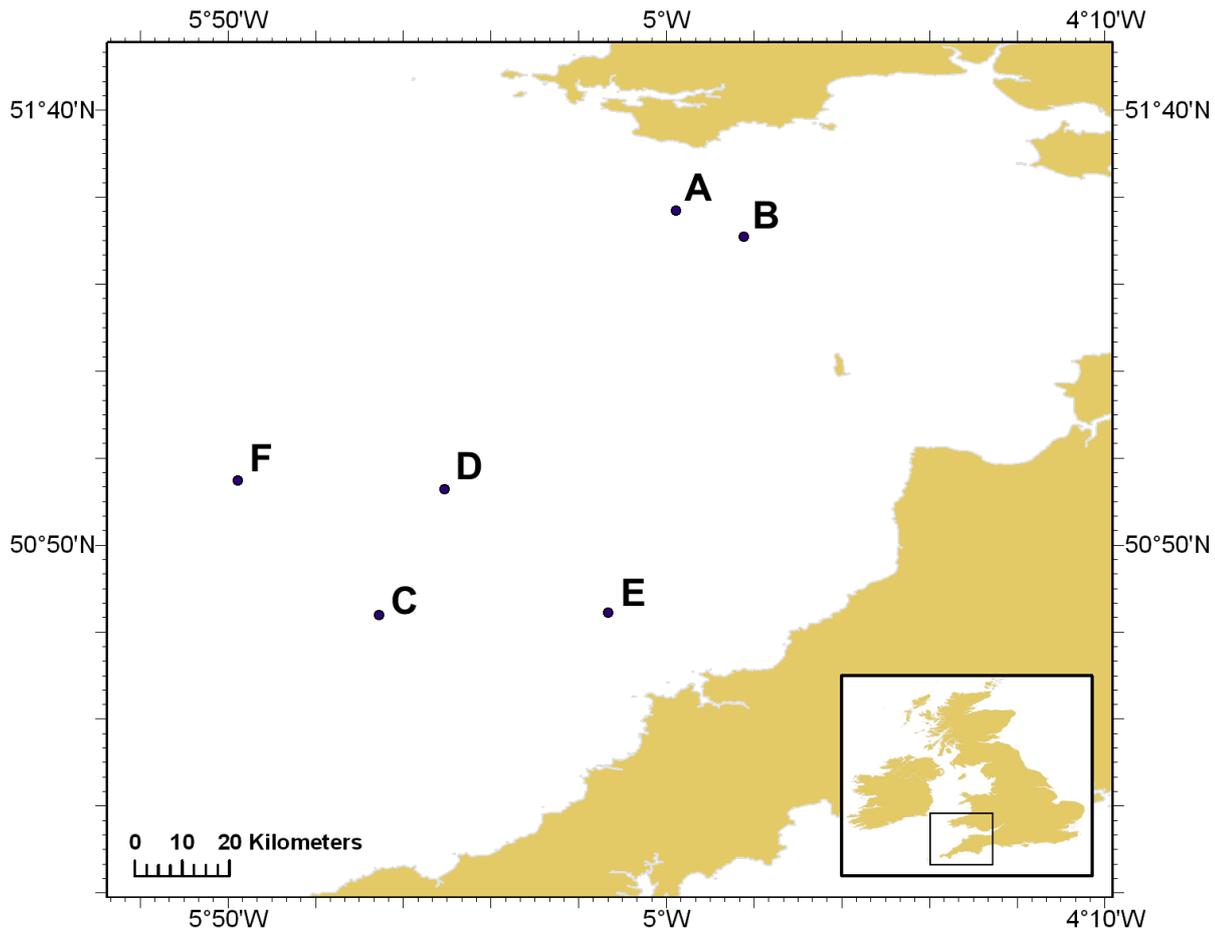
111 velocity (m s^{-1}), distance to land (m) and primary production ($\text{mg C m}^{-2} \text{ yr}^{-1}$) were also calculated from existing GIS
112 layers. The average annual primary production at each site was determined using MODIS satellite sensor data
113 collected between 2009 and 2013 by NEODAAS (www.neodaas.ac.uk), at a resolution of 1.1 km². Daily mean
114 horizontal velocity was extracted from the data derived from the North West Shelf Reanalysis CMEMS
115 (www.marine.copernicus.eu).

116 *Sampling*

117 Both *P. platessa* and *M. squinado* were collected using a 4m beam trawl with a 50mm net, deployed for 30 minutes
118 at each site. Initial dissection was conducted onboard, and the digestive tracts were individually preserved in
119 formalin. Removal of the digestive tract of *M. squinado* was carried out as outlined in Welden and Cowie (2016a);
120 each individual was sexed and measured for carapace length and width, following which their carapace was
121 cracked and the oesophagus and hind gut cut away, keeping the stomach intact until analysis. Stomachs of *P.*
122 *platessa* were removed as described in Lusher et al. (2013); the length and weight of the individual were recorded,
123 and the stomach was excised by separating at the oesophagus and midgut.

124 *Microplastic extraction and analysis*

125 Stomach contents were sorted under dissecting microscope at between x7.5 and x25 magnification. Potential
126 plastics were identified using a combination of by colour, regularity of shape, surface texture, ductility and
127 resistance to breaking. Largest fibres and fragments were removed using forceps, those too small to handle in this
128 manner were transferred to a clean filter paper using a pipette. Identifiable prey remains were recorded and whole
129 *A. tobianus* were individually dissected. Suspected microplastics were transferred to microcentrifuge tubes prior to
130 confirmation of polymer composition. Whilst procedural blanks were not deployed during the initial dissection,
131 petri dishes containing filter papers dampened with de-ionized water were used to determine the potential for
132 contamination by ambient airborne microplastic levels during the analysis.



133

134 *Figure 1. Sampling sites in the Celtic Sea: A, 51° 28' 29.0532" N, 4° 58' 56.1144" W; B, 51° 25' 28.254" N, 4° 51'*
 135 *11.34" W; C, 50° 41' 57.0444" N, 5° 32' 48.8436" W; D, 50° 56' 24.9756" N, 5° 25' 18.5772" W; E, 50° 42' 10.4508"*
 136 *N, 5° 6' 41.2308" W; F, 50° 57' 27.4356" N, 5° 48' 52.8408" W.*

137

138 Fourier Transformed Infrared Spectrometry (FT-IR) was carried out on half of the samples to verify the identity of
 139 the suspected microplastics. Samples were selected for FT-IR analysis using a random number generator. Analysis
 140 was carried out at wavelengths between 800 and 4000 cm^{-1} using a Thermo Nicolet Nexus FT-IR spectrometer with
 141 attached Continuum IR microscope. The abundance of the remaining suspected plastics was rounded down by the
 142 number of miss-identified samples.

143

144 *Statistical analysis*

145 Statistical analysis of the results was carried out as outlined in Welden and Cowie (2016a). Data were analysed using
146 R Studio version 1.0.44. Prior to analysis, the mean number and deviation of microplastics per individual was
147 calculated. A Kolmogorov-Smirnov analysis was used to assess the distribution of the data for each species, after
148 which generalised linear models (GLM) were used to examine the variation in microplastic in the focal species. In the
149 first model, microplastic uptake in *P. platessa* was examined in relation to fishing intensity, primary production,
150 mean velocity, distance from land, and fish weight. A second GLM compared microplastic uptake in *M. squinado* in
151 relation to fishing intensity, primary production, mean velocity, and distance from land, as well as to the carapace
152 length and sex of the individual. To produce the best model for plastic abundance in analysis with intercorrelated
153 independent variables were examined in separate GLMs, after which stepwise reduction and lowest AIC were used
154 to select the most relevant result.

155

156 **Results**

157 In total 140 potential microplastics were recovered, 76 of which were successfully analysed using FT-IR. Of these
158 12 (15.7%) were found to be misidentified. It was hoped that more than 50% of the sample would be analysed,
159 however, the model of FTIR microscope available proved unable to provide consistent readings for smallest
160 samples. The minimum reliable size varied in relation to the dimensions of the sample, for example fragments
161 produced more consistent results at low dimensions than thin fibres. The level of plastics observed in the procedural
162 blanks averaged less than 2 MP per petri dish per hour.

163 Gut content analysis revealed microplastic contamination in 50% of *P. platessa* and 42.5% *M. squinado* (Table 1).
164 Microplastic was identified in *P. platessa* recovered from all sites at which they were sampled, whereas *M. squinado*
165 from site C did not contain microplastic. *P. platessa* exhibited the highest rates of microplastic contamination at
166 site B, whilst *M. squinado* exhibited highest rates of microplastic contamination at A and B, with 73% and 67%
167 occurrence respectively (Figure 2). The shells of *M. squinado* were also covered with plastic fibres similar to those

168 from fishing nets and trawl chafers (Figure 3). In both species, the most frequently ingested plastics were fibres,
 169 and when subjected to micro-FTIR analysis, the most commonly identified plastics were polypropylene, polyester,
 170 polyamide. The variation in the proportion of microplastic contaminated individuals at each site was highest in *M.*
 171 *squinado*, whilst the variation in microplastic uptake per individual at each site was highest in *P. platessa*.

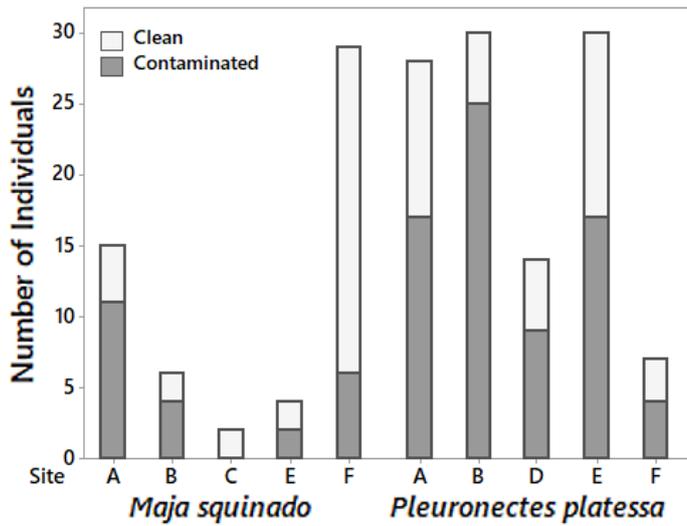
172 Identifiable prey remains were similar in both focal species. Observed taxa were predominantly crustacean, small
 173 bivalves and polychaete worms. In addition to invertebrate prey, nine intact sand eels, *A. tobianus*, were recovered
 174 the foreguts of *P. platessa*, with up to seven individuals recorded in the stomach of a single *P. platessa*. Dissection
 175 and analysis of *A. tobianus* revealed that 44.4% percent contained microplastic particles (Table 1). A Mann-Whitney
 176 test was used to analyse the variation in microplastic abundance between *P. platessa* and their *A. tobianus* prey,
 177 however, no significant difference was identified ($W = 6523, P < 0.6835$). Prey commonly identified in the stomachs
 178 of *A. tobianus* were crustacean larvae and copepods.

179

180 *Table 1. The abundance of microplastic recovered in Pleuronectes platessa, Maja squinado, and Ammodytes*
 181 *tobianus*

Species	N	Number of contaminated individuals	Number of microplastics	Mean number of microplastics per animal (SD)
<i>Pleuronectes platessa</i>	109	54	79	1.46 (1.02)
<i>Maja squinado</i>	54	23	32	1.39 (0.79)
<i>Ammodytes tobianus</i>	9	4	7	1.75 (0.83)

182



183

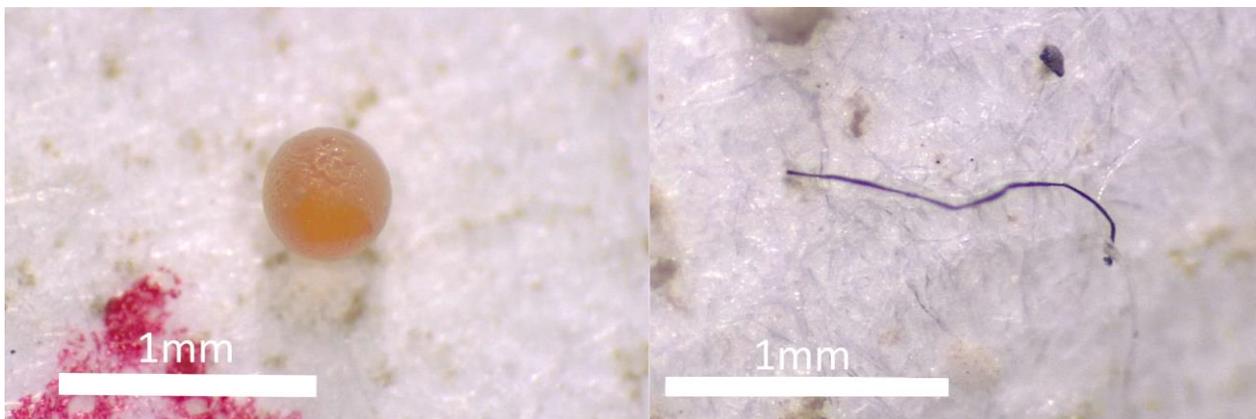
184 Figure 2. Plastic uptake by *Pleuronectes platessa* and *Maja squinado* at each sampling site

185



186

187 Figure 3. Macroplastic fibres adhered to the carapace of *Maja squinado*



188

189 Figure 4. Microplastic pellet and fibre recovered from *M. squinado* (site A and F respectively)

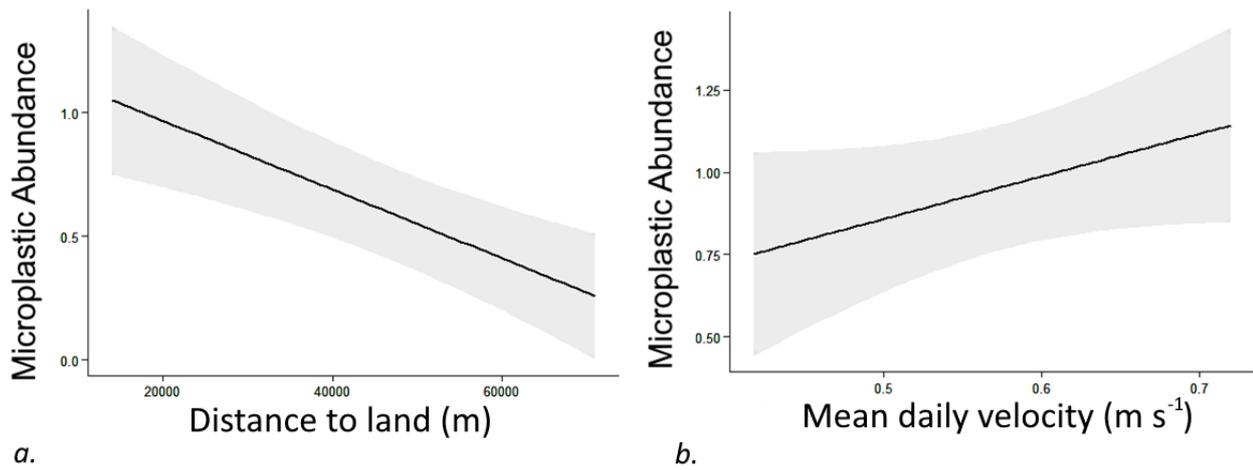
190 GLM analysis of the factors significantly correlated to the uptake of microplastic by *P. platessa* and *M. squinado*
 191 revealed differing relationships between the two species (Table 2). In *P. platessa*, the abundance of microplastic
 192 was positively correlated with weight and daily mean velocity (Figure 5), and negatively correlated with primary
 193 production (Table 2). Analysis of the factors responsible for microplastic uptake in *M. squinado* revealed that only
 194 distance to land influenced the level of contamination, with animals nearshore found to contain microplastic at
 195 higher abundances (Figure 5). Notably, fishing intensity was not found to be significant in either of the analyses.

196

197 Table 2. Observed relationships between microplastic abundance and dependent variables

<i>P. platessa</i>					<i>M. squinado</i>				
	Estimate	St. Error	T - value	P		Estimate	St. Error	T - value	P
Intercept	1.037e+00	4.951e-01	2.094	0.038648	Intercept	1.243e+00	1.945e-01	6.389	4.65e-08
Distance from land	1.235e-05	6.953e-06	1.777	0.078537	Distance from land	-1.385e-05	3.608e-06	-3.840	0.000336
Mean daily velocity	2.984e+00	1.255e+00	2.378	0.019236					
Primary Production	-3.754e-03	1.062e-03	-3.536	0.000608					
Weight	5.249e-03	1.214e-03	4.324	3.52e-05					

198



199

200 Figure 5. The statistical relationship between microplastic abundance and distance from land in *M. squinado* (a) and
 201 microplastic abundance and mean daily velocity in *P. platessa* (b).

202

203 Discussion

204 Microplastics were recorded at all sample sites; however, there was variation in the proportion of contaminated
205 individuals. It is worth noting that mechanical sorting of samples limited the minimum size of plastic selected to
206 approximately 500µm. As a result the smallest microplastic fractions may be under represented.

207 The carapaces of many *M. squinado* were covered in fibres, presumably as a result of contact with fishing nets or
208 accidental addition as masking material (Parapar et al., 1997). However, high external contamination of
209 macroplastic did not correspond to increased levels of microplastic ingestion; the maximum number of
210 microplastics recovered per individual was 3, with an average of 1.39 items per individual. The proportion of *M.*
211 *squindao* seen to contain microplastic and the abundance of microplastics recovered falls between that observed
212 in similarly sized wild-caught crustaceans, *N. norvegicus*, from remote fishing grounds off the coast of North
213 Scotland and those observed in the highly impacted Clyde Sea Area (Welden and Cowie, 2016a). The level of spatial
214 heterogeneity recorded in *M. squinado* and the low level of microplastics per individual is also similar to that of the
215 brown shrimp, *Crangon crangon*, from the southern North Sea and Channel (1.23 ± 0.99 microplastics per
216 individual) (Devriese et al., 2015).

217 Analysis of the occurrence of microplastic in *P. platessa* revealed contamination in 50% of the animals sampled;
218 however, previous studies have revealed levels of microplastic contamination far lower than those recorded here.
219 The degree of uptake varies greatly, for example, 5.5% in Rummel et al. (2016), 11% in Lusher et al. (2015), 29% by
220 Murphy et al. (2017), and 35% in Boerger et al. (2010). Many of the lowest occurrences of microplastic have been
221 recorded in fish caught in offshore and mid-water trawls. Due to such extreme variation between locations and
222 target species, comparisons between the results presented above and existing studies have been limited to those
223 that examine nearshore environments and similar demersal feeders.

224 The percentage of contaminated *P. platessa* recorded here is commensurate with other studies of microplastic
225 uptake by fish in UK waters. Whilst the abundance of fish seen to have consumed plastic is higher than that of

226 demersal species reported from the nearby English Channel (35%) (Lusher et al., 2013), it is similar to that observed
227 in fish in nearshore Scottish waters (45%) (Murphy et al., 2017), and below that of flounder, *Platichthys flesus*, in
228 the Thames Estuary (McGoran et al., 2017). The average number of microplastics per animal (1.46) was also similar
229 to that seen around the UK; higher than that recorded by Murphy et al (2017) (0.9 ± 1.79), but similar to that
230 observed in demersal fish studied by Lusher et al. (2013) (1.2 ± 0.54).

231 The difference in the level of microplastic uptake and retention in *M. squinado* and *P. platessa* is not unexpected
232 due to the distinct morphology and feeding modes of the two taxa. Indeed, similar variation between species with
233 different feeding modes has been observed in species found at >2200m in the Rockall Trough (Courtene-Jones et
234 al., 2017). In addition to feeding mode, age or body mass may also affect the dimensions and number of
235 microplastics that are ingested and retained. For example, body mass has previously been linked to lower
236 microplastic uptake in *N. norvegicus* (Welden and Cowie, 2016a); however, this is driven by the presence of the
237 gastric mill, a structure not found in fish. In our studied species, microplastic uptake may be affected by numerous
238 factors linked to body size. In *P. platessa*, the relationship between length and stomach volume is a linear one, and
239 it is predicted that a greater weight of food is consumed by larger individuals (Jobling, 1980). Increased food
240 consumption in larger fish may result in the higher microplastic uptake seen in the statistical analysis. In *M. squinado*
241 dietary composition may vary with age (Bernárdez et al., 2000), altering the level of microplastic to which individuals
242 of different ages are exposed, however, in this study there was no significant link between microplastic abundance
243 and carapace size.

244 Variation in plastic retention may also be related to rates of gastric evacuation or regurgitation. In *N. norvegicus*,
245 plastics have been evacuated with the stomach lining at ecdysis (Welden and Cowie, 2016a). Many spider crabs,
246 including *M. squinado*, do not moult after reaching sexual maturity (González-Gurriarán et al., 1995). Species which
247 no longer undergo ecdysis must rely on microplastics being sufficiently small or appropriately oriented to pass
248 through the gastric mill. The period between pre-pubertal moults in *M. squinado* is also highly variable (Corgos et
249 al., 2007), the regularity with which juveniles may expel any retained plastics will have a further impact on variation
250 in microplastic contamination in this species.

251 The apparent differences in microplastic uptake between the two focal species indicate that observations of high
252 microplastic abundance in one species cannot be used to infer high contamination throughout a community. Whilst
253 this variation between taxa is expected, the significance of these results will be determined by whether the
254 relationship between the microplastic uptake rates of the two species remains consistent across a range of
255 environmental microplastic concentrations. Consistent relationships between species may allow extrapolation of
256 contamination between species, reducing the level of potentially damaging sampling required.

257

258 *The Effect of Site Specific Factors and Local Fishing Intensity on Microplastic Uptake*

259 The abundance of recovered microplastic in both *M. squinado* and *P. platessa* varied between the six sampled sites;
260 however no common factors influencing microplastic abundance were found between the two species. In addition
261 to the different factors affecting microplastic abundance, *M. squinado* exhibited higher variation between locations.

262 The composition of recovered polymers indicates a probable combination of land based and fisheries sourced
263 plastics. Local fishing pressure may affect microplastic availability and uptake in a number of ways, either by
264 introducing microplastic via the weathering of fishing gear or by the resuspension of deposited plastics during the
265 disturbance of seabed sediments (Churchill, 1989). However, statistical analysis indicated no significant links
266 between the abundance of microplastics and fishing intensity. In addition to being a potential source of
267 microplastics, trawling results in the resuspension of sediment. Plumes of sediment and microplastics are re-
268 distributed as a result of the tidal state, circulation and wind patterns apparent during the resuspension event
269 (Floderus and Pihl, 1990). As a result, environmental factors may mediate and diffuse microplastic inputs from
270 trawling.

271 It is probable that microplastic uptake by benthic and demersal species in coastal environments is driven by
272 proximity to shore and land based sources. This is supported by the significantly higher levels of microplastics in *M.*
273 *squinado* at nearshore sites. This is particularly apparent at site A and B and is also visible in *P. platessa* at site B,

274 the two nearshore sites. Similar observations have been made in *N. norvegicus* in Scottish coastal waters, in which
275 microplastic abundance was highest in the nearshore site (Welden and Cowie, 2016a).

276 In *P. platessa*, microplastic abundance was positively correlated with the mean daily water velocity and primary
277 production. The reduction in microplastic abundance per individual at sites of high primary production may be the
278 result of local increases in population density. At sites of high primary production there may be increases in the
279 abundance of primary consumers and predatory species (Frederiksen et al., 2006). Assuming a similar level of
280 microplastic between locations, the higher density of feeding animals may result in fewer microplastics available to
281 an individual. However, as we are not able to definitively state the level of microplastic in the water column a great
282 deal of additional observation is needed for this explanation to be accepted.

283 The higher average microplastic abundance in *P. platessa* has also been linked to areas of increased water velocity.
284 Faster currents may result in the refloatation and reduced deposition of microplastic. For example, it is known that
285 increased water movement during storm events results in higher concentrations of suspended plastic (Lattin et al.,
286 2004), and periods of elevated wave activity have been linked to increases in the mean abundance and mean size
287 of microplastics (Reisser et al., 2015). Elevated levels of microplastic at the water-sediment interface may be the
288 source of increased microplastic uptake in these fish.

289

290 *Trophic Transfer to P. platessa*

291 Observation of microplastic in the stomach content of ingested *A. tobianus* indicates an active route of trophic
292 transfer to *P. platessa*. *A. tobianus* are predominantly plankton feeders, and microplastic may be taken up from
293 seawater or from copepods and other zooplankton, a group known to ingest microplastic (Desforges et al., 2015).
294 *Ammodytes* sp. are a key prey species for many piscivorous organisms (Frederiksen et al., 2006; O'Connell and Fives,
295 1995). Although the number of individuals sampled in this study is small, there is clear evidence that predatory fish,
296 seabirds and marine mammals are at risk of microplastic uptake via trophic transfer (Furness, 2002; Rindorf et al.,
297 2000). A similar observation has been made in captive common seals, *Halichoerus grypus*, fed wild caught mackerel,

298 *Scomber scombrus*; in which microplastics were found in 32% of the fish analysed and 48% of seal scat subsamples
299 (Nelms et al., 2018).

300 In addition the presence of *Ammodytes* sp., previous dietary observations of *P. platessa* have revealed polychaetes
301 such as *Pectinaria* and *Nereis*, bivalves including *Ensis* and *Spisula*, and crustaceans including *Upoebia* and
302 *Macropipus* (Rijnsdorp and Vingerhoed, 2001). Their diet is temporally variable; the relative importance of
303 polychaete prey is seasonal, with the weight recorded increasing to over 60% of the stomach contents in summer
304 months. Similarly, larger individuals have a reduced dependence on annelids, increasing their consumption of
305 bivalves, echinoderms and vertebrate prey (Rijnsdorp and Vingerhoed, 2001). Polychaetes, such as *Nereis*
306 (Lourenço et al., 2017) and *Arenicola* (Van Cauwenberghe et al., 2015) have also been seen to take in plastics in the
307 wild, and may represent a further route of transfer to plaice; however, this could not be confirmed in the current
308 analysis due to the highly degraded state of soft bodied prey.

309 Mann-Whitney analysis comparing the abundance of plastic in *A. tobianus* and *P. platessa* species revealed no
310 significant difference in plastic loads between the trophic levels. This indicates that whilst *P. platessa* may consume
311 multiple individuals containing microplastic, most will be readily egested and not retained in the gut. A positive
312 relationship was observed between microplastic abundance and individual weight in *P. platessa*. As indicated in
313 the previous section, this may be the result of increased feeding rates by larger individuals, with a greater number
314 of contaminated prey items being consumed in a shorter time period.

315

316 *Impacts of microplastic ingestion on Maja squinado and Pueronectes platessa*

317 Ingestion of microplastics may have a range of effects on the health of our focal species. Retention of plastics by
318 crustaceans has been seen to result in aggregation in the gut, reduced feeding and lower nutritional state (Blarer
319 and Burkhardt-Holm, 2016; Watts et al., 2015; Welden and Cowie, 2016b). Microplastics at the lower end of the
320 size range may also translocate into the tissues (Brennecke et al., 2015; Farrell and Nelson, 2013), resulting in a
321 range of physiological effects such as reduced mobility and survivorship (Tosetto et al., 2016). However, these

322 effects may not be apparent in animals which do not contain large aggregations of microplastic or are not exposed
323 for extended periods (Hämer et al., 2014), as observed in *Echinogammarus marinus* (Bruck and Ford, 2018) and *Uca*
324 *rapax* (Imhof and Laforsch, 2016).

325 In fish, microplastic ingestion has been linked to translocation (Lu et al., 2016), reduced predatory performance and
326 feeding efficiency in *Pomatoschistus microps* (de Sá et al., 2015), changes in the histology and lipid uptake in the
327 liver of *Danio rerio* (Lu et al., 2016), and altered histology and function in the intestine of *Dicentrarchus labrax*
328 (Pedà et al., 2016). As in crustaceans, the potential of microplastics to negatively affect an organism may be
329 dependent on the degree of aggregation and retention time. Analysis of microplastic consumption in goldfish has
330 suggested that particles over 63µm are not held in the gut for extended periods (Grigorakis et al., 2017). Low
331 retention time may be responsible for lack of plastic contamination observed in a number of species; for example,
332 eelpout, *Zoarces viviparus*, sampled from the Baltic Sea and North Sea, which were not seen to contain plastic
333 (Wesch et al., 2016).

334 In addition to the potential direct impacts of microplastic uptake on *P. platessa*, there may be indirect effects
335 related to contamination of prey species. *A. tobianus* that have ingested microplastic may have lower feeding rates
336 and reduced nutrient assimilation similar to those outlined in the species above. As a result, microplastic
337 contaminated prey may have lower nutritional value. Regularly consuming prey of lower quality would reduce the
338 foraging efficiency of *P. platessa* and other predators, requiring individuals to spend a greater time foraging and
339 feeding. As more information is generated describing the energetic and nutritional costs of microplastic uptake, so
340 models must be developed to project these effects through the trophic levels.

341

342 *Conclusions*

343 Crustaceans and fish from the Celtic Sea were both seen take in plastic at levels commensurate with other studies
344 of coastal waters around the UK; however, the pattern of microplastic uptake varied between the two taxa. Fishing
345 intensity and the associated microfibrils released from trawl nets did not significantly raise the level of microplastic

346 in either species. Instead, proximity to shore resulted in a greater contamination in *M. squinado* and body weight,
347 mean water velocity and increased primary production were linked to variation in microplastic abundance in *P.*
348 *platessa*. This study is the first to confirm the trophic transfer of plastics in progress in the marine environment,
349 raising concerns over the relative nutritional value of prey and the effect on the foraging efficiency of *P. platessa*.
350 In addition to highlighting the traditional issue of impacts on the contaminated organism, researchers must now
351 consider the impacts of microplastic uptake on the commercial value of economically important fish and shellfish
352 species.

353

354 **Acknowledgements**

355 Analysis of samples was carried out with the assistance of a grant from the Fisheries Society of the British Isles.

356

357 Alomar, C., Deudero, S., 2017. Evidence of microplastic ingestion in the shark *Galeus melastomus* Rafinesque,
358 1810 in the continental shelf off the western Mediterranean Sea. *Environmental Pollution* 223, 223-229.

359

360 Batel, A., Linti, F., Scherer, M., Erdinger, L., Braunbeck, T., 2016. Transfer of benzo [a] pyrene from microplastics to
361 *Artemia nauplii* and further to zebrafish via a trophic food web experiment: CYP1A induction and visual tracking of
362 persistent organic pollutants. *Environmental toxicology and chemistry* 35, 1656-1666.

363

364 Bern, L., 1990. Size-related discrimination of nutritive and inert particles by freshwater zooplankton. *Journal of*
365 *Plankton Research* 12, 1059-1067.

366

367 Bernárdez, C., Freire, J., González-Gurriarán, E., 2000. Feeding of the spider crab *Maja squinado* in rocky subtidal
368 areas of the Ría de Arousa (north-west Spain). *Journal of the Marine Biological Association of the United Kingdom*
369 80, 95-102.

370

371 Blarer, P., Burkhardt-Holm, P., 2016. Microplastics affect assimilation efficiency in the freshwater amphipod
372 *Gammarus fossarum*. *Environmental Science and Pollution Research* 23, 23522-23532.

373

374 Boerger, C.M., Lattin, G.L., Moore, S.L., Moore, C.J., 2010. Plastic ingestion by planktivorous fishes in the North
375 Pacific Central Gyre. *Marine Pollution Bulletin* 60, 2275-2278.

376

377 Brennecke, D., Ferreira, E.C., Costa, T.M., Appel, D., da Gama, B.A., Lenz, M., 2015. Ingested microplastics (>
378 100µm) are translocated to organs of the tropical fiddler crab *Uca rapax*. *Marine pollution bulletin* 96, 491-495.

379

380 Bruck, S., Ford, A., 2018. Chronic ingestion of polystyrene microparticles in low doses has no effect on food
381 consumption and growth to the intertidal amphipod *Echinogammarus marinus*? *Environmental Pollution* 233,
382 1125-1130.

383

384 Churchill, J.H., 1989. The effect of commercial trawling on sediment resuspension and transport over the Middle
385 Atlantic Bight continental shelf. *Continental Shelf Research* 9, 841-865.

386

387 Cole, M., Lindeque, P., Fileman, E., Halsband, C., Goodhead, R., Moger, J., Galloway, T.S., 2013. Microplastic
388 Ingestion by Zooplankton. *Environmental Science & Technology* 47, 6646-6655.

389

390 Cole, M., Lindeque, P., Halsband, C., Galloway, T.S., 2011. Microplastics as contaminants in the marine
391 environment: A review. *Marine Pollution Bulletin* In Press.

392

393 Corgos, A., Sampedro, M.P., González-Gurriarán, E., Freire, J., 2007. Growth at Moulting, Intermoult Period, and
394 Moulting Seasonality of the Spider Crab *Maja Brachydactyla*: Combining Information from Mark-Recapture and
395 Experimental Studies. *Journal of Crustacean Biology* 27, 255-262.

396

397 Courtene-Jones, W., Quinn, B., Gary, S.F., Mogg, A.O.M., Narayanaswamy, B.E., 2017. Microplastic pollution
398 identified in deep-sea water and ingested by benthic invertebrates in the Rockall Trough, North Atlantic Ocean.
399 *Environmental Pollution* 231, 271-280.

400

401 de Sá, L.C., Luís, L.G., Guilhermino, L., 2015. Effects of microplastics on juveniles of the common goby
402 (*Pomatoschistus microps*): Confusion with prey, reduction of the predatory performance and efficiency, and
403 possible influence of developmental conditions. *Environmental Pollution* 196, 359-362.

404

405 Desforges, J.-P.W., Galbraith, M., Ross, P.S., 2015. Ingestion of microplastics by zooplankton in the Northeast
406 Pacific Ocean. *Archives of environmental contamination and toxicology* 69, 320-330.

407

408 Devriese, L.I., van der Meulen, M.D., Maes, T., Bekaert, K., Paul-Pont, I., Frère, L., Robbens, J., Vethaak, A.D., 2015.
409 Microplastic contamination in brown shrimp (*Crangon crangon*, Linnaeus 1758) from coastal waters of the
410 Southern North Sea and Channel area. *Marine Pollution Bulletin* 98, 179-187.

411

412 Dunn, M., Pawson, M., 2002. The stock structure and migrations of plaice populations on the west coast of
413 England and Wales. *Journal of Fish Biology* 61, 360-393.

414

415 Eigaard, O.R., Bastardie, F., Hintzen, N.T., Buhl-Mortensen, L., Buhl-Mortensen, P., Catarino, R., Dinesen, G.E.,
416 Egekvist, J., Fock, H.O., Geitner, K., 2016. The footprint of bottom trawling in European waters: distribution,
417 intensity, and seabed integrity. *ICES Journal of Marine Science* 74, 847-865.

418

419 Farrell, P., Nelson, K., 2013. Trophic level transfer of microplastic: *Mytilus edulis* (L.) to *Carcinus maenas* (L.).
420 *Environmental Pollution* 177, 1-3.

421

422 Floderus, S., Pihl, L., 1990. Resuspension in the Kattegat: impact of variation in wind climate and fishery.
423 Estuarine, Coastal and Shelf Science 31, 487-498.

424

425 Foekema, E.M., De Gruijter, C., Mergia, M.T., van Franeker, J.A., Murk, A.J., Koelmans, A.A., 2013. Plastic in North
426 Sea Fish. Environmental Science & Technology 47, 8818-8824.

427

428 Frederiksen, M., Edwards, M., Richardson, A.J., Halliday, N.C., Wanless, S., 2006. From plankton to top predators:
429 bottom-up control of a marine food web across four trophic levels. Journal of Animal Ecology 75, 1259-1268.

430

431 Froese, R., 2004. Keep it simple: three indicators to deal with overfishing. Fish and fisheries 5, 86-91.

432

433 Furness, R.W., 2002. Management implications of interactions between fisheries and sandeel-dependent seabirds
434 and seals in the North Sea. ICES Journal of Marine Science 59, 261-269.

435

436 Galloway, T.S., 2015. Micro-and nano-plastics and human health, Marine anthropogenic litter. Springer, pp. 343-
437 366.

438

439 González-Gurriarán, E., Freire, J., Bernárdez, C., 2002. Migratory patterns of female spider crabs *Maja squinado*
440 detected using electronic tags and telemetry. Journal of Crustacean Biology 22, 91-97.

441

442 González-Gurriarán, E., Freire, J., Parapar, J., Sampedro, M.P., Urcera, M., 1995. Growth at moult and moulting
443 seasonality of the spider crab, *Maja squinado* (Herbst) (Decapoda: Majidae) in experimental conditions:
444 implications for juvenile life history. Journal of Experimental Marine Biology and Ecology 189, 183-203.

445

446 Green, D.S., 2016. Effects of microplastics on European flat oysters, *Ostrea edulis* and their associated benthic
447 communities. Environmental Pollution 216, 95-103.

448

449 Grigorakis, S., Mason, S.A., Drouillard, K.G., 2017. Determination of the gut retention of plastic microbeads and
450 microfibers in goldfish (*Carassius auratus*). Chemosphere 169, 233-238.

451

452 Hämer, J., Gutow, L., Köhler, A., Saborowski, R., 2014. Fate of Microplastics in the Marine Isopod *Idotea*
453 *emarginata*. Environmental Science & Technology 48, 13451-13458.

454

455 Howarth, L.M., Roberts, C.M., Thurstan, R.H., Stewart, B.D., 2014. The unintended consequences of simplifying
456 the sea: making the case for complexity. Fish and Fisheries 15, 690-711.

457

458 Imhof, H.K., Laforsch, C., 2016. Hazardous or not – Are adult and juvenile individuals of *Potamopyrgus*
459 *antipodarum* affected by non-buoyant microplastic particles? Environmental Pollution 218, 383-391.

460

461 Jobling, M., 1980. Gastric evacuation in plaice, *Pleuronectes platessa* L.: effects of temperature and fish size.
462 Journal of Fish Biology 17, 547-551.

463

464 Lattin, G.L., Moore, C.J., Zellers, A.F., Moore, S.L., Weisberg, S.B., 2004. A comparison of neustonic plastic and
465 zooplankton at different depths near the southern California shore. *Marine Pollution Bulletin* 49, 291-294.

466

467 Li, J., Qu, X., Su, L., Zhang, W., Yang, D., Kolandhasamy, P., Li, D., Shi, H., 2016. Microplastics in mussels along the
468 coastal waters of China. *Environmental Pollution* 214, 177-184.

469

470 Lourenço, P.M., Serra-Gonçalves, C., Ferreira, J.L., Catry, T., Granadeiro, J.P., 2017. Plastic and other microfibers in
471 sediments, macroinvertebrates and shorebirds from three intertidal wetlands of southern Europe and west Africa.
472 *Environmental Pollution* 231, 123-133.

473

474 Lu, Y., Zhang, Y., Deng, Y., Jiang, W., Zhao, Y., Geng, J., Ding, L., Ren, H., 2016. Uptake and accumulation of
475 polystyrene microplastics in zebrafish (*Danio rerio*) and toxic effects in liver. *Environmental science & technology*
476 50, 4054-4060.

477

478 Lusher, A., McHugh, M., Thompson, R., 2013. Occurrence of microplastics in the gastrointestinal tract of pelagic
479 and demersal fish from the English Channel. *Marine pollution bulletin* 67, 94-99.

480

481 Lusher, A., Welden, N., Cole, M., 2017a. Critical Review: Sampling, isolating and identifying microplastics ingested
482 by fish and invertebrates. *Anal Methods*. DOI 10, c6ay02415g.

483

484 Lusher, A., Welden, N., Sobral, P., Cole, M., 2017b. Sampling, isolating and identifying microplastics ingested by
485 fish and invertebrates. *Analytical Methods* 9, 1346-1360.

486

487 Lusher, A.L., O'Donnell, C., Officer, R., O'Connor, I., 2015. Microplastic interactions with North Atlantic
488 mesopelagic fish. *ICES Journal of Marine Science: Journal du Conseil*.

489

490 Mazurais, D., Ernande, B., Quazuguel, P., Severe, A., Huelvan, C., Madec, L., Mouchel, O., Soudant, P., Robbens, J.,
491 Huvet, A., Zambonino-Infante, J., 2015. Evaluation of the impact of polyethylene microbeads ingestion in
492 European sea bass (*Dicentrarchus labrax*) larvae. *Marine Environmental Research* 112, 78-85.

493

494 McGoran, A., Clark, P., Morritt, D., 2017. Presence of microplastic in the digestive tracts of European flounder,
495 *Platichthys flesus*, and European smelt, *Osmerus eperlanus*, from the River Thames. *Environmental Pollution* 220,
496 744-751.

497

498 Millner, R., Walsh, S.J., Diaz de Astarloa, J.M., 2005. Atlantic flatfish fisheries. *Flatfishes: biology and exploitation*,
499 240-271.

500

501 Miranda, D.d.A., de Carvalho-Souza, G.F., 2016. Are we eating plastic-ingesting fish? *Marine Pollution Bulletin*
502 103, 109-114.

503

504 Murphy, F., Russell, M., Ewins, C., Quinn, B., 2017. The uptake of macroplastic & microplastic by demersal &
505 pelagic fish in the Northeast Atlantic around Scotland. *Marine Pollution Bulletin* 122, 353-359.

506

507 Nelms, S.E., Galloway, T.S., Godley, B.J., Jarvis, D.S., Lindeque, P.K., 2018. Investigating microplastic trophic
508 transfer in marine top predators. *Environmental Pollution*.

509

510 O'Connell, M., Fives, J.M., 1995. The Biology of the Lesser Sand-Eel *Ammodytes tobianus* L. In the Galway Bay
511 Area. *Biology and Environment: Proceedings of the Royal Irish Academy* 95B, 87-98.

512

513 Parapar, J., Fernández, L., González-Gurriarán, E., MUÍNO, R., 1997. Epibiosis and masking material in the spider
514 crab *Maja squinado* (Decapoda: Majidae) in the Ria de Arousa (Galicia, NW Spain). *Cahiers de Biologie marine* 38,
515 221-234.

516

517 Pedà, C., Caccamo, L., Fossi, M.C., Gai, F., Andaloro, F., Genovese, L., Perdichizzi, A., Romeo, T., Maricchiolo, G.,
518 2016. Intestinal alterations in European sea bass *Dicentrarchus labrax* (Linnaeus, 1758) exposed to microplastics:
519 Preliminary results. *Environmental Pollution* 212, 251-256.

520

521 Reisser, J., Slat, B., Noble, K., Du Plessis, K., Epp, M., Proietti, M., De Sonneville, J., Becker, T., Pattiaratchi, C.,
522 2015. The vertical distribution of buoyant plastics at sea: an observational study in the North Atlantic Gyre.
523 *Biogeosciences* 12, 1249.

524

525 Rijnsdorp, A.D., Vingerhoed, B., 2001. Feeding of plaice *Pleuronectes platessa* L. and sole *Solea solea* (L.) in
526 relation to the effects of bottom trawling. *Journal of Sea Research* 45, 219-229.

527

528 Rindorf, A., Wanless, S., Harris, M., 2000. Effects of changes in sandeel availability on the reproductive output of
529 seabirds. *Marine Ecology Progress Series* 202, 241-252.

530

531 Rochman, C.M., Tahir, A., Williams, S.L., Baxa, D.V., Lam, R., Miller, J.T., Teh, F.-C., Werorilangi, S., Teh, S.J., 2015.
532 Anthropogenic debris in seafood: Plastic debris and fibers from textiles in fish and bivalves sold for human
533 consumption. *Scientific Reports* 5, 14340.

534

535 Romeo, T., Pietro, B., Pedà, C., Consoli, P., Andaloro, F., Fossi, M.C., 2015. First evidence of presence of plastic
536 debris in stomach of large pelagic fish in the Mediterranean Sea. *Marine Pollution Bulletin* 95, 358-361.

537

538 Rummel, C.D., Löder, M.G.J., Fricke, N.F., Lang, T., Griebeler, E.-M., Janke, M., Gerdts, G., 2016. Plastic ingestion
539 by pelagic and demersal fish from the North Sea and Baltic Sea. *Marine Pollution Bulletin* 102, 134-141.

540

541 Tanaka, K., Takada, H., 2016. Microplastic fragments and microbeads in digestive tracts of planktivorous fish from
542 urban coastal waters. *Scientific reports* 6, 34351.

543

544 Tosetto, L., Brown, C., Williamson, J.E., 2016. Microplastics on beaches: ingestion and behavioural consequences
545 for beachhoppers. *Marine Biology* 163, 199.

546

547 Van Cauwenberghe, L., Claessens, M., Vandegehuchte, M.B., Janssen, C.R., 2015. Microplastics are taken up by
548 mussels (*Mytilus edulis*) and lugworms (*Arenicola marina*) living in natural habitats. *Environmental Pollution* 199,
549 10-17.

550

551 Van Cauwenberghe, L., Janssen, C.R., 2014. Microplastics in bivalves cultured for human consumption.
552 Environmental Pollution 193, 65-70.

553

554 Watts, A.J., Lewis, C., Goodhead, R.M., Beckett, S.J., Moger, J., Tyler, C.R., Galloway, T.S., 2014. Uptake and
555 retention of microplastics by the shore crab *Carcinus maenas*. Environmental science & technology 48, 8823-
556 8830.

557

558 Watts, A.J., Urbina, M.A., Corr, S., Lewis, C., Galloway, T.S., 2015. Ingestion of Plastic Microfibers by the Crab
559 *Carcinus maenas* and Its Effect on Food Consumption and Energy Balance. Environmental science & technology
560 49, 14597-14604.

561

562 Welden, N.A., Cowie, P.R., 2016a. Environment and gut morphology influence microplastic retention in
563 langoustine, *Nephrops norvegicus*. Environmental Pollution 214, 859-865.

564

565 Welden, N.A., Cowie, P.R., 2016b. Long-term microplastic retention causes reduced body condition in the
566 langoustine, *Nephrops norvegicus*. Environmental Pollution 218, 895-900.

567

568 Wesch, C., Barthel, A.-K., Braun, U., Klein, R., Paulus, M., 2016. No microplastics in benthic eelpout (*Zoarces*
569 *viviparus*): An urgent need for spectroscopic analyses in microplastic detection. Environmental research 148, 36-
570 38.

571

572