



[Ghashut, R. A.](#), McMillan, D. C., Kinsella, J., and Talwar, D. (2017) Erythrocyte concentrations of B1, B2, B6 but not plasma C and E are reliable indicators of nutrition status in the presence of systemic inflammation. *Clinical Nutrition ESPEN*, 17, pp. 54-62. (doi:[10.1016/j.clnesp.2016.10.007](https://doi.org/10.1016/j.clnesp.2016.10.007))

This is the author's final accepted version.

There may be differences between this version and the published version. You are advised to consult the publisher's version if you wish to cite from it.

<http://eprints.gla.ac.uk/139670/>

Deposited on: 19 April 2017

Enlighten – Research publications by members of the University of Glasgow
<http://eprints.gla.ac.uk>

1 **Erythrocyte concentrations of B1, B2, B6 but not plasma C and E are reliable indicators**
2 **of nutrition status in the presence of systemic inflammation.**

3 Rawia A Ghashut^{1, 2}, Donald C McMillan², John Kinsella ¹, Dinesh Talwar³.

4 1. Academic Unit of Anaesthesia, College of Medical, Veterinary and Life of Sciences-
5 University of Glasgow, Royal Infirmary, Glasgow G31 2ER.

6 2. Academic Unit of Surgery, College of Medical, Veterinary and Life of Sciences-
7 University of Glasgow, Royal Infirmary, Glasgow G31 2ER.

8 3. The Scottish Trace Element and Micronutrient Reference Laboratory, Department of
9 Biochemistry, Royal Infirmary, Glasgow G31 2ER.

10 Short running head: Vitamin B1, B2, B6, C and E concentrations and systemic inflammation.

11

12 Presented in part as an oral abstract at the European Society of Parenteral and Enteral
13 Nutrition (2012) in Gothenburg and Barcelona.

14

15 Correspondence to;

16 Dr Rawia A Ghashut

17 Academic unit of Anaesthesia College of Medical, Veterinary and Life of Sciences-
18 University of Glasgow, Royal Infirmary, Glasgow G31 2ER, United Kingdom.

19 Tel No. 0141 211 8647

20 Fax No. 0141 211 1191

21 Email: rgashot@yahoo.co.uk

22 **Abstract**

23 **Background & aim:** There is increasing evidence that the plasma concentration of vitamin
24 D, carotenoids, zinc and selenium are associated with the magnitude of the systemic
25 inflammatory response. In order to examine whether other vitamins may be affected and
26 whether red cell concentrations are less affected by systemic inflammation the aim of the
27 present study was to examine the effect of the systemic inflammatory response on red cell
28 measurements of vitamins B1, B2 and B6, and plasma concentration of vitamin C and E in a
29 large cohort of patients referred for a nutritional screen.

30 **Methods:** Patients referred for nutritional assessment of B1 (n=551), B2 (n=251), B6
31 (n=313), ascorbic acid (n=494) and α -tocopherol (n=395) concentrations. These vitamins
32 were measured using routine laboratory methods.

33 **Results:** The median concentrations of vitamin B1 grouped according to C-reactive protein
34 concentrations ≤ 10 , 11-80 and >80 mg/L were 543, 664 and 766 ng/g Hb respectively
35 ($p < 0.001$, 41% higher). The median concentration of vitamin B1 grouped according to
36 albumin concentrations ≥ 35 , 25-34 and < 25 g/l were 547, 664 and 701 ng/g Hb respectively
37 ($p < 0.001$, 28% higher). The median concentrations of red cell vitamin B2 grouped according
38 to CRP concentrations ≤ 10 , 11-80 and >80 mg/L were 2.2, 2.3 and 2.4 nmol/g Hb
39 respectively ($p < 0.001$, 9% higher). The median red cell concentrations of vitamin B2
40 grouped according to albumin concentrations ≥ 35 , 25-34 and < 25 g/l were 2.1, 2.4 and 2.3
41 nmol/g Hb respectively ($p < 0.001$, 14% higher). The median concentrations of red cell
42 vitamin B6 grouped according to CRP concentrations ≤ 10 , 11-80 and >80 mg/L were 534,
43 548 and 767 pmol/g Hb respectively ($p < 0.001$, 44% higher). The median red cell
44 concentrations of vitamin B6 grouped according to albumin concentrations ≥ 35 , 25-34 and
45 < 25 g/l were 462, 644 and 840 pmol/g Hb respectively ($p < 0.001$, 82% higher).

46 In contrast, the median plasma concentrations of ascorbic acid grouped according to CRP
47 concentrations ≤ 10 , 11-80 and >80 mg/L were 25.0, 15.0 and 6.0 $\mu\text{mol/l}$ respectively (78%
48 lower, $p < 0.001$). The median plasma concentrations of ascorbic acid grouped according to
49 albumin concentrations ≥ 35 , 25-34 and < 25 g/l were 32.0, 13.0 and 5.0 $\mu\text{mol/l}$ respectively
50 (84% lower, $p < 0.001$). The median α -tocopherol/ cholesterol grouped according to CRP
51 concentrations ≤ 10 , 11-80 and >80 mg/L were 5.9, 4.6 and 2.1 umol/l respectively (64%
52 lower, $p < 0.001$). The median α -tocopherol/cholesterol grouped according to albumin
53 concentrations ≥ 35 , 25-34 and < 25 g/l were 6.0, 5.5 and 2.1 umol/l respectively (65% lower,
54 $p < 0.001$).

55 **Conclusion:** Red cell concentrations of vitamins B1, B2 and B6 were not lower with an
56 increasing systemic inflammatory response. In contrast, plasma concentrations of vitamin C
57 and E were lower. Therefore, compared with plasma concentration, red cell concentrations of
58 B1, B2 and B6 are likely to be more reliable measures of status in the presence of a systemic
59 inflammatory response.

60 **Key words:** Vitamin B1, B2; B6, ascorbic acid; α -tocopherol; C-reactive protein; albumin.

61 **1.1. Introduction**

62 There is increasing evidence that the plasma concentration of a variety of trace
63 elements and vitamins are associated with the magnitude of the systemic inflammatory
64 response [1-4]. There was significant lowering of plasma concentrations of vitamins A, B6,
65 C, E, carotenoids and trace elements zinc and selenium associated with an elevated systemic
66 inflammatory response, when adjusted for CRP concentrations alone [3]. Indeed, it was
67 shown that as the severity of systemic inflammatory response increased, the proportion of
68 results below the lower reference limit increased. However, the variability of the association
69 was considerable such that plasma α -tocopherol and ascorbic acid concentrations were not
70 readily adjustable by CRP concentrations. Ghashut and co-workers (2013; 2014; 2016) have
71 recently reported that plasma concentrations of fat soluble micronutrients such as 25 (OH) D,
72 carotenoids, zinc and selenium may be confounded in the presence of a systemic
73 inflammatory response with both CRP and albumin having an independent effect on plasma
74 concentrations [5-7].

75 Therefore, it is of interest that red cell concentrations of some micronutrients appear
76 to be less affected by the magnitude of the systemic inflammatory response [8;9]. Therefore,
77 the aim of the present study was to examine the effect of the systemic inflammatory response,
78 as evidenced by both CRP and albumin, on red cell measurements of vitamins B1, B2 and
79 B6, and plasma measurements of vitamins C and E in a cohort of patients referred for
80 nutritional screen.

81 **1.2 Patients and Methods**

82 **1.2.1 Nutritional screen cohort red cell B1, B2, B6, C and E**

83 Consecutive heparin-treated whole-blood samples from individual patients that had
84 red cell B1 (n=553), B2 (n=251) and B6 (n=313) measurements were examined. These
85 samples were received from hospitals throughout Scotland between January 2008 and March
86 2013 for routine laboratory analysis. Consecutive heparin-treated whole-blood samples from
87 individual patients that had plasma vitamin C (n=494) and E (n=359) measurements were
88 examined. As a regional centre blood samples were sent for analysis if the patient was
89 considered at nutritional risk and was often secondary to a number of disease states. Only the
90 patients who had both CRP and albumin measured with the vitamins were included.

91 This investigation was conducted with the intent of developing local guidelines to aid
92 in the interpretation of vitamin B1, B2, B6, C and E results. This was a large convenience
93 retrospective sample that arose from an audit of patients who had a sample sent to a regional
94 laboratory for a nutritional screen. The patient was anonymised and de-identified prior to
95 analysis (Andrew Duncan and Donald C McMillan respectively). Approval for audit purposes
96 was obtained from the local ethics committee of the North Glasgow NHS Trust.

97 **1.2.2. Analytical methods**

98 Vitamins B1, B2 and B6 were assessed by measuring TDP, FAD and PLP
99 respectively in red blood cells. Vitamins C and E were assessed by measuring ascorbic acid
100 and α -tocopherol. These were measured by HPLC using established routine laboratory
101 methods previously described [1;10;11]. The inter assay precision was <10% for all analytes.

102 C-reactive protein and albumin were measured using an automated analyzer
103 (Architect; Abbot Diagnosis, Maidenhead, UK). The limits of detection for albumin and C-

104 reactive protein were 10g/L and 0.2 mg/L respectively. The inter-assay CV was <6% for
105 both analytes.

106 **1.2.3. Statistical analysis**

107 Data was presented in median and (range) value. Correlations between variables in
108 the convenience sample were carried out using the Spearman rank correlation. The cohorts
109 were divided into three groups according to CRP concentrations ≤ 10 , 11-80 and >80 mg/L as
110 previously described [12]. With each CRP group the concentration of individual vitamins
111 B1, B2, B6, ascorbic acid and α -tocopherol/ cholesterol were grouped according to 3
112 categories of albumin concentrations ≥ 35 , 25-34, < 25 g/L as previously described [13]. A P-
113 value < 0.05 was considered significant and the analysis of the data was carried out using
114 SPSS software (version 19; SPSS Inc, Chicago, Ill).

115 1.3. Results

116 1.3.1 Nutritional screen cohorts

117 The characteristics of the convenience sample for whole blood vitamin B1 (n=553)
118 are shown in **Table 1**. The majority were older than 50 years (median 54 years), male (50%)
119 and had plasma albumin concentrations below the normal range whereas plasma CRP and
120 whole blood vitamin B1 were in the normal range. Whole blood vitamin B1 was
121 significantly associated with CRP ($r_s= 0.290, p<0.001$) and albumin ($r_s= -0.265, p<0.001$).
122 These relationships are shown in Figures 1a, 1b and 1c.

123 The characteristics of the convenience sample for red cell vitamin B2 (n=251) are
124 shown in **Table 1**. The majority were older than 50 years (median 51 years), female (56%)
125 and had plasma albumin were below the normal range and plasma CRP concentration and red
126 cell vitamin B2 in the normal range. Red cell vitamin B2 was significantly associated with
127 CRP ($r_s= 0.189, p<0.01$) and albumin ($r_s= 0.186, p<0.001$). These relationships are shown in
128 Figures 2a, 2b and 2c.

129 The characteristics of the convenience sample for red cell vitamin B6 (n=313) are
130 shown in **Table 1**. The majority were older than 50 years (median 51 years), female (60%)
131 and had plasma albumin below the normal range and plasma CRP and red cell vitamin B6 in
132 the normal range. Red cell vitamin B6 was significantly associated with and albumin ($r_s=-$
133 $0.217, p<0.001$). These relationships are shown in Figures 3a, 3b and 3c.

134 The characteristics of the convenience sample for ascorbic acid (n= 494) are also
135 shown in **Table 1**. The majority were older than 50 years (median 54 years), female (53%),
136 and had plasma CRP and albumin and plasma ascorbic acid in the normal range. Plasma

137 ascorbic acid was significantly associated with CRP ($r_s = -0.333$, $p < 0.001$) and albumin
138 ($r_s = 0.427$, $p < 0.001$). These relationships are shown in Figures 4a, 4b and 4c.

139 The characteristics of the convenience sample for α -tocopherol ($n = 359$) are shown in
140 **Table 1**. The majority were older than 50 years (median 52 years), male (52%) and had
141 plasma albumin and cholesterol below the normal range, while plasma CRP and plasma α -
142 tocopherol were in the normal range. Plasma α -tocopherol adjusted to cholesterol was
143 significantly associated with CRP ($r_s = -0.424$, $p < 0.001$) and albumin ($r_s = 0.531$, $p < 0.001$).
144 These relationships are shown in Figures 5a, 5b and 5c.

145 **Nutritional screen cohort for whole blood vitamin B1**

146 The median concentrations of whole blood vitamin B1 (**Table 2**) grouped according
147 to CRP concentrations ≤ 10 , 11-80 and > 80 mg/L were 543, 664 and 766 ng/g Hb respectively
148 ($p < 0.001$) with an overall elevation of 41%. The median whole blood concentrations of
149 vitamin B1 grouped according to albumin concentrations ≥ 35 , 25-34 and < 25 g/l were 547,
150 664 and 701 ng/g Hb respectively ($p < 0.001$) with an overall elevation of 28%.

151 When albumin concentrations were ≥ 35 g/L, the median concentrations of whole
152 blood vitamin B1 grouped according to CRP concentrations ≤ 10 , 11-80 and > 80 were 535,
153 619 and 812 ng/g Hb respectively ($p = 0.003$) with an overall elevation of 52%. When albumin
154 concentrations were 25-34 g/L, the median concentrations of whole blood vitamin B1
155 grouped according to CRP concentrations ≤ 10 , 11-80 and > 80 mg/L were 611, 693 and 776
156 ng/g Hb respectively ($p = 0.011$) with an overall elevation of 27%. When albumin
157 concentrations were < 25 g/L, the median concentrations of whole blood vitamin B1 grouped
158 according to CRP concentrations ≤ 10 , 11-80 and > 80 mg/L were 574, 671 and 745 ng/g Hb
159 respectively ($p = 0.118$) with an overall elevation of 30%. Median concentrations of vitamin

160 B1/ albumin x100 were significantly increased from 1450 to 2250 to 4030 respectively
161 ($p<0.001$).

162 **1.3.2. Nutritional screen cohort for red cell vitamin B2**

163 The median concentrations of red cell vitamin B2 (**Table 3**) grouped according to
164 CRP concentrations ≤ 10 , 11-80 and >80 mg/L were 2.2, 2.3 and 2.4 nmol/g Hb respectively
165 ($p<0.001$) with an overall elevation of 9%. The median concentrations of red cell vitamin B2
166 grouped according to albumin concentrations ≥ 35 , 25-34 and <25 g/l were 2.1, 2.4 and 2.3
167 nmol/g Hb respectively ($p<0.001$).

168 When albumin concentrations were ≥ 35 g/L, the median red cell concentrations of
169 vitamin B2 grouped according to CRP concentrations ≤ 10 and 11-80 were 2.1 and 2.2 nmol/g
170 Hb respectively ($p=0.147$) with an overall elevation of 4%. When albumin concentrations
171 were 25-34 g/L, the median red cell concentrations of vitamin B2 grouped according to CRP
172 concentrations ≤ 10 and 11-80 mg/L were 2.4 and 2.4 nmol/g Hb respectively ($p=0.265$).
173 When albumin concentrations were < 25 g/L, the median red cell concentrations of vitamin
174 B2 grouped according to CRP concentrations ≤ 10 , 11-80 and >80 mg/L were 2.5, 2.3 and 2.5
175 nmol/g Hb respectively ($p=0.334$). Median red cell concentrations of vitamin B2/ albumin
176 x100 were significantly increased from 6 to 8 to 14 respectively ($p<0.001$).

177 **1.3.3. Nutritional screen cohort for red cell vitamin B6**

178 The median concentrations of red cell vitamin B6 (**Table 4**) grouped according to
179 CRP concentrations ≤ 10 , 11-80 and >80 mg/L were 534, 548 and 767 pmol/g Hb respectively
180 ($p<0.001$) with an overall elevation of 9%. The median red cell concentrations of vitamin B2
181 grouped according to albumin concentrations ≥ 35 , 25-34 and <25 g/l were 462, 644 and 840
182 pmol/g Hb respectively ($p<0.001$).

183 When albumin concentrations were ≥ 35 g/L, the median red cell concentrations of
184 vitamin B6 grouped according to CRP concentrations ≤ 10 and 11-80 were 478 and 413
185 pmol/g Hb respectively were not significantly different ($p=0.286$). When albumin
186 concentrations were 25-34 g/L, the median red cell concentrations of vitamin B6 grouped
187 according to CRP concentrations ≤ 10 and 11-80 mg/L were 818, 547 and 669 pmol/g Hb
188 respectively ($p=0.056$). When albumin concentrations were < 25 g/L, the median red cell
189 concentrations of vitamin B6 grouped according to CRP concentrations ≤ 10 , 11-80 and >80
190 mg/L were 1037, 822 and 81 pmol/g Hb respectively ($p=0.802$). Median red cell
191 concentrations of vitamin B6/ albumin x100 were significantly increased from 1380 to 2030
192 to 3560 respectively ($p<0.001$).

193 **1.3.4. Nutritional screen cohort for plasma ascorbic acid.**

194 The median plasma concentrations of ascorbic acid (**Table 5**) grouped according to
195 CRP concentrations ≤ 10 , 11-80 and >80 mg/L were 25.0, 15.0 and 6.0 $\mu\text{mol/l}$ respectively
196 ($p<0.001$) with an overall reduction of 78%. The median plasma concentrations of ascorbic
197 acid grouped according to albumin concentrations ≥ 35 , 25-34 and <25 g/l were 32.0, 13.0 and
198 5.0 $\mu\text{mol/l}$ respectively ($p<0.001$) with an overall reduction of 84%.

199 When albumin concentrations were ≥ 35 g/L, the median plasma concentrations of
200 ascorbic acid grouped according to CRP concentrations ≤ 10 , 11-80 and >80 mg/L were 33.0,
201 30 and 31 $\mu\text{mol/l}$ ($p=0.765$). When albumin concentrations were 25-34 g/L, the median
202 plasma concentrations of ascorbic acid grouped according to CRP concentrations ≤ 10 , 11-80
203 and >80 mg/L were 12.0, 15 and 10.0 $\mu\text{mol/l}$ ($p=0.409$) with an overall reduction of 17%.
204 When albumin concentrations were < 25 g/L, the median plasma concentrations of ascorbic
205 acid grouped according to CRP concentrations ≤ 10 , 11-80 and >80 mg/L were 3, 10 and 5.0
206 $\mu\text{mol/l}$ ($p=0.121$). The median vitamin C/albumin ration x 100 for CRP concentrations ≤ 10 ,
207 11-80 and >80 mg/L were 71.8, 53.5 and 37.3 respectively (48%, $p=0.050$). The median

208 vitamin C /CRP x 100 for albumin concentrations ≥ 35 , 25-34 and < 25 g/l were 711.5, 65.8
209 and 10.3 (99%, $p < 0.001$).

210 **1.3.5. Nutritional screen cohort for α -tocopherol / cholesterol**

211 The median α -tocopherol/ cholesterol (**Table 6**) grouped according to CRP
212 concentrations ≤ 10 , 11-80 and > 80 mg/L were 5.9, 4.6 and 2.1 $\mu\text{mol/l}$ respectively ($p < 0.001$)
213 with an overall reduction of 64%. The median α -tocopherol/cholesterol grouped according to
214 albumin concentrations ≥ 35 , 25-34 and < 25 g/l were 6.0, 5.5 and 2.1 $\mu\text{mol/l}$ respectively
215 ($p < 0.001$) with an overall reduction of 65%.

216 When albumin concentrations were ≥ 35 g/L, the median α -tocopherol/cholesterol
217 grouped according to CRP concentrations ≤ 10 and 11-80 mg/L were 6.2 and 5.4 ($p = 0.134$).
218 When albumin concentrations were 25-34 g/L, the median α -tocopherol/cholesterol grouped
219 according to CRP concentrations ≤ 10 , 11-80 and > 80 mg/L were 5.2, 6.0 and 4.6 ($p = 0.518$)
220 with an overall reduction of 12%. When albumin concentrations were < 25 g/L, The median
221 α -tocopherol/cholesterol grouped according to CRP concentrations ≤ 10 , 11-80 and > 80 mg/L
222 were 3.8, 2.4 and 1.7 ($p = 0.032$) with an overall reduction of 55%.

223 **1.4. Discussion**

224 The results of the present study show that red cell concentrations of vitamin B1, B2
225 and B6 were not lower with an increasing magnitude of the systemic inflammatory response
226 (as evidenced by both CRP and albumin concentrations). Indeed, there was a relatively small
227 but significant increase in red cell B1, B2 and B6 concentrations. In contrast, plasma
228 measurements of vitamin C and E were lower with an increasing magnitude of the systemic
229 inflammatory response with the potential for misclassification of deficiency. Therefore, red
230 cell measurements appear to obviate the potential for misclassification of vitamin B1, B2 and
231 B6 deficiency in the presence of systemic inflammatory response. On this basis red cell
232 measurements are recommended for the routine assessment of B-vitamin status.

233 The present relationship between the systemic inflammatory response and low plasma
234 vitamin C and E results are consistent with that previously reported for vitamins A, D and
235 carotenoids. Taken together these results would suggest that the systemic inflammatory
236 response is associated with a redistribution of plasma vitamins, perhaps to specific tissues and
237 organs irrespective of vitamin status. Despite this strong and consistent association
238 recognition of this phenomenon remains limited and plasma measurements of vitamins
239 continue to be used to assess status. Therefore, in order to obviate such interpretative
240 problems it is essential that plasma vitamin measurements are accompanied by a measure of
241 the systemic inflammatory response such as CRP and albumin. Alternatively, that new
242 methods of measurements are developed for the assessment of vitamins A, C, D, E and the
243 carotenoids, that better reflect nutritional status.

244 The results of the present study are particularly pertinent to plasma vitamin
245 measurements carried out in patients with critical illness since almost all patients will have
246 evidence of a systemic inflammatory response and may have a true deficiency and a need for
247 supplementation. This may explain the mixed results of vitamin supplementation in the

248 critically ill [14]. In contrast, measurements of thiamine, FAD and PLP concentrations in
249 blood may be considered to more reliably reflect nutritional status of vitamin B1, B2 and B6
250 [1-4] and more reliably benefit from supplementation [14].

251 Vitamin C and E status is usually assessed by plasma measurement of α -tocopherol
252 and ascorbic acid respectively, although their correlation with concentrations in tissues is not
253 clearly established. Moreover, plasma concentrations of vitamin E are strongly associated
254 with their carrier lipids, principally lipoprotein complexes, such as high density lipoprotein
255 (HDL), very low density lipoprotein (VLDL) and chylomicrons. To overcome this limitation
256 it has been proposed that the plasma α -tocopherol concentrations should be expressed in
257 relation to plasma lipid concentrations, usually cholesterol [15-17]. In particular, the results
258 of the present study question the use of the α -tocopherol adjusted for cholesterol to
259 compensate for the effect of the systemic inflammatory response. Since, when expressed per
260 mmol of cholesterol, α -tocopherol, concentrations were lower with decreasing concentrations
261 of albumin.

262 The role of vitamin C in alleviating the oxidant stress and recycling the oxidized α -
263 tocopherol is well recognised. Plasma α -tocopherol concentrations, in addition to being
264 redistributed as part of the systemic inflammatory response, have been reported to be
265 regenerated by vitamin C. Early work on fat autoxidation performed by Golumbic and
266 Mattill (1941) reported the antioxygenic action of ascorbate in association with tocopherols
267 [18]. More recently, in vitro studies have shown that ascorbic acid reduces the tocopheroxyl
268 radical [19] and thereby restores the radical-scavenging activity of tocopherol [20-24]. It
269 would appear that the tocopheroxyl radical that forms in membranes reacts with ascorbic acid
270 to yield tocopherol and the ascorbyl radical, the result of which is to maintain radical
271 scavenging potential within the membrane by regenerating tocopherol and to transfer the
272 oxidative challenge to the aqueous phase. This scheme is consistent with the observation in

273 the present study that plasma vitamin C concentrations were lower compared with vitamin E
274 concentrations for the same magnitude of the systemic inflammatory response and even in
275 patients with critical illness in whom vitamin C concentrations were recognised to be
276 profoundly low [25-27].

277 The importance of the systemic inflammatory response in determining plasma vitamin
278 C concentrations is highlighted by the relative inefficiency, whereby supplementation can
279 prevent low concentrations following an inflammatory insult. For example, in healthy
280 patients with normal pre-operative plasma concentrations of ascorbic acid, a single oral
281 supplementation with 1,000 mg of ascorbic acid was unable to prevent the fall in
282 postoperative plasma concentrations [28]. Therefore, it is reasonable to conclude that
283 ascorbic acid deficiency and the need for supplementation should only be considered on the
284 basis of low plasma ascorbic acid concentrations in the presence of plasma CRP and albumin
285 concentrations in the normal range.

286 The present study has a number of limitations. In particular there was no information
287 on the presence of chronic disease, co-morbidity or on BMI and dietary intake. Nevertheless,
288 it is likely that the impact of these clinical factors on vitamins B1, B2, B6 C and E will
289 involve the systemic inflammatory response, at least in part.

290 In summary, the results of the present study show that, unlike plasma concentrations
291 of vitamins C and E, red cell concentrations of vitamin B1, B2 and B6 were not significantly
292 affected by the systemic inflammatory response. Therefore, red cell B1, B2 and B6
293 concentrations are likely to be a more reliable measure of status in patients with evidence of a
294 systemic inflammatory response

295 **Acknowledgement**

296 The authors acknowledge the Libyan government for funding.

297 The authors gratefully acknowledge the assistance of Denis O'Reilly, Andrew
298 Duncan, Pamela Moyes, Karen Elliot, Allison McLaughlin, Lesley Stuart and Isobel Hogg,
299 Scottish Reference Laboratory for Vitamins and Trace Elements, Glasgow Royal Infirmary.

300 **Declaration**

301 The authors confirm that there are no conflicts of interest.

302 **Authors' statements**

303 DCM, JK and DT conceived the idea of examining the relationship between red cell vitamin
304 B1, B2 and B6, CRP and albumin concentrations in a large cohort and in patients with critical
305 illness and funded the study. RAG and DCM performed the statistical analysis. All authors
306 contributed to the drafts and final version of the paper and are the guarantors.

307 Reference List

308

- 309 [1] Talwar D, Quasim T, McMillan DC, Kinsella J, Williamson C, O'Reilly DS.
310 Pyridoxal phosphate decreases in plasma but not erythrocytes during systemic
311 inflammatory response. *Clinical Chemistry* 2003; 49: 515-518.
- 312 [2] Quasim T, McMillan DC, Talwar D, Vasilaki A, O'Reilly DSJ, Kinsella J. The
313 relationship between plasma and red cell B-vitamin concentrations in critically-ill
314 patients. *Clinical Nutrition* 2005; 24: 956-960.
- 315 [3] Duncan A, Talwar D, McMillan DC, Stefanowicz F, O'Reilly DS. Quantitative data
316 on the magnitude of the systemic inflammatory response and its effect on
317 micronutrient status based on plasma measurements. *American Journal of Clinical
318 Nutrition* 2012; 95: 64-71.
- 319 [4] Gray A, McMillan DC, Wilson C, Williamson C, O'Reilly DSJ, Talwar D. The
320 relationship between plasma and red cell concentrations of vitamins thiamine
321 diphosphate, flavin adenine dinucleotide and pyridoxal 5-phosphate following
322 elective knee arthroplasty. *Clinical Nutrition* 2004; 23: 1080-1083.
- 323 [5] Ghashut A, McMillan D, Kinsella J, Duncan A, Talwar D. Quantitative data on the
324 magnitude of the systemic inflammatory response and its effect on carotenoids status
325 based on plasma measurements. *Journal of clinical nutrition* 2013; 8: e193-e199.
- 326 [6] Ghashut RA, Talwar D, Kinsella J, Duncan A, McMillan DC. The effect of the
327 systemic inflammatory response on plasma vitamin 25 (OH) D concentrations
328 adjusted for albumin. *Plos One* 2014; 9: e92614.
- 329 [7] Ghashut RA, McMillan DC, Kinsella J, Vasilaki AT, Talwar D, Duncan A. The effect
330 of the systemic inflammatory response on plasma zinc and selenium adjusted for
331 albumin. *Clinical Nutrition* 2016; 35: 381-388.
- 332 [8] Stefanowicz F, Gashut RA, Talwar D, Duncan A, Beulshausen JF, McMillan DC,
333 Kinsella J. Assessment of plasma and red cell trace element concentration, disease
334 severity, and outcome in patients with critical illness. *Journal of Critical Care* 2013; 1-
335 5.
- 336 [9] Talwar D, Quasim T, McMillan DC, Kinsella J, Williamson C, O'Reilly DS.
337 Optimisation and validation of a sensitive high-performance liquid chromatography
338 assay for routine measurement of pyridoxal 5-phosphate in human plasma and red
339 cells using pre-column semicarbazide derivatisation. *Journal of Chromatography B-
340 Analytical Technologies in the Biomedical and Life Sciences* 2003; 792: 333-343.
- 341 [10] Margolis SA, Davis TP. Stabilization of Ascorbic-Acid in Human-Plasma, and Its
342 Liquid-Chromatographic Measurement. *Clinical Chemistry* 1988; 34: 2217-2223.

- 343 [11] Talwar D, Ha TKK, Cooney J, Brownlee C, SO'Reilly D. A routine method for the
344 simultaneous measurement of retinol, alpha-tocopherol and five carotenoids in human
345 plasma by reverse phase HPLC. *Clinica Chimica Acta* 1998; 270: 85-100.
- 346 [12] Marsik C, Kazemi-Shirazi L, Schickbauer T, Winkler S, Joukhadar C, Wagner O,
347 Endler G. C-Reactive Protein and All-Cause Mortality in a Large Hoepital-Based
348 Cohort. *Clinical Chemistry* 2008; 54: 343-349.
- 349 [13] Goldwasser P, Feldman J. Association of serum albumin and mortality risk. *Journal of*
350 *Clinical Epidemiology* 1997; 50: 693-703.
- 351 [14] Berger M. Micronutrients. In: Preiser J (ed.), *The stress response of critical illness:*
352 *Metabolic and hormonal aspects.* Springer International Publishing Switzerland 2016;
353 2016: 107-122.
- 354 [15] Vasilaki AT, Leivaditi D, Talwar D, Kinsella J, Duncan A, O'Reilly DS, McMillan
355 DC. Assessment of vitamin E status in patients with systemic inflammatory response
356 syndrome: Plasma, plasma corrected for lipids or red blood cell measurements?
357 *Clinica Chimica Acta* 2009; 409: 41-45.
- 358 [16] Doise C, Aho LS, Quenot JP, Guillard JC, Zeller M, Vergely C, Aude H, Blettery B,
359 Rochette L. Plasma antioxidant status in septic critically ill patients: a decrease over
360 time. *Fundamental & Clinical Pharmacology* 2008; 22: 203-209.
- 361 [17] Thurnham DI, Davies JA, Crump BJ, Situnayake RD, Davis M. The use of different
362 lipids to express serum tocopherol-Lipid ratios for the measurment of vitamin E
363 status. *Annals of Clinical Biochemistry* 1986; 23: 514-520.
- 364 [18] Golumbic C, Mattill H. Antioxidants and the Autoxidation of Fats. XIII.
365 Antioxygenic Action of Ascorbic Acid in Association with Tocopherols,
366 Hydroquinones and Related Compounds. *Journal of the american chemical society*
367 1941; 63: 1279-1280.
- 368 [19] Packer JE, Slater TF, Willson RL. Direct observation of a free redical interaction
369 between vitamin E and vitamin C. *Nature* 1979; 278: 737-738.
- 370 [20] Doba T, Burton GW, Ingold KU. Antioxidant and cooxidant activity of vitamin C-
371 The effect of vitamin C either alone, or in the presence of vitamin E or a water soluble
372 vitamin E analog, upon of peroxidation of aqueous multilamellar phospholipid
373 liposomes. *Biochimica et Biophysica Acta* 1985; 835: 298-303.
- 374 [21] Lambelet P, Saucy F, Lölliger J. Chemical evidence for interactions between vitamin
375 E and vitamin C. *Experientia* 1985; 41: 1384-1388.
- 376 [22] Niki E, Tsuchiya J, Tanimura R, Kamiya Y. Regeneration of vitamin E from á-
377 chromanoxyl radical by glutathione and vitamin C. *Chemistry Letters* 1982; 11: 789-
378 792.
- 379 [23] Niki E. Interaction of ascorbate and á-tocopherol. *Annals of the New York Academy*
380 *of sciences* 1987; 498: 186-199.

- 381 [24] Wayner DDM, Burton GW, Ingold KU, Barclay LRC, Locke S.J. The relative
382 contributions of vitamin E, urate, ascorbate and proteins to the total peroxy radical
383 trapping activity of human blood plasma. *Biochimica et Biophysica Acta* 1987; 924:
384 408-419.
- 385 [25] Nathens AB, Neff MJ, Jurkovich GJ, Klotz P, Farver K, Ruzinski JT, Radella F,
386 Garcia I, Maier RV. Randomized, prospective trial of antioxidant supplementation in
387 critically ill surgical patients. *Annals of Surgery* 2002; 236: 814-822.
- 388 [26] Mishra V, Baines M, Wenstone R, Shenkin A. Markers of oxidative damage,
389 antioxidant status and clinical outcome in critically ill patients. *Annals of Clinical*
390 *Biochemistry* 2005; 42: 269-276.
- 391 [27] Schorah C, Downing C, Piriptsi A, Gallivan L, AL-Hazaa A, Sanderson M,
392 Bodenham A. Total vitamin C, ascorbic acid, and dehydroascorbic acid
393 concentrations in plasma of critically ill patients. *American Journal of Clinical*
394 *Nutrition* 1996; 63: 760-765.
- 395 [28] Ruemelin A, Doerr S, Fauth U. Single preoperative oral application of ascorbic acid
396 does not affect postoperative plasma levels of ascorbic acid. *Annals of Nutrition and*
397 *Metabolism* 2002; 46: 211-214.
398
399

Tables legends

Table 1. Characteristics of nutritional screen cohorts with measurements of vitamins B1, B2, B6, C and E concentrations.

Table 2. Distribution of whole blood vitamin B1 (ng/g Hb) according to C-reactive protein and albumin concentrations (n=553).

Table 3. Distribution of red cell vitamin B2 according to C-reactive protein and albumin concentrations (n=251).

Table 4. Distribution of red cell vitamin B6 (pmol/g Hb) according to C-reactive protein and albumin concentrations (n=312).

Table 5: Distribution of plasma vitamin C (ascorbic acid) ($\mu\text{mol/l}$) according to CRP and albumin concentrations (n=494).

Table 6: Distribution of plasma α -tocopherol adjusted to cholesterol according to CRP and albumin concentrations (n=359).

Figure legends

Figure 1a.The relationship between CRP (log 10) and whole blood vitamin B1 (ng/g Hb) ($r_s = 0.290, p < 0.001$).

Figure 1b.The relationship between albumin and whole blood vitamin B1 (nmol/l) ($r_s = -0.265, p < 0.001$).

Figure 1c.The relationship between vitamin B1 adjusted for albumin and CRP ($r_s = 0.583, p < 0.001$)

Figure 2a.The relationship between CRP (log 10) and red cells vitamin B2 (nmol/g Hb) ($r_s = 0.189, p = 0.003$).

Figure 2b.The relationship between albumin and red cells vitamin B2 (nmol/g Hb) ($r_s = 0.186, p < 0.001$).

Figure 2c.The relationship between vitamin B2 adjusted for albumin and CRP ($r_s = 0.543, p < 0.001$).

Figure 3a.The relationship between CRP (log 10) and red cells vitamin B6 (pmol/g Hb) ($r_s = 0.079, p = 0.161$)

Figure 3b.The relationship between albumin and red cells vitamin B6 (pmol/g Hb) ($r_s = -0.217, p < 0.001$)

Figure 3c.The relationship between vitamin B6 adjusted to albumin and CRP ($r_s = 0.238, p < 0.001$) **4a:** The relationship between CRP (log 10) and vitamin C ($\mu\text{mol/l}$) ($r_s = -0.333, p < 0.001$)

Figure 4b: The relationship between albumin and vitamin C ($\mu\text{mol/l}$) ($r_s = 0.427, p < 0.001$).

Figure 4c: The relationship between CRP (log 10) and vitamin C adjusted to albumin ($r_s = -0.179, p < 0.001$).

Figure 5a: The relationship between α -tocopherol adjusted to cholesterol and CRP (log 10) ($r_s = -0.424, p < 0.001$)

Figure 5b: The relationship between α -tocopherol adjusted to cholesterol and albumin ($r_s = 0.531, p < 0.001$).

Figure 5c: The relationship between CRP (log 10) and vitamin α -tocopherol adjusted to cholesterol and albumin ($r_s = -0.094, p = 0.359$).

Table 1. Characteristics of nutritional screen cohorts with measurements of vitamins B1, B2, B6, C and E concentrations.

| | Reference interval | Nutritional screen cohort Whole blood vitamin B1 (n=553) | Nutritional screen cohort Vitamin B2 (n=251) | Nutritional screen cohort Vitamin B6 (n=313) | Nutritional screen cohort Vitamin C (n=494) | Nutritional screen cohort Vitamin E (n=359) |
|----------------------------------|---------------------------|---|---|---|--|--|
| Age (years) | NA | 54 (18-100) | 51 (16-82) | 51 (16-82) | 54 (16-90) | 52 (15-100) |
| Sex (Male/Female) | NA | 276 (50%)/ 277 (50%) | 111 (44%)/ 140 (56%) | 125 (40%)/ 188 (60%) | 234 (47%)/ 260 (53%) | 185 (52%)/ 174 (48%) |
| C-reactive protein (mg/l) | <10 | 14.0 (<10- 565.0) | 5.0 (<10- 565.0) | 4.3 (<10- 301.0) | 10.0 (0.20- 565.0) | 9.0 (0.20- 565.0) |
| Albumin (g/l) | 35-55 | 32 (6-50) | 36 (9-49) | 35 (9-49) | 33 (9-50) | 31 (9-50) |
| B1 (ng/g Hb) | 275- 675 | 613 (175-3650) | | | | |
| B2 (nmol/g Hb) | 1.0 – 3.4 | | 2.2 (1.2-27.3) | | | |
| B6 (pmol/g Hb) | 250 – 680 | | | 540 (79-100755) | | |
| Plasma ascorbic acid | 11-114 | | | | 17.0 (0.9-262.0) | |

| | | | | | | |
|---|---------------|--|--|--|--|-----------------|
| ($\mu\text{mol/l}$) | | | | | | |
| Plasma α-tocopherol ($\mu\text{mol/l}$) | 12 - 46 | | | | | 21.0 (2.9-58.0) |
| Cholesterol (mmol/l) | <5.2 | | | | | 4.5 (1.1-15.7) |
| α-tocopherol/ cholesterol ratio ($\mu\text{mol/ mmol}$) | 5.2 (3.5-8.7) | | | | | 5.2 (0.4-19.5) |

(Median and range)

Table 2. Distribution of whole blood vitamin B1 (ng/g Hb) according to C-reactive protein and albumin concentrations (n=553).

| whole blood vitamin B1 (ng/g Hb) | CRP ≤10 (mg/l) | 11-80 (mg/l) | >80 (mg/l) | whole blood vitamin B1 according to albumin |
|--|---------------------------|---------------------------|--------------------------|--|
| Albumin ≥35 (g/l) | 535 (211-2067) (n=175) | 619 (201-1508) (n=61) | (n=2) | 547 (201-2067) (n=238) |
| 34-25 (g/l) | 611 (175-1781) (n=56) | 693 (260-2552) (n=77) | 776 (385-3650) (n=18) | 664 (175-3650) (n=151) |
| <25 (g/l) | 574 (233-1248) (n=19) | 671 (288-2129) (n=71) | 745 (196-3524) (n=74) | 701 (196-3524) (n=164) |
| whole blood vitamin B1 according to CRP | 543 (175-2067) (n=250) | 664 (201-2552) (n=209) | 766 (196-3650) (n=94) | |

Median (range), where n<10 median and range not calculated.

Table 3. Distribution of red cell vitamin B2 according to C-reactive protein and albumin concentrations (n=251).

| Red cell B2 (nmol/g Hb | CRP ≤10 (mg/l) | 11-80 (mg/l) | >80 (mg/l) | Red cell B2 according to albumin |
|---|---------------------------|--------------------------|-------------------------|---|
| Albumin ≥35 (g/l) | 2.1 (1.2-5.9) (n=121) | 2.2 (1.9-3.1) (n=15) | - (n=0) | 2.1 (1.2-5.9) (n=136) |
| 34-25 (g/l) | 2.4 (1.2-27.3) (n=38) | 2.4 (1.6-5.8) (n=38) | - (n=1) | 2.4 (1.2-27.3) (n=77) |
| <25 (g/l) | (n=6) | 2.3 (1.3-3.5) (n=21) | 2.5 (2.0-5.5) (n=11) | 2.3 (1.3-5.5) (n=38) |
| Red cell B2 according to CRP | 2.2 (1.2-27.3) (n=165) | 2.3 (1.3-57.8) (n=74) | 2.4 (1.6-5.5) (n=12) | |

Median (range), where n<10 median and range not calculated.

Table 4. Distribution of red cell vitamin B6 (pmol/g Hb) according to C-reactive protein and albumin concentrations (n=312).

| Red cells B6 (pmol/g Hb) | CRP ≤10 (mg/l) | 11-80 (mg/l) | >80 (mg/l) | Red cells B6 according to albumin |
|--------------------------------------|----------------------------|----------------------------|---------------------------|--|
| Albumin ≥35 (g/l) | 478 (138-72374) (n=155) | 413 (181-2812) (n=18) | - (n=0) | 462 (138-72374) (n=173) |
| 34-25 (g/l) | 818 (79-61870) (n=47) | 547 (164-9245) (n=42) | (n=2) | 644 (79-61870) (n=91) |
| <25 (g/l) | 1037 (206-3036) (n=12) | 822 (183-100755) (n=24) | 881 (291-23294) (n=12) | 840 (183-100755) (n=48) |
| Red cells B6 according to CRP | 534 (79-27374) (n=214) | 548 (164-100775) (n=85) | 767 (291-23294) (n=14) | |

Median (range), where n<10 median and range not calculated

Table 5: Distribution of plasma vitamin C (ascorbic acid) ($\mu\text{mol/l}$) according to CRP and albumin concentrations (n=494).

| Plasma vitamin C (ascorbic acid) ($\mu\text{mol/l}$) | CRP ≤ 10 (mg/l) | 11-80 (mg/l) | >80 (mg/l) | Plasma C (ascorbic acid) according to albumin |
|---|--------------------------|----------------------------|----------------------------|--|
| Albumin ≥ 35 (g/l) | 33 (0.99-235) (n=175) | 30 (0.99-79.0) (n=44) | 31 (17.0-66.0) (n=4) | 32 (0.99-235) (n=223) |
| 34-25 (g/l) | 12 (0.90-126) (n=61) | 15 (0.99-136) (n=66) | 10 (0.99-61.0) (n=19) | 13 (0.90-136) (n=146) |
| <25 (g/l) | 3 (0.90-40.0) (n=12) | 10 (0.90-262) (n=50) | 5.0 (0.90-102.0) (n=63) | 5 (0.90-262) (n=125) |
| Plasma C (ascorbic acid) according to CRP | 25 (0.90-235) (n=248) | 15 (0.90-262.0) (n=160) | 6.0 (0.90-102.0) (n=86) | |

Median (range) (number of observation)

Table 6: Distribution of plasma α -tocopherol adjusted to cholesterol according to CRP and albumin concentrations (n=359).

| Plasma α -tocopherol / cholesterol ratio | CRP ≤ 10 (mg/l) | 11-80 (mg/l) | >80 (mg/l) | Plasma α -tocopherol / cholesterol ratio according to albumin |
|--|---------------------------|--------------------------|--------------------------|--|
| Albumin ≥ 35 (g/l) | 6.2 (3.2-10.7) (n=131) | 5.4 (3.3-10.8) (n=18) | - (n=1) | 6.0 (3.2-10.8) (n=150) |
| 34-25 (g/l) | 5.2 (0.97-10.4) (n=48) | 6.0 (1.2-9.7) (n=38) | 4.6 (1.3-8.3) (n=12) | 5.5 (1.0-10.4) (n=98) |
| <25 (g/l) | 3.8 (1.1-7.6) (n=15) | 2.4 (0.9-17.0) (n=31) | 1.7 (0.4-19.4) (n=65) | 2.1 (0.4-19.5) (n=111) |
| Plasma α -tocopherol / cholesterol ratio according to CRP | 5.9 (1.0-10.7) (n=194) | 4.6 (0.9-17.0) (n=87) | 2.1 (0.4-19.5) (n=78) | |

Median (range) (number of observation)

