



Gracia-Lor, E. et al. (2017) Measuring biomarkers in wastewater as a new source of epidemiological information: current state and future perspectives. *Environment International*, 99, pp. 131-150.
(doi:[10.1016/j.envint.2016.12.016](https://doi.org/10.1016/j.envint.2016.12.016))

This is the author's final accepted version.

There may be differences between this version and the published version. You are advised to consult the publisher's version if you wish to cite from it.

<http://eprints.gla.ac.uk/133949/>

Deposited on: 27 February 2017

1 **Measuring biomarkers in wastewater as a new source of epidemiological**
2 **information: current state and future perspectives**

3
4 **AUTHORS:**

5 Emma Gracia-Lor^{a,b*}, Sara Castiglioni^b, Richard Bade^a, Frederic Been^c, Erika Castrignanò^d, Adrian
6 Covaci^c, Iria González-Mariño^b, Evroula Hapeshi^e, Barbara Kasprzyk-Hordern^d, Juliet Kinyua^c,
7 Foon Yin Lai^c, Thomas Letzel^f, Luigi Lopardo^d, Markus R. Meyer^g, Jake O'Brien^h, Pedram Raminⁱ,
8 Nikolaos I. Rousis^b, Axel Rydevik^d, Yeonsuk Ryu^j, Miguel M. Santos^{k,l}, Ivan Senta^m, Nikolaos S.
9 Thomaidisⁿ, Sofia Veloutsou^f, Zhugen Yang^o, Ettore Zuccato^b, Lubertus Bijlsma^a

10
11
12 ^aResearch Institute for Pesticides and Water, Universitat Jaume I, Castellon, Spain

13 ^bIRCCS-Istituto di Ricerche Farmacologiche "Mario Negri", Department of Environmental Health
14 Sciences, Milan, Italy

15 ^cToxicological Center, University of Antwerp, 2610 Wilrijk, Belgium

16 ^dDepartment of Chemistry, Faculty of Science, University of Bath, Bath BA2 7AY, UK

17 ^eNIREAS-International Water Research Center, University of Cyprus, P.O. Box 20537, 1678
18 Nicosia, Cyprus

19 ^fAnalytical Group, Chair of Urban Water Systems Engineering, Technical University of Munich,
20 Germany

21 ^gDepartment of Experimental and Clinical Toxicology, Institute of Experimental and Clinical
22 Pharmacology and Toxicology, Saarland University, 66421 Homburg, Germany

23 ^hNational Research Center for Environmental Toxicology, The University of Queensland, Coopers
24 Plains, QLD 4108, Australia

25 ⁱDept. of Environmental Engineering, Technical University of Denmark, Denmark

26 ^jEcotoxicology and Risk Assessment, Norwegian Institute for Water Research, Oslo, Norway

27 ^k CIMAR/CIIMAR, LA-Interdisciplinary Centre for marine and Environmental Research,
28 University of Porto, Portugal

29 ^l FCUP—Dept of Biology, Faculty of Sciences, University of Porto, Rua do Campo Alegre, 4169-
30 007 Porto, Portugal

31 ^m Rudjer Boskovic Institute, Zagreb, Croatia

32 ⁿ Laboratory of Analytical Chemistry, Department of Chemistry, National and Kapodistrian
33 University of Athens, Panepistimiopolis Zografou, 15771 Athens, Greece

34 ^o Division of Biomedical Engineering, School of Engineering, University of Glasgow, G128LT
35 Glasgow, United Kingdom

36

37

38 * **Corresponding author:** Emma Gracia-Lor

39 E-mail: lor@uji.es; emma.gracialor@marionegri.it

40

41 **E-mail address of each author:**

42 Sara Castiglioni: sara.castiglioni@marionegri.it

43 Richard Bade: bade@uji.es

44

45 Frederic Been: frederic.been@uantwerpen.be

46 Erika Castrignanò: E.Castrignano@bath.ac.uk

47 Adrian Covaci: adrian.covaci@uantwerpen.be

48 Iria González-Mariño: iria.gonzalez@usc.es

49 Evroula Hapeshi: hapeshi.evroula@ucy.ac.cy

50 Barbara Kasprzyk-Hordern: B.Kasprzyk-Hordern@bath.ac.uk

51 Juliet Kinyua: juliet.kinyua@uantwerpen.be

52 Foon Yin Lai: FoonYin.Lai@uantwerpen.be

53 Thomas Letzel: t.letzel@tum.de
54 Luigi Lopardo: l.lopardo@bath.ac.uk
55 Markus R. Meyer: markus.meyer@med.uni-heidelberg.de
56 Jake O'Brien: j.obrien2@uq.edu.au
57 Pedram Ramin: pear@env.dtu.dk
58 Nikolaos I. Rousis: nikolaos.rousis@marionegri.it
59 Axel Rydevik: a.rydevik@bath.ac.uk
60 Yeonsuk Ryu: Yeonsuk.Ryu@niva.no
61 Miguel M. Santos: santos@ciimar.up.pt
62 Ivan Senta: isenta@irb.hr
63 Nikolaos S. Thomaidis: ntho@chem.uoa.gr
64 Sofia Veloutsou: sofia.veloutsou@tum.de
65 Zhugen Yang: Zhugen.Yang@glasgow.ac.uk
66 Ettore Zuccato: ettore.zuccato@marionegri.it
67 Lubertus Bijlsma: bijlsma@uji.es
68
69
70

71 **ABSTRACT**

72 The information obtained from the chemical analysis of specific human excretion products
73 (biomarkers) in urban wastewater can be used to estimate the exposure or consumption of the
74 population under investigation to a defined substance. A proper biomarker can provide relevant
75 information about lifestyle habits, health and wellbeing, but its selection is not an easy task as it
76 should fulfil several specific requirements in order to be successfully employed. This paper aims to
77 summarize the current knowledge related to the most relevant biomarkers used so far. In addition,
78 some potential wastewater biomarkers that could be used for future applications were evaluated. For
79 this purpose, representative chemical classes have been chosen and grouped in four main categories:
80 (i) those that provide estimates of lifestyle factors and substance use, (ii) those used to estimate the
81 exposure to toxicants present in the environment and food, (iii) those that have the potential to
82 provide information about public health and illness and (iv) those used to estimate the population
83 size. To facilitate the evaluation of the eligibility of a compound as a biomarker, information, when
84 available, on stability in urine and wastewater and pharmacokinetic data (*i.e.* metabolism and
85 urinary excretion profile) has been reviewed. Finally, several needs and recommendations for future
86 research are proposed.

87

88 **Key words**

89 Wastewater; Epidemiology; Biomarker; Consumption; Exposure; Population

90

91 **INTRODUCTION**

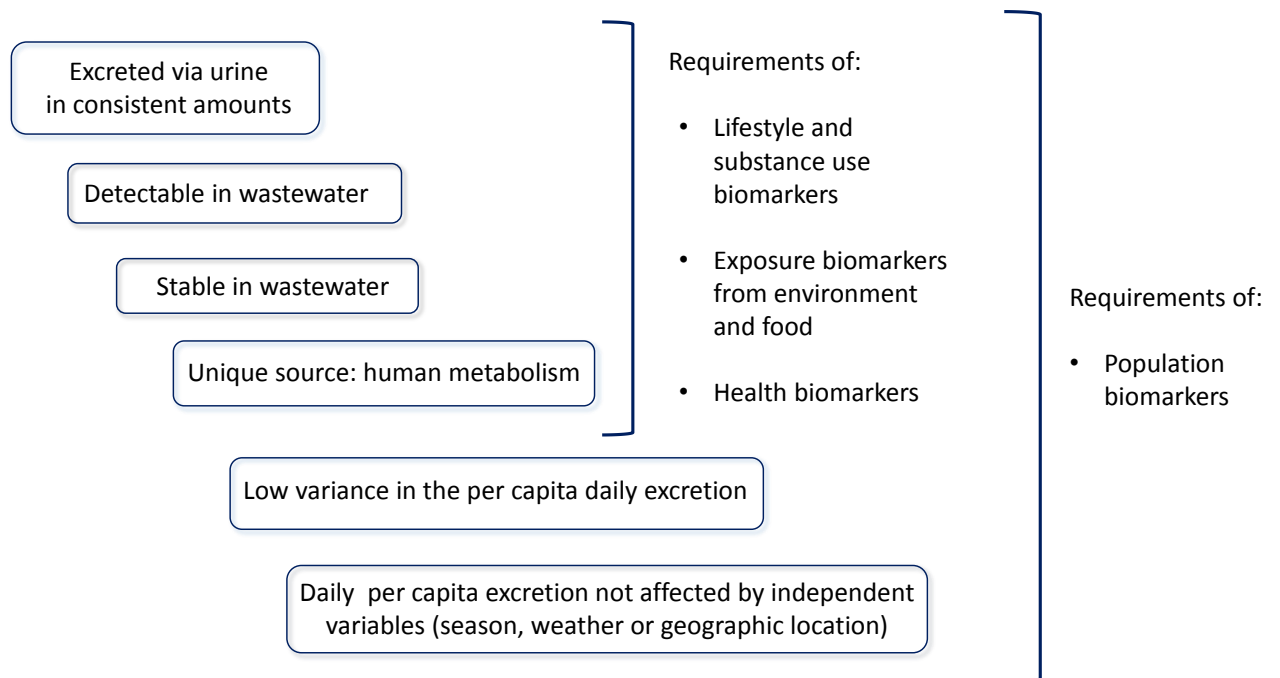
92 Relevant epidemiological information about lifestyle habits, public health and wellbeing can
93 be obtained from the chemical analysis of urban wastewater. This approach, called *wastewater-*
94 *based epidemiology* (WBE), is based on the analysis of specific human metabolic excretion
95 products (biomarkers) in wastewater as indicators of consumption or exposure of the population
96 served by the sewer network under investigation to different substances. WBE has been
97 successfully applied as a suitable approach for the estimation of illicit drugs consumption (Ort et al.,
98 2014; Thomaidis et al., 2016; Thomas et al., 2012; van Nuijs et al., 2011a; Zuccato et al., 2008), but
99 it has also recently been employed to assess other lifestyle-related factors such as alcohol
100 (Rodríguez-Álvarez et al., 2015; Ryu et al., 2016), nicotine (Castiglioni et al., 2015b; Lopes et al.,
101 2014; Rodríguez-Álvarez et al., 2014b), caffeine (Senta et al., 2015a) and new psychoactive
102 substances (NPS) (Kinyua et al., 2015; Reid et al., 2014a; van Nuijs et al., 2014). WBE has also
103 been applied to verify community-wide exposure to endocrine disruptors and antimicrobial agents
104 in personal care and household products (O'Brien et al., 2015; Rydevik et al., 2015). The broad
105 range of information that can be gathered from wastewater opens up the possibility of expanding
106 WBE to other human biomarkers providing clues about diet, health, diseases and exposure to
107 contaminants. For example by linking exposure to environmental or food contaminants with health
108 outcomes such as diabetes or cancer.

109 In general, a human biomarker can be an endogenous compound (produced naturally in the
110 body) or a metabolite of a xenobiotic/exogenous substance (produced through metabolic processes
111 after intentional consumption of a substance, accidental exposure to environmental contaminants, as
112 well as through diet or ingestion of a substance). Biomarkers can be classified on the basis of their
113 function as biomarkers of exposure (compounds that give information about substances consumed
114 or ingested) and biomarkers of effect (indicators of measurable changes or alterations in an
115 organism that can be associated with health problems or wellbeing) and on the basis of biological

116 nature (e.g. metabolites, hormones), or of the disease they can indicate (e.g. cardiovascular
117 biomarkers, obesity biomarkers) (Pischon, 2009).

118 The selection of a specific biomarker is not an easy task, as it needs to satisfy different
119 criteria (**Figure 1**) (Castiglioni and Gracia-Lor, 2015; Gracia-Lor et al., 2016). From a WBE
120 perspective, a suitable biomarker must be excreted mainly via urine and concentration levels in
121 urine should be at least in the $\mu\text{g/L}$ range to ensure its detection in raw wastewater after dilution
122 (Chen et al., 2014).

123



124

125 **Figure 1.** Main requirements of a biomarker

126

127 A biomarker should also be sufficiently stable in wastewater during the transport (in-sewer
128 stability) from the input (i.e. toilet) to the sampling point and during sampling, storage and analysis
129 (in-sample stability) (McCall et al., 2016a). In wastewater biomarkers can undergo further
130 transformation due to microbial activity (Mardal and Meyer, 2014) and/or sorption to particulate
131 matter (Baker and Kasprzyk-Hordern, 2011; Daughton, 2012a; McCall et al., 2016a). The fate of

132 biomarkers in the sewer can be also predicted by using mathematical models to simulate
133 physicochemical and microbial processes (Bisceglia and Lippa, 2014; McCall et al., 2016b; Ramin
134 et al., 2016). It is important to note that biomarker transformation pathways in the sewer might be
135 different from human metabolic pathways.

136 Furthermore, a biomarker should preferably be specific to the compound under investigation
137 and unique to human metabolism, thus ensuring that its presence only derives from human
138 excretion and not from exogenous sources (Daughton, 2012b). Therefore, pharmacokinetic data on
139 human metabolism are necessary but unfortunately this information is not always feasible as for
140 many substances it is very limited or do not even exist. This information, however, is highly
141 relevant not only to back-calculate the consumption/exposure of/to a certain substance by a
142 community, but also to distinguish the amount of a substance originating from human metabolism
143 or other sources. Unfortunately, pharmacokinetic studies are time-consuming and have to fulfil
144 strict ethical rules. Alternative approaches, which allow for the identification and selection of
145 appropriate biomarkers, are therefore required; for example, *in-vitro* studies using liver enzymes,
146 which metabolize the parent compound, help in the elucidation of the chemical structure of the
147 metabolites formed (i.e. possible biomarkers) formed (Mardal et al., 2016). Computer-based *in-*
148 *silico* modelling also allow the prediction of pharmacokinetics (Reid et al., 2014a). However these
149 alternatives provide qualitative information on metabolism, but not data regarding excretion rates of
150 parent substances and their metabolites (Gracia-Lor et al., 2016).

151 The present manuscript emerges within the framework of the pan-European inter-
152 disciplinary network (Sewage analysis CORE group-SCORE), which brings together experts from
153 different disciplines interested in standardizing the WBE approach and in coordinating international
154 studies (<http://score-cost.eu/>). The aim of this review is to describe the criteria for selecting suitable
155 biomarkers and to give an overview of relevant human (urinary) metabolites and potential
156 wastewater biomarkers. Biomarkers have been grouped in four sections: (i) those that provide

157 estimates of lifestyle factors and substance use, (ii) those used to estimate the exposure to toxicants
158 present in the environment and food, (iii) those giving information about public health and (iv)
159 those used to estimate the population size. For each group and biomarker, a thorough review of the
160 available pharmacokinetic data (*i.e.* metabolism and excretion profile) and stability in urine and
161 wastewater (if known) is provided. This information can be used to evaluate their suitability
162 according to the criteria described above. Finally, potential gaps or limitations are discussed and
163 future research directions are proposed.

164

165 **2. LIFESTYLE AND SUBSTANCE USE BIOMARKERS**

166 Initially, WBE was applied to evaluate lifestyle, in particular illicit drug use within a
167 community. Its ability to deliver objective and near-real-time data on drug use, being able to detect
168 changes over time and local patterns of use, suggests that this method can be used as a
169 complementary and extended data source to existing epidemiological tools. WBE has been well
170 established for monitoring the use of cocaine, cannabis, amphetamine, methamphetamine and
171 MDMA (3,4-methylenedioxymethamphetamine).

172 Additional applications to estimate consumption of other substances, such as alcohol,
173 tobacco, caffeine and NPS, have been employed more recently. Alcohol and nicotine (tobacco) are
174 probably the most popular and accepted recreational drugs. However, many negative social,
175 economic and health aspects have been linked to their use, causing millions of deaths every year
176 (World Health Organization, 2015, 2014). It is therefore important and of particular interest for
177 policy makers to obtain continuous monitoring data on consumption levels and patterns of use, in
178 order to reduce the disease burden related to alcohol and tobacco use. Caffeine use has been
179 limitedly investigated, although it is one of the most extensively used legal stimulants, found in
180 widely-consumed products, such as coffee, tea, soft and “energy” drinks. Besides monitoring its
181 consumption, caffeine has also been proposed as a human biomarker for assessing the size and

182 dynamics of the population served (see section 5.3) by a particular wastewater treatment plant
183 (WWTP) (Senta et al., 2015a). NPS are emerging narcotic or psychotropic substances which may
184 pose similar threats to public health such as classical illicit drugs (European Union, 2005; Papaseit
185 et al., 2014). Due to the delay between their appearance on the market and their addition to the list
186 of banned (or controlled) substances, many NPS can be legally purchased, thus promoting their
187 proliferation worldwide. Furthermore, new substances appear continuously on the market (Bijlsma
188 et al., 2016; EMCDDA, 2015a). WBE has been proposed as a tool for providing useful information
189 on temporal and regional trends in the use of NPS.

190 Current state and some new features of WBE, with regard to lifestyle and substance use are
191 presented in this chapter. Furthermore, specific biomarkers of each lifestyle factor are suggested
192 (**Table S1**) and conceptual approaches for dealing with NPSs using biomarkers in wastewater are
193 proposed.

194

195 **2.1. Illicit drugs**

196 Among the available epidemiological indicators, general population surveys have been
197 traditionally used to assess illicit drug use at the population level. Yet, due to their inherent biases,
198 complementary and real-time approaches are needed. The determination of illicit drug consumption
199 through wastewater was first theorized by Daughton (Daughton, 2001) and implanted by Zuccato *et*
200 *al.* using cocaine as an example (Zuccato et al., 2005). Since then, WBE has been widened to
201 include other illicit drugs (Asimakopoulos and Kannan, 2016; Castiglioni et al., 2008; Hernández et
202 al., 2016; van Nuijs et al., 2011a).

203 The biomarkers currently used are either the illicit drug itself (i.e. amphetamine,
204 methamphetamine, and 3,4-methylenedioxy-methamphetamine-MDMA) or one of its metabolites

205 (i.e. benzoylecgonine (BEG) for cocaine, 11-nor-9-carboxy-delta9-tetrahydrocannabinol (THC-
206 COOH) for cannabis and morphine or 6-acetylmorphine for heroin).

207 Cocaine, the first substance studied in WBE, is considered unstable in wastewater; however,
208 its unique and stable metabolite (BEG) makes back-calculation to drug consumption more
209 straightforward. It must be noted that significant degradation of BEG from cocaine in sewage is also
210 reported (Plósz et al., 2013), which could result in over estimation of cocaine consumption if this
211 formation is neglected. Considering human excretion rates, a cocaine: BEG ratio around 0.1 or
212 lower can indicate consumption, and any value higher (between 0.1 and 0.7) could indicate other
213 sources of cocaine, such as direct disposal (Castiglioni et al., 2011a). However, more research is
214 needed in this regard (Bijlsma et al., 2012; Postigo et al., 2010; Van Nuijs et al., 2009).

215 Δ 9-tetrahydrocannabinol (THC), the active ingredient of cannabis, is metabolized to more
216 than 20 metabolites after consumption, with 11-nor- Δ 9-carboxy-THC (THC-COOH) and 11-
217 hydroxy-THC (THC-OH) being those primarily excreted. THC-COOH has been shown to be highly
218 stable and is thus normally used to estimate cannabis consumption, albeit with some analytical
219 difficulties arising in multi-residue methods resulting from its non-polarity compared to other illicit
220 drugs (Bijlsma et al., 2014; Ort et al., 2014; Pedrouzo et al., 2011).

221 Two more recently works studied illicit drugs are ketamine and methadone. Ketamine is a
222 dissociative anaesthetic which has been used as a recreational drug, whilst methadone is a synthetic
223 opioid used clinically to relieve pain and also as maintenance treatment of opioid addicts
224 (Castiglioni et al., 2011b; Preston et al., 2003). Both ketamine and its metabolite norketamine are
225 fairly stable in wastewater (Castiglioni et al., 2015a; McCall et al., 2016a), with the parent
226 compound generally used as a biomarker for reliable estimation of drug usage. Variable stability for
227 methadone has, however, been reported i.e. from high (Senta et al., 2014) to low (González-Mariño
228 et al., 2010).

229 Opioids use in Europe remains a central issue, reflecting the significant impact these drugs
230 still have on mortality and morbidity (EMCDDA, 2015b). In recent years, the production of high
231 purity heroin has been rising, thereby increasing heroin-related mortality (UNODC, 2015). In the
232 human body, heroin is rapidly hydrolyzed to 6-monoacetylmorphine (6-MAM) by blood esterases
233 (Bencharit et al., 2003) and further hydrolyzed to morphine in the liver (Smith, 2009). In
234 wastewater, heroin shows low stability (González-Mariño et al., 2010). Although 6-MAM detected
235 in urine is used as a marker of heroin consumption (Staub et al., 2001), 6-MAM is not always
236 detected in wastewater as it is not stable in wastewater (Thai et al., 2014). Back-calculations using
237 6-MAM as biomarker provides inconsistent results (Been et al., 2015). Therefore, morphine is
238 considered as an alternative biomarker for heroin. However, therapeutic consumption of morphine
239 should be subtracted from the total measured morphine in sewage (Khan and Nicell, 2011; van
240 Nuijs et al., 2011a; Zuccato et al., 2016), which necessitates the availability of registered prescribed
241 morphine at the time of wastewater sampling. Morphine is also formed in the sewer due to
242 deconjugation of morphine glucuronide and deacetylation of 6-MAM, which imposes new
243 challenges in back-calculation schemes. Although fractions of morphine originating from codeine
244 can be considered negligible (Zuccato et al., 2008), more research is needed to find a drug
245 biomarker for heroin which fulfils all the aforementioned criteria.

246 As shown in **Table 1**, the most frequently used illicit drug biomarkers are benzoylecgonine,
247 amphetamine, methamphetamine, MDMA and THC-COOH (Thomas et al., 2012). Information
248 about excretion and stability in urine and wastewater of these and other illicit drug biomarkers less
249 frequently studied is presented in **Table S1**. One of the most current analytical challenges associated
250 with WBE is represented by chirality. Amphetamine, methamphetamine and MDMA are among the
251 illicit drugs that are chiral and as a result they can exist as enantiomers (one enantiomeric pair per
252 each chiral centre). The verification of their chiral signature in wastewater (i.e. relative proportion

253 of two enantiomers within each enantiomeric pair) allows to distinguish between illicit or licit use
254 and direct disposal (Emke et al., 2014). It has been shown that the distinction between the
255 consumption or the disposal of MDMA could be made by differentiating the loads of the
256 enantiomers present in wastewater. Indeed, enantiomeric fractions (EFs) greater than 0.5 indicated
257 illicit use, whilst EFs equal to 0.5 indicated direct disposal, when EF was calculated as follows:

$$EF = \frac{(-) - MDMA}{(-) - MDMA + (+) - MDMA}$$

258
259 Enantiomeric profiling of MDMA's metabolites were recently investigated in wastewater by
260 Castrignanò et al., suggesting enantioselective metabolism for HMMA (Castrignanò et al., 2016).
261 Amphetamine and methamphetamine can also be investigated at enantiomeric level, however due to
262 both legal and illicit uses, a clear understanding between consumption and direct disposal is
263 difficult (Emke et al., 2014; Kasprzyk-Hordern and Baker, 2012).

264

265 **2.2. Alcohol**

266 Following the consumption of alcoholic beverages, the majority of ingested ethanol is
267 rapidly metabolized in the human body in a two-stage oxidation process, first to acetaldehyde and
268 then to acetic acid. The remaining part is excreted unchanged in urine, sweat and exposed breath
269 (Jones, 1990). However, a very small fraction (<0.1%) undergoes a conjugation reaction with
270 glucuronic acid to produce ethyl glucuronide (EtG) (Dahl et al., 2002) and with 3'-
271 phosphoadenosine 5'-phosphosulfate to produce ethyl sulphate (EtS) (Helander and Beck, 2005).
272 These metabolites are excreted within a few hours and are detectable in urine for considerably
273 longer times (up to 1-2 days, depending on the subject and the alcohol dose) (Helander and Beck,
274 2005; Høiseth et al., 2008), making them unequivocal indicators of recent alcohol consumption
275 (Dahl et al., 2011; Dresen et al., 2004).

276 EtG was found to degrade ~50% after 18 hours, whereas EtS showed little or no degradation
277 (Reid et al., 2011). In addition, no significant differences were found between its stability in sewage
278 and in an ethanol-fortified wastewater sample (Reid et al., 2011), indicating that it is unlikely to be
279 formed from unconsumed alcohol discarded into the sewer system. Taking into account these
280 observations, EtS has been used by several researchers to estimate community-wide alcohol
281 consumption through wastewater analysis (**Table 1**). Typically, its determination in this matrix is
282 performed by direct injection, after filtration and/or centrifugation, into a liquid chromatography-
283 mass spectrometry system. The alcohol consumption rates estimated through WBE have revealed
284 specific drinking patterns, temporal and spatial variations. The study conducted by Reid et al. (Reid
285 et al., 2011), for example, clearly showed the weekend elevated drinking pattern in Oslo.
286 Furthermore, the estimated consumption rates were in good agreement with sales statistics (Reid et
287 al., 2011). The increase in alcohol consumption during the weekend was also found in three Spanish
288 cities, eight Belgian cities and one Italian city (Andrés-Costa et al., 2016; Boogaerts et al., 2016;
289 Mastroianni et al., 2014; Rodríguez-Álvarez et al., 2015, 2014a; Ryu et al., 2016). However, a
290 different consumption pattern was observed during a special event in Valencia, where an increased
291 alcohol use was noticeable, reaching the maximum rate on Wednesday, which corresponded to the
292 last day of the “Fallas” festivities (Andrés-Costa et al., 2016). Co-consumption of alcohol and
293 cocaine was also evaluated through WBE by analyzing cocaethylene, a specific biomarker excreted
294 when the two substances are consumed together (Mastroianni et al., 2014; Rodríguez-Álvarez et al.,
295 2015). In the studies carried out in Belgium (Boogaerts et al., 2016) and Greece (Gatidou et al.,
296 2016) higher alcohol consumption in urbanized cities than in smaller villages was evidenced.
297 Although all these studies highlight the potential of EtS as a reliable biomarker for estimating
298 alcohol consumption in relative terms, the main limitation is the uncertainty associated with its
299 percentage of excretion, which might lead to inaccurate back-calculations in absolute amounts.
300 Until now, there have been insufficient pharmacokinetic studies evaluating this percentage to

301 provide a unique, representative figure (Halter et al., 2008; Høiseth et al., 2008; Lostia et al., 2013;
302 Schneider and Glatt, 2004; Wurst et al., 2006). In the aforementioned WBE studies, the range
303 0.010-0.016% (on molar basis) was used by (Andrés-Costa et al., 2016; Reid et al., 2011); the
304 median value of the excretion rates provided by Høiseth et al. (Høiseth et al., 2008), 0.011%, was
305 used by (Mastroianni et al., 2014; Rodríguez-Álvarez et al., 2014a). Finally, four studies (Boogaerts
306 et al., 2016; Gatidou et al., 2016; Rodríguez-Álvarez et al., 2015; Ryu et al., 2016), employed a
307 people-weighted value of 0.012%, based on the data provided by (Høiseth et al., 2008) and (Wurst
308 et al., 2006).

309

310 **2.3. Tobacco**

311 Nicotine is the principal alkaloid found in tobacco and, although not being directly
312 associated with diseases, its addictiveness is the major cause of continued use of tobacco products
313 (Hukkanen, 2005). Nicotine is extensively metabolized in humans, with 70-80% of the initial dose
314 being converted to cotinine (Benowitz and Jacob, 1994), which is then further metabolized into
315 various compounds, the most abundant being *trans*-3'-hydroxycotinine (Byrd et al., 1992). Nicotine
316 and its major metabolites are also excreted as glucuronides. Globally, nicotine is excreted
317 unchanged at rates between 8 and 10%, whilst its glucuronide makes up for 3-5% of the initial dose
318 (Byrd et al., 1992). Cotinine and its glucuronide are excreted at rates between 10-15% and 12-17%,
319 respectively, while *trans*-3'-hydroxycotinine and its glucuronide make up for 33-40% and 7-9% of
320 the initial dose, respectively (Hukkanen, 2005).

321 Nicotine and its metabolites, cotinine and *trans*-3'-hydroxycotinine, have been analyzed in
322 wastewater as biomarkers (**Table S1**) to estimate tobacco use in various communities (Castiglioni et
323 al., 2015b; Lopes et al., 2014; Mackuľak et al., 2015; Rodríguez-Álvarez et al., 2014b; Senta et al.,
324 2015a). The three compounds were shown to be stable in wastewater samples stored at 4° C and 20°
325 C during 24 h (Chen et al., 2014; Rodríguez-Álvarez et al., 2014b; Senta et al., 2015a). However,

326 the concentration of the glucuronide of *trans*-3'-hydroxycotinine was shown to decrease even in
327 refrigerated samples (i.e., 35% decrease over 8 h at 4° C). The authors of the study thus suggested
328 to enzymatically deconjugate the compounds prior to extraction and analysis (Rodríguez-Álvarez
329 et al., 2014b).

330 The amounts of these compounds in wastewater range from 0.1 to 7 µg/L (Buerge et al.,
331 2008; Mackul'ak et al., 2015; Rodríguez-Álvarez et al., 2014b; Senta et al., 2015a), and the levels of
332 cotinine and *trans*-3'-hydroxycotinine reflected the excretion profiles expected from
333 pharmacokinetic studies, whilst nicotine was found at higher levels (Rodríguez-Álvarez et al.,
334 2014b; Senta et al., 2015a). The contribution from ashes and cigarettes butts has been advanced as a
335 possible explanation for this observation (Castiglioni et al., 2015b; Rodríguez-Álvarez et al., 2014b;
336 Senta et al., 2015a). In fact, higher nicotine levels have been reported during rain events, supporting
337 the hypothesis that ashes and cigarette butts found on streets eventually contribute to measured
338 nicotine loads (Senta et al., 2015a). Thus, cotinine and *trans*-3'-hydroxycotinine were used as
339 biomarkers to estimate the amount of nicotine used per capita in a population, as indicated in **Table**
340 **1** (Castiglioni et al., 2015b; Mackul'ak et al., 2015; Rodríguez-Álvarez et al., 2014b; Senta et al.,
341 2015a).

342 In some studies, figures were corrected to account for the portion of nicotine absorbed
343 during smoking (Castiglioni et al., 2015b; Mackul'ak et al., 2015), thus providing estimates of the
344 gross amount of number of cigarettes. Additionally, Mackul'ak and co-workers (Mackul'ak et al.,
345 2015) included a factor to account for losses due to degradation, based on the mean residence time
346 of wastewater in sewers. From the estimated nicotine consumption, the number of cigarettes
347 smoked per capita was also calculated using as reference value 0.8 mg of nicotine per cigarette
348 (Gorrod and Wahren, 1993; Lopes et al., 2014; Rodríguez-Álvarez et al., 2014b) or 1.25 mg of
349 nicotine (Castiglioni et al., 2015b). The obtained figures highlighted substantial differences in
350 consumption within the same country. For example, researchers from Italy found significant

351 differences between the north, centre and south of the country (Castiglioni et al., 2015b; Senta et al.,
352 2015a). These results were in agreement with epidemiological data, which suggested a higher
353 prevalence of tobacco use in the south (Castiglioni et al., 2015b). Similarly, important differences
354 were found in cities in Slovakia and Spain (Mackuľak et al., 2015; Rodríguez-Álvarez et al.,
355 2014b). In Portugal, estimates of nicotine consumption derived from wastewater analysis were in
356 line with findings from a European survey (Lopes et al., 2014).

357 Mass loads measured in wastewater were also used to investigate weekly consumption
358 patterns and findings suggested that this was stable throughout the week (Chen et al., 2014;
359 Rodríguez-Álvarez et al., 2014b; Senta et al., 2015a). Public holidays and specific touristic
360 locations, attracting larger crowds, were the only exceptions (Lopes et al., 2014; Mackuľak et al.,
361 2015).

362 The results obtained show that the measurement of nicotine metabolites is a useful tool
363 which could potentially be used to complete current knowledge about the prevalence of tobacco
364 use.

365

366 **2.4. Caffeine**

367 Caffeine (1,3,7-trimethylxanthine) is the world's most widely consumed stimulating agent
368 (Garattini, 1993). It is found in many globally popular products, including tea and cola drinks, as
369 well as in some medications and dietary supplements, but the most important source of this alkaloid
370 is coffee.

371 Caffeine metabolism is extensive (Baselt, 2004), with at least 17 urinary metabolites
372 identified in humans (Garattini, 1993). The major metabolites include 1-methyluric acid (excretion
373 rate 12-25%), 1-methylxanthine (9-18%), 7-methylxanthine (2-8%), paraxanthine (1,7-
374 dimethylxanthine; 4-7%), 1,7-dimethyluric acid (5-8%) and unstable product 5-acetylamino-6-
375 formylamino-3-methyluracil (4-15%), with a small percentage (1-4%) of the initial dose excreted as

376 the parent compound (Carrillo and Benitez, 1994; Garattini, 1993). The list of caffeine metabolites
377 identified in humans, together with the excretion rates can be found in **Table S1**. Besides being
378 complex, caffeine metabolism is also rather variable, with the different excretion rates observed not
379 only in different studies, but also between individuals within the same studies (Carrillo and Benitez,
380 1994; Grant et al., 1983). These variations can be related with genetic differences (Blanchard et al.,
381 1985; Grant et al., 1983) or influenced by other factors, such as age (Blanchard et al., 1985; Grant et
382 al., 1983), pregnancy ((Carrillo and Benitez, 1994; Garattini, 1993) or medications (Callahan et al.,
383 1983). However, certain metabolites, such as paraxanthine, 1,7-dimethyluric acid and 1-
384 methylxanthine were found to be less affected by the genetic background compared to the parent
385 compound and they were, therefore, suggested as potential biomarkers for caffeine dietary intake
386 (Crews et al., 2001). Furthermore, most of the pharmacokinetic data on caffeine metabolism in
387 humans are quite old (Blanchard et al., 1985; Grant et al., 1983) and some of them include a
388 relatively low number of subjects (Blanchard et al., 1985).

389 Due to its wide usage in modern societies, caffeine is among the most ubiquitous wastewater
390 micro-contaminants, usually detected at relatively high concentration levels ($\mu\text{g/L}$) in untreated
391 wastewater (Martínez-Bueno et al., 2011; Rosal et al., 2010; Santos et al., 2009). Due to this,
392 caffeine was proposed as anthropogenic marker to indicate the discharge of domestic wastewater in
393 rivers and lakes (Buerge et al., 2003), but so far has been rarely used as a biomarker in a WBE
394 approach. Caffeine has also been proposed as a human biomarker for assessing population size and
395 the dynamics of people served by a particular WWTP (Daughton, 2012b) (see section 5.3).

396 However, with the exception of paraxanthine, data on the occurrence of caffeine metabolites
397 in wastewater are still very scarce. In fact, the first comprehensive study which included most of the
398 major caffeine metabolites (1-methylxanthine, 7-methylxanthine and paraxanthine) was published
399 just recently (Senta et al., 2015a). Concentrations of these metabolites found in Italian wastewater
400 were similar to those of the parent compound, i.e. in the $\mu\text{g/L}$ range. In the same work temporal and

401 spatial patterns of use were also studied and the mean mass loads of caffeine and its major
402 metabolites revealed to be slightly lower during the weekend, probably due to the lower
403 consumption of coffee. Similar findings for caffeine was reported by Rico et al. (Rico et al., 2016;
404 Senta et al., 2015a). On the other hand, no clear geographical trends could be observed. Besides
405 being easily detectable, caffeine, 1-methylxanthine, 7-methylxanthine and paraxanthine fulfill
406 additional important requirement for an ideal biomarker - they are stable in wastewater samples
407 stored at 4 °C and 20 °C for 24 h (Senta et al., 2015a). However, it is noteworthy that more research
408 is needed in order to select the most suitable caffeine biomarker in wastewater for the correct
409 interpretation of the obtained results within the concept of WBE.

410

411 **2.5. New Psychoactive Substances**

412 The detection of NPS and the estimation of their use are especially challenging for drug
413 epidemiology, since new compounds appear continuously on the market and consumers do not
414 always know the composition of the drugs they take. WBE can shed some light and provide
415 additional information, but it is also affected by important challenges. First, pharmacokinetic data
416 are essentially non-existent for most NPS, making it extremely difficult to define appropriate
417 biomarkers. Second, the prevalence of abuse of a single substance is generally low, leading to very
418 low concentrations in wastewater. Finally, their stability in this matrix is largely unknown
419 (EMCDDA, 2016; Reid and Thomas, 2016). Based on the limited information available, this
420 section attempts to present a selection of potential biomarkers, to be used in WBE studies, for the
421 most common classes of NPSs: synthetic cannabinoids, synthetic cathinones, phenethylamines,
422 piperazines, tryptamines, arylcycloalkylamines and benzodiazepines (EMCDDA, 2015a). The two
423 first groups constitute the largest categories and also account for the majority of seizures in Europe
424 (EMCDDA, 2015a).

425 Synthetic cannabinoids include a broad range of structurally different compounds sharing
426 affinity for the cannabinoid receptors in the brain (Pertwee, 2008). Due to their recent increased
427 popularity, their human metabolism is a growing area of research. Several *in vitro* and *in vivo*
428 experiments have been performed over the past few years and, although individual pharmacokinetic
429 profiles remain to be elucidated for many of them, it is generally thought that synthetic
430 cannabinoids are extensively oxidized in the human body and excreted as a complex mixture of
431 phase I and phase II metabolites (Fantegrossi et al., 2014; Seely et al., 2012). JWH-type
432 cannabinoids are the most popular drugs within this class. Monohydroxylation, either at the N-alkyl
433 side chain, the naphthyl moiety or the indole moiety (followed by the corresponding
434 glucuronidation) has been identified as their major metabolic pathway and, in fact,
435 monohydroxylated metabolites have been detected in urine from JWH-type cannabinoids
436 consumers (Hutter et al., 2012; Ozturk et al., 2015; Wohlfarth et al., 2013). However, the lack of
437 rigorous pharmacokinetic data, essential to calculate excretion rates, prevents from extrapolating
438 these analyses to whole communities by the WBE approach. Another important limitation concerns
439 their instability in wastewater: the scarce literature available suggests that some synthetic
440 cannabinoids and their metabolites are highly labile and tend to get adsorbed to particle matter,
441 hindering their determination and sub-estimating the potentially derived abuse calculations (Reid et
442 al., 2014a, 2014b). As a reflection of these intrinsic difficulties, to the best of our knowledge only
443 the metabolite JWH 018 N-5-hydroxypentyl and the parent compounds JWH-210 and JWH-122,
444 have been positively detected in wastewater in two out of all the studies dealing with NPS in this
445 matrix (Borova et al., 2015; Reid et al., 2014b) (see **Table S1**).

446 Synthetic cathinones are known to have been abused for approximately 15 years and the
447 synthesis of cathinone derivatives has been reported since the late 1920s (Hyde and Adams, 1928;
448 Prosser and Nelson, 2012). They all refer to cathinone ((S)-2-amino-1-phenyl-1-propanone), a
449 naturally occurring stimulant found in the leaves of *Catha edulis* (Khat) (Prosser and Nelson, 2012).

450 In general, the drugs are in part extensively metabolized in humans. However, some of the synthetic
451 cathinones are also excreted unchanged in urine (Uralets et al., 2014). Details on the metabolism
452 and detectability of synthetic cathinones can be found in original articles and are summarized in
453 several review articles (Ellefsen et al., 2015; Helfer et al., 2007; Meyer et al., 2014, 2012, 2010a,
454 2010b; Meyer and Maurer, 2010; Pawlik et al., 2012; Pozo et al., 2014; Shima et al., 2014; Staack
455 and Maurer, 2005; Uralets et al., 2014; Welter-Luedeke and Maurer, 2015). Also, data on the
456 stability, especially under storage conditions, were published (Senta et al., 2015b) and highlighted
457 the possible instability of the parent compounds under alkaline conditions (Johnson and Botch-
458 Jones, 2013; Tsujikawa et al., 2012). However, detailed and comprehensive studies are missing on
459 their chemical stability in wastewater and also biotransformation in the sewer or wastewater should
460 be considered (McCall et al., 2016a). Several studies were published on the analysis of synthetic
461 cathinones in wastewater samples, with mephedrone, methylenedioxypropylone, methcathinone,
462 methylone and α -pyrrolidinovalerophenone (α -PVP) being the most frequently detected (Borova et
463 al., 2015; Chen et al., 2013; González-Mariño et al., 2016a, 2016b; Kinyua et al., 2015;
464 Mwenesongole et al., 2013; Ocaña-González et al., 2015; Thai et al., 2016; Tschärke et al., 2016).

465 Phenylethylamines are a class of substances related to amphetamine and methamphetamine,
466 possessing psychoactive and stimulant effects; however, modification of these compounds can lead
467 to potent hallucinogens (Zaitsev et al., 2011; Zawilska and Andrzejczak, 2015). They include
468 amphetamine derivatives such as MDMA, 2C and 'D' series drugs. However, the phenethylamine
469 core is shared among several compounds including cathinones and catecholamines. Several
470 metabolism studies have been conducted in an effort to understand their metabolic profiles (Ewald
471 et al., 2008, 2006; Lai et al., 2015b; Staack et al., 2003) but more information is needed.

472 Piperazine-like compounds include the original member 1-benzylpiperazine (BZP), its
473 methylenedioxy analogue and several phenylpiperazines. They are mainly known to bind to
474 serotonin receptors, with BZP additionally producing amphetamine-like stimulant effects (Bye et

475 al., 1973; De Boer et al., 2001). A summary with details on the metabolism of piperazines can be
476 found in some articles (Maurer et al., 2004; Staack et al., 2001; Staack and Maurer, 2005);
477 furthermore, one study showed the detection of metabolites in human urine (Tsutsumi et al., 2005).
478 Some examples are shown in **Table S1**.

479 Tryptamine is a primary amine alkaloid found widely in nature in both the plant and animal
480 kingdoms and known for its hallucinogenic effects (Collins, 2011). Metabolism of some synthetic
481 tryptamines has been studied (Kamata et al., 2006; Michely et al., 2015; Narimatsu et al., 2008).

482 Arylcycloalkylamines, which include the ketamine derivative methoxetamine (MXE) and
483 phencyclidine derivatives, have emerged as legal alternatives to ketamine (Roth et al., 2013). MXE,
484 which has gained popularity in several European countries (EMCDDA, 2014), is extensively
485 metabolized (Meyer et al., 2013) but it was detected as parent MXE in wastewater from Belgium
486 and Switzerland (Kinyua et al., 2015).

487 Benzodiazepines are psychoactive substances whose core structure is a benzene ring fused
488 to a diazepine ring. Benzodiazepines are known as tranquilizers and are among the most commonly
489 prescribed antidepressant medications. Although a useful pharmaceutical, there is potential for
490 abuse due to their hypnotic and sedative effects – even to the extent of being used as “date rape”
491 drugs (Schwartz et al., 2000). From now on we will refer to those benzodiazepines used illegally as
492 design benzodiazepines. Designer benzodiazepines have become a rapidly growing class of drugs
493 on the NPS online market, since a medical prescription is not needed. Since designer
494 benzodiazepines have increased in popularity, studies have been conducted characterizing their
495 human metabolism (Huppertz et al., 2015; Moosmann et al., 2013).

496 Up to now, no designer phenethylamines, tryptamines or designer benzodiazepines and
497 metabolites have been detected in wastewater and only two studies has reported the stability of
498 some phenylethylamines in wastewater (Bade et al., 2016; Senta et al., 2015b).

499 Although the interpretation of quantitative results should be done carefully for NPS due to
500 the lack of metabolic information, the qualitative monitoring could lead to a better understanding of
501 the frequency of use and could identify changes in consumption.

502

503 **3. EXPOSURE BIOMARKERS FROM ENVIRONMENT AND FOOD**

504 Two important exposure pathways for potentially harmful compounds are the dietary intake
505 and the exposure from the surrounding daily environment. The monitoring of various classes of
506 compounds for which exposure commonly occurs through these routes is necessary to safeguard
507 public health. Representative chemical classes have been chosen as examples for this paper.
508 Pesticides, mycotoxins and parabens are three classes of compounds for which exposure occurs
509 through the intake of contaminated food or absorption through the skin and adverse health effects
510 can be foreseen for humans (Błędzka et al., 2014; Heyndrickx et al., 2015; Rizzati et al., 2016;
511 Warth et al., 2013). Exposure through the indoor environment (furniture, electronics, packaging and
512 personal care products (PCPs)) is characteristic for UV-filters, plasticizers and brominated flame
513 retardants.

514 This section reviews the specific biomarkers of each of the above mentioned chemical
515 classes which could be measured in wastewater in order to assess the overall exposure to these
516 compounds through a WBE approach. When relevant, we have also included the metabolites of
517 these chemicals to be explored as a suitable biomarker. The suggested biomarkers are reported in
518 **Table S2** including also metabolites, whenever such information is available.

519

520 **3.1 Pesticides**

521 Pesticides are chemicals commonly used for control of harmful organisms, such as fungi,
522 insects and weeds. They are mostly used for crop protection, but can also be used for livestock
523 protection, as well as for other industrial and household purposes, such as termite prevention. The

524 general population is exposed to pesticides mainly through diet (Ntzani et al., 2013), but also
525 through household use (Trunnelle et al., 2013) and inhalation of polluted air - particularly in
526 agricultural areas where aerial spraying of pesticides occurs (Coscollà et al., 2010). Exposure to
527 pesticides is of public concern as they may cause health effects such as elevated rates of chronic
528 diseases, like cancer or diabetes, as well as neurodegenerative disorders such as Parkinson disease,
529 birth defects and reproductive diseases (Rizzati et al., 2016). Young children are the most
530 susceptible to be at risk (European Food Safety Authority, 2013).

531 There are several types of pesticides and they are generally classified by their chemical
532 structure: carbamate, organophosphate or triazine pesticides (**Table S2**). They may also be
533 classified by the type of pest they control, such as herbicides, which are intended to kill weeds and
534 other unwanted plants, and insecticides, which kill insects and other arthropods. Pesticides are
535 mostly formulated as mixtures with individual components which may act independently of each
536 other, interact or have dose-addition effects (Hernández et al., 2013).

537 Until now, there are only two WBE studies (Rousis et al., 2016a, 2016b) published on
538 human exposure to pesticides. The first work (Rousis et al., 2016a) proposed for the first time a new
539 application for pesticides, where pyrethroid, triazine and organophosphate metabolites were
540 monitored in influent wastewater of seven Italian cities. The most frequently detected compounds
541 were the specific metabolite of chlorpyrifos and chlorpyrifos-methyl, 3,5,6-trichloro-2-pyridinol
542 (TCPY), the metabolite of diazinon (2-isopropyl-6-methyl-4-pyrimidinol, IMPY), the pyrethroid
543 metabolites 3-phenoxybenzoic acid (3-PBA, common metabolite of about 20 pyrethroids), 3-(2,2-
544 dichlorovinyl)-2,2-dimethyl-(1-cyclopropane)carboxylic acid (DCCA, common metabolite of
545 permethrin, cypermethrin and cyfluthrin) and two alkyl phosphate metabolites. The second work
546 (Rousis et al., 2016b) applied the novel WBE approach to assess further exposure to pyrethroids,
547 concretely 3-PBA, cis-DCCA and trans-DCCA. The obtained results were in agreement with the

548 Human Biomonitoring (HBM) profiles in urine samples of the general population, reported in the
549 literature.

550 Yusa et al. 2015 reviewed analytical methods for HBM of pesticides and found that the most
551 commonly biomonitored ones are carbamates, herbicides, neonicotinoids, organophosphates,
552 pyrethroids and sulfonylurea herbicides – all of which can be monitored in urine samples and they
553 can be good potential biomarkers for WBE. However, some other pesticide classes, such as
554 organochlorines, are probably not suited to WBE due to their non-polar characteristics and their
555 poor excretion in urine (Yusa et al., 2015).

556 As described previously for other substances, the metabolites of pesticides rather than the
557 parent substances should be measured in wastewater to avoid contributions from sources other than
558 human metabolism. It has to be emphasized that some pesticide metabolites are also formed in the
559 environment (i.e. atrazine undergoes dealkylation in water systems forming human metabolites) and
560 therefore more research is needed. Moreover, there are some common metabolites produced by
561 different classes of compounds, such as organophosphate pesticides, organophosphate plasticizers
562 and flame retardants, and this should be taken into account in a WBE approach. The novel method
563 developed by Rousis et al. is considered as a valuable tool for obtaining objective, direct
564 information on pesticide exposure levels and could provide complementary information for HBM
565 studies. **Table S2** presents the main potential biomarkers of exposure to pesticides selected by
566 considering the detection frequency in urine, and the concentration levels (Barr, 2008; Yusa et al.,
567 2015).

568

569 **3.2 Mycotoxins**

570 Mycotoxins are toxic fungal metabolites that can be found in food and feed which are
571 intended for human and animal consumption (i.e. cereals such as rice, maize and wheat). There is

572 huge concern of human health risks related to the ingestion of these substances, since they are stable
573 in food processing and cooking. Maximum tolerable levels in food commodities were therefore
574 legally established in many countries (*Comission Regulation 1881/2006*, 2006). While, nowadays,
575 approximately 400 compounds belong to this group, only 10-15 are considered to be priority
576 mycotoxins, due to higher occurrence and toxicity. These latter compounds belong to the groups of
577 aflatoxins, ochratoxins, patulin and fusarium toxins (tricothecenes, fumonisins, zearalenone and
578 zearalenone derivatives) (Anfossi et al., 2016; Turner et al., 2015). HBM studies performed on
579 general population have shown that the most studied mycotoxin biomarkers in urine samples are
580 aflatoxin M1 (AFM1), ochratoxin A (OTA), deoxynivalenol (DON), nivalenol (NIV), fumonisin B1
581 (FB1) and zearalenone (ZON) (H Fromme et al., 2016; Heyndrickx et al., 2015). If mycotoxin
582 contaminations are going to be increased in the near future due to higher global food demand and
583 global climate and environment changes, new methods are needed to evaluate the human exposure
584 to mycotoxins (Marroquín-Cardona et al., 2014). Thus, a novel approach such as the WBE can be
585 useful to provide complementary information to existing methods.

586 Few studies dealing with the determination of mycotoxins in wastewater have been
587 published. The studied analytes were detected at very low concentrations (few ng/L), but at high
588 detection frequency. In addition to parent compounds, some human metabolites were also
589 investigated. The detected mycotoxins were DON, beauvericin (BEA), 3-Acetyldeoxynivalenol (3-
590 AcDON), NIV, ZON, α -zearalenol (α -ZOL) and β -zearalenol (β -ZOL) (Kolpin et al., 2014; Laganà
591 et al., 2004; Schenzel et al., 2012, 2010; Wettstein and Bucheli, 2010). None of these studies
592 attempted to apply the WBE approach to these substances; they had only a monitoring scope. In the
593 present paper a selection of mycotoxins and their related potential biomarkers for a WBE approach
594 were reported for the first time (**Table S2**).

595

596 **3.3 Parabens**

597 Parabens are a group of chemicals that is drawing a lot of interest in the current discussion
598 given their potential endocrine disrupting properties, since studies have shown that they have
599 potential adverse health effects (Hu et al., 2013; Kim et al., 2015; Zhang et al., 2013). This has
600 raised concern considering their widespread use. Parabens are used as preservatives in many
601 different products, such as cosmetics, PCPs and foods, and can be commonly found in household
602 products.

603 Some studies also investigated the occurrence and fate of parabens in wastewater (González-
604 Mariño et al., 2009; Gracia-Lor et al., 2012a; Kasprzyk-Hordern et al., 2008), but not from a WBE
605 perspective. Therefore, a list of known urinary biomarkers for paraben exposure is reported in
606 **Table S2**. Future research should be addressed in order to explore paraben biomarkers for WBE.

607

608 **3.4. UV-Filters**

609 Overexposure to ultraviolet (UV) radiation has been associated with skin disorders, such as
610 cancer (Ramos et al., 2016). This led to the widespread usage of UV filters in a variety of personal
611 care products to protect against UV radiation, i.e., sunscreen, cosmetics, beauty creams, body
612 lotions, hair sprays and shampoos (Brausch and Rand, 2011). UV filters are also used in food
613 packages, plastics and textiles to prevent polymer degradation. Hence, human exposure occurs
614 through multiple routes such as dermal absorption, ingestion of contaminated food and tap water
615 (Valle-Sistac et al., 2016). Two major types of UV filters are currently available; organic UV filters
616 are used to absorb UVA and/or UVB radiation, whereas inorganic UV filters mainly reflect the
617 radiation. Given the high photostability and lipophilicity, many UV filters can enter biological
618 membranes and bioaccumulate in the body, including in the placental tissues (Valle-Sistac et al.,
619 2016). However, it is important to note that most UV-Filters are released into the sewers without
620 going through the body (Daughton and Ruhoy, 2009; Ruhoy and Daughton, 2008). This fact would
621 contribute to a large uncertainty in its estimation.

622 Urinary analysis has frequently detected UV filters at various levels, demonstrating human
623 exposure (Dewalque et al., 2014; Louis et al., 2015). Despite their widespread use, between 2010
624 and 2015 only 20 studies have been published in peer reviewed journals dealing with UV filters
625 detection in wastewater (Ramos et al., 2016). Yet, available data indicates that major UV filters
626 groups, i.e. benzophenone derivatives, p-aminobenzoic acid derivatives, camphor derivatives,
627 benzotriazole derivatives, salicylate derivatives, benzimidazole derivatives, triazine derivatives,
628 cinnamate derivatives, crylene derivatives, and dibenzoyl methane derivatives, are ubiquitous in
629 wastewater with concentrations ranging from the ng/L to the mg/L level (Gago-Ferrero et al., 2011;
630 Rodil et al., 2012). Evidence from mammalian studies indicate that various UV filters are endocrine
631 disruptors, acting as estrogenic, antiestrogenic, antiandrogenic or antithyroid (Louis et al., 2014).
632 These results find support in recent epidemiologic studies reporting an association between human
633 urinary levels of certain UV filters and couples fecundity, i.e. BP-2 (Louis et al., 2014), and
634 decrease semen quality, i.e. BP-3 and BP-8. Therefore, (Louis et al., 2015) highlighted the
635 importance of further studies exploring human exposure to UV filters. Despite the presence of UV
636 filters has been reported in wastewater (Ramos et al., 2016; Tsui et al., 2014) no WBE approaches
637 have been yet tested to evaluate human exposure to these substances. However, the high stability of
638 these compounds and the indication of particular metabolite signatures (Le Fol et al., 2015) suggest
639 potential biomarkers for UV filters in wastewater based biomarkers to support epidemiological
640 studies (**Table 1 and S2**).

641

642 **3.5. Plasticizers**

643 Plastics are very versatile materials typically consisting of organic polymers of high
644 molecular mass, which may contain other substances. Manufacturers often add different chemicals
645 to plastics to give them specific characteristics, such as flexibility, resilience and pliability. These
646 plasticizers mainly include phthalates and adipates, and because of their environmental persistence

647 and their widespread use, it is unsurprising that they can be found in wastewater and in the
648 receiving environment (Barnabé et al., 2008; Gao and Wen, 2016; Olofsson et al., 2013; Zolfaghari
649 et al., 2014). Some of these chemicals and/or their derivatives interfere with endogenous hormone
650 signalization in animals and humans, raising concerns about their potential to cause long-term
651 diseases (Joint Fao Oms Expert Committee On Food Additives, 2010). In particular phthalates (e.g.
652 bis(2-ethylhexyl) phthalate and, dibutyl phthalate) were associated with the disruption of
653 hormonally-mediated pathways, as well as increased risk for cancer (“Toxicological profile for
654 di(2-ethylhexyl)phthalate (DEHP),” 2002, “Toxicological profile for Di-n-butyl-Phthalate,” 2001).
655 Furthermore, epidemiological observational studies suggest that there is a consistent association of
656 blood and urine concentrations of phthalates, and some effects, such as those mentioned above
657 (Joint Fao Oms Expert Committee On Food Additives, 2010; Kim et al., 2015; Wang et al., 2016).
658 Due to a better toxicological profile (Bhat et al., 2014) and a better blood compatibility (Zhong et
659 al., 2013), other plasticizers, such as di-isononyl cyclohexane-1,2-dicarboxylate (DINCH), have
660 been increasingly used in recent years as alternatives in PVC films and medical devices.
661 Metabolites of phthalates, adipates, and DINCH have been found in urine (Fromme et al., 2016;
662 Guo et al., 2011; Herrero et al., 2015; Loftus et al., 1993; Silva et al., 2007), but their presence in
663 wastewater has never been investigated. For a list of known biomarkers in urine see **Table S2**.

664

665 **3.6 Flame retardants**

666 Flame retardants (FRs) are chemical additives for manufactured materials, such as plastics
667 and textiles, to inhibit, suppress, or delay the production of flames to prevent the spread of fire.
668 Brominated flame retardants (BFRs) and organophosphorus flame retardants (PFRs) are the most
669 used classes of organic FRs. Due to their high log K_{ow} , BFRs are lipophilic and preferentially
670 retained in the human body, e.g. in the blood or adipose tissue. They are only slowly metabolized to
671 hydroxylated metabolites (e.g. HO-PBDEs), which are also retained in the body and thus not

672 excreted in the urine. The presence of BFRs in the sewer system is largely due to direct input from
673 the indoor environment, following washing out of dust and being associated with particles. PFRs
674 are less persistent and rapidly metabolized in the human body (Van den Eede et al., 2013), they
675 have been measured in municipal wastewater in Europe (Loos et al., 2012; Marklund et al., 2005),
676 Australia (O'Brien et al., 2014) and United States (Schreder and La Guardia, 2014). PFRs
677 metabolites are excreted via urine and they are thus suitable biomarkers to assess human exposure
678 to PFRs (Van den Eede et al., 2015); however, there are no reports on the presence of PFR
679 metabolites in wastewater and no studies testing them in a WBE approach (**Table S2**).

680

681 **4. HEALTH BIOMARKERS**

682 Community health programs play an essential role for public health agencies to monitor and
683 evaluate the present status of health in a community and measure the success of programs aimed at
684 improving it. Current challenges mainly consist of the quick and reliable evaluation of the overall
685 health of a population, and detect possible health and illness threats such as pandemics or higher
686 prevalence of diabetes or cancer.

687 The quantitative measurement of specific exogenous and endogenous biomarkers related to
688 these diseases in wastewater has the potential to provide rapid information on different factors
689 related to public health and illness. Specific classes of pharmaceuticals such as antibiotics and
690 benzodiazepines and their metabolites are exogenous compounds, which can be related to their use
691 for specific illnesses or diseases, whereas endogenous compounds, such as α -fetoprotein,
692 chorionic gonadotropin (hCG) and isoprostanes, are more directly related to cancer or stress.

693 In this section, both exogenous and endogenous specific biomarkers are presented and
694 suggested to monitor health issues (**Table S3**) through the WBE approach. In addition, DNA-based
695 approaches, currently applied in the field of WBE, have been reviewed.

696

697 **4.1 Pharmaceuticals**

698 **4.1.1 Antibiotics**

699 Antibiotics (ABs) can be suitable biomarkers for representing human health status
700 associated with bacterial infections. The determination of reliable data on their consumption is of
701 interest as AB use is one of the main factors responsible for AB resistance (Euro-CDC, 2012). WBE
702 may give a better understanding of real time use and misuse of ABs at the population level, by
703 supporting for example prescription data from official sources and annual sales.

704 Many ABs are excreted unchanged in urine (Castiglioni et al., 2006; Huang et al., 2011),
705 hence, parent drugs are generally targeted as biomarkers (**Table S3**). However, the selection of a
706 significant AB biomarker should not be limited to the parent drug only; in fact, the investigation of
707 specific metabolites is adding specificity to the analysis avoiding biases coming from the direct
708 disposal of the AB. This is particularly relevant for ABs widely used for veterinary treatments. The
709 most targeted classes of ABs are β -lactams, quinolones and fluoroquinolones, sulphonamides,
710 tetracyclines and macrolides. Apart from β -lactams that undergo easy hydrolysis, sulphonamides
711 and macrolides are very persistent, and are therefore also detected in treated wastewater (Jelic et al.,
712 2012). Stability of the ABs metabolites in wastewater is less understood.

713 The occurrence of ABs in influent wastewater has been widely investigated in several
714 countries (Gracia-Lor et al., 2012b; Kümmerer, 2009; Verlicchi et al., 2012). Seasonal variability of
715 population-normalized mass loads was observed by Castiglioni et al. 2006, using the WBE
716 approach, showing a difference in percentage from winter to summer of 47, 77 and 100 for
717 ciprofloxacin, ofloxacin and sulphamethoxazole, respectively (Castiglioni et al., 2006). Temporal
718 monitoring of ABs at several time scales showed a higher variability monthly/hourly than
719 daily/weekly along with seasonality in mass fluxes for ciprofloxacin, ofloxacin and clindamycin
720 (Coutu et al., 2013). Deconjugation during in-sewer transport may influence the influent loading of
721 sulfamethoxazole (Snip et al., 2016) depending on the type and size of the served catchment

722 (Polesel et al., 2016). Application of WBE helped in determining the usage of ABs in areas where
723 consumption data were scarce or a proper regulation was missing, revealing an excessive use in
724 China (Yuan et al., 2015).

725

726 **4.1.2 Benzodiazepines**

727 Benzodiazepines are used therapeutically for a considerable number of applications,
728 including anxiety and sleep disorders. Their primary mode of action is an enhancement of the action
729 of the neurotransmitter gamma-aminobutyric acid which may result in anticonvulsant, anxiolytic,
730 hypnotic, muscle relaxant and sedative effects. Benzodiazepines and benzodiazepine analogs are
731 commonly prescribed; however, they are also among the most frequently abused prescription
732 medications (Button, 2015). Despite the risk for abuse, approximately 5.2% of US adults between
733 18 and 80 years of age used benzodiazepines in 2008, with a double prevalence for women than
734 men (Olfson et al., 2015). As such, monitoring of benzodiazepines is of public concern.

735 Monitoring benzodiazepines in populations could be achievable via WBE as they are
736 normally halogenated and hence resistant to biodegradation (Kosjek et al., 2012). Multiple studies
737 have already identified both parent benzodiazepines and their urinary metabolites in wastewater
738 influent (Baker et al., 2014; Borova et al., 2014; Castrignanò et al., 2016; Fernández et al., 2014;
739 Hummel et al., 2006; Kosjek et al., 2012; Racamonde et al., 2015, 2014). Differences in the
740 behavior of benzodiazepines are associated with differences in functional substituent groups, and
741 mainly the hydroxylated tranquilizers, oxazepam, and temazepam, were reported to be present in
742 influent and effluent wastewater (Bijlsma et al., 2012; Hummel et al., 2006; Löffler et al., 2005).

743 A summary of the most commonly prescribed and detected benzodiazepine parent
744 compounds and their metabolites, which have been identified in urine, in addition to identification
745 in wastewater and stability data, when available, are presented in **Table S3**.

746

747 **4.1.3 Other pharmaceuticals**

748 Even if many works have analysed the presence of pharmaceuticals in urban wastewater,
749 only a few studies investigated these chemicals as WBE biomarkers. Some examples can be found
750 in **Table 1**. Furthermore, a list of proposed pharmaceuticals is given in **Table S3** with their
751 excretion rates.

752

753 **4.1.4. Chiral pharmaceuticals**

754 More than 50% of pharmaceuticals currently used are chiral although they are usually
755 manufactured as racemic mixtures (Petrie et al., 2015; Vazquez-Roig et al., 2014). Human
756 metabolism and microbial processes during wastewater treatment can result in the enrichment of
757 one specific enantiomer. Thus, the analysis of chiral compounds in wastewater allows to distinguish
758 between usage of pharmaceuticals due to intentional human ingestion and from accidental release
759 (direct disposal). For instance, enantioselective analysis was used by (Vazquez-Roig et al., 2014) to
760 tentatively propose direct disposal of atenolol where a moderate higher average daily load was
761 observed. Recently, (Petrie et al., 2016) identified direct disposal of the antidepressant fluoxetine
762 via the sewer network using wastewater analysis.

763

764 **4.2. Endogenous compounds**

765 Endogenous chemicals are produced by biological processes associated with stress or
766 normal metabolism. Changes in biological mechanisms may result in alterations of the endogenous
767 compound production and, therefore, measurement of such compounds can be used as indicator of
768 health status and disease (Daughton, 2012b; Group, 2001; Hagger et al., 2006). Endogenous
769 biomarker analysis has been extensively studied as diagnostic or prognostic tools in clinical
770 medicine, and can be further applied to the field of WBE (Daughton, 2012b). Thus far, the

771 investigation of endogenous biomarkers has been more focused on diseases such as cancer, diabetes
772 and cardiovascular disorder than on the overall health status. However, the number of biomarkers
773 validated for routine clinical practice is rather limited (Poste, 2011; Rifai et al., 2006), which falls
774 into even smaller numbers of biomarkers for WBE when considering only those excreted into urine.
775 Nevertheless, a range of endogenous compounds have been suggested as wastewater biomarkers of
776 effect including cancer (prostate specific antigen, α -fetoprotein) (Thomas and Reid, 2011; Yang et
777 al., 2015c), oxidative stress (isoprostanes) (Daughton, 2012b; Ryu et al., 2015; Thomas and Reid,
778 2011) and health (anti-inflammatory eicosanoids) (Daughton, 2012b). To date, studies conducted on
779 candidate endogenous biomarkers in wastewater are based on targeted analysis of specific markers
780 such as isoprostanes (Ryu et al., 2015) and cancer biomarkers (Yang et al., 2015c). However, it is
781 important to note that omics approaches also hold promising and important roles in future
782 developments and applications of endogenous biomarkers analysis in WBE (Rice et al., 2015). The
783 added value of analyzing these compounds would reside mainly in relative comparisons, both intra-
784 and inter- communities (Daughton, 2012b). Compared to the interpretation of the exogenous
785 biomarkers, where absolute values are emphasized, the use of endogenous biomarkers is more
786 focused on detecting changes over time or between communities. Such data can reveal emerging
787 trends (i.e., early warning system) and health disparities caused by various factors (e.g., exposure,
788 lifestyle).

789

790 **4.3. DNA**

791 The demand for sensitive, low-cost and high-throughput methods to characterize DNA/RNA
792 sequences has driven the development of molecular biology techniques and bioinformatics, i.e.,
793 PCR-based approaches and next generation sequencing (NGS) (Ryoo et al., 2013). Massive
794 sequencing is nowadays possible, owing to the development of different NGS platform that allows
795 an entire genome to be sequenced in less than one week. These technical advances led to a rapid

796 increase in new applications, including DNA-based health biomarkers. During the last decade an
797 increasing number of studies took advantage of these developments, and applied them to the field of
798 WBE. Several examples highlight the potential of the approach. In the field of virological
799 surveillance, wastewater screening has been used to identify the viral strains that are circulating in
800 the community, supporting epidemiological studies of the related viral infections and working as an
801 early warning tool (Hellmér et al., 2014; Kokkinos et al., 2011; McLellan et al., 2013; Zhou et al.,
802 2014). Hellmér et al. 2014 investigated the presence of eight pathogenic viruses (norovirus,
803 astrovirus, rotavirus, adenovirus, Aichi virus, parechovirus, hepatitis A virus [HAV], and hepatitis E
804 virus) in wastewater from Sweden to explore whether their identification could be used as an early
805 warning of outbreaks. Results show that two strains were involved in an ongoing outbreak in
806 Scandinavia and were also identified in samples from patients with acute hepatitis A in Gothenburg
807 during spring of 2013.

808 A similar framework has been applied in other areas such as the study of the epidemiology
809 of the emerging human pathogens (McLellan et al., 2013; Webb et al., 2015), and antibiotic
810 resistance patterns of populations (Colomer-Lluch et al., 2014; Kumaraswamy et al., 2014;
811 McLellan and Eren, 2014). One of the most recent applications has been in the field of human
812 metabolic disorders. With the obesity epidemic reaching alarming levels, there is a need to set
813 biomarkers to identify populations or sub-populations at risk (Lyssimachou et al., 2015). Recently,
814 a good correlation has been established between the gut microbiome and obesity. In fact, only a few
815 bacterial species are sufficient to distinguish between lean and obese individuals (Le Chatelier et al.,
816 2013). These findings prompted a large study in the US using oligotyping of high-throughput 16S
817 rRNA gene sequence data to screen wastewater from 71 cities. It was demonstrated that cities could
818 be differentiated by their sewage bacterial communities, and the community structures were good
819 predictors of a city's estimated level of obesity (Newton et al., 2015). This example illustrates that

820 once specific biomarkers are identified, DNA-based analysis in wastewater can work as a powerful
821 tool to support epidemiological studies

822

823 **5. POPULATION BIOMARKERS**

824 Accurate estimation of population size is necessary to normalize WBE data to the per capita
825 level, which allows for temporal and spatial comparisons to be made (van Nuijs et al., 2011b). A
826 review of all uncertainties associated with WBE found that there is a direct relationship between the
827 uncertainty in measuring the population size and the uncertainty in the calculated daily loads of
828 drugs (Castiglioni et al., 2013; Lai et al., 2015a). Therefore, accurate data on population size are
829 needed to make decisions involved with planning and forecasting, assessing services and
830 infrastructure, policy making, informing legislation and resource allocation at the level of
831 neighborhood, city, province or country.

832 Current methodologies to estimate population size are based on public surveys (such as
833 census taking), complemented with a wide array of demographic statistics, such as tourism and
834 potential commuters. Census, however, can become increasingly outdated and cannot be easily
835 updated to accommodate change such as births, deaths, and migration (movement). Ideally, the
836 census should be able to estimate both the *de jure* and the *de facto* population. The *de jure*
837 population comprises all “usual” residents, mainly those with formal residences. The *de facto*
838 population comprises all those who are present, regardless of the location of their formal or usual
839 residence (Daughton, 2012a). A *de facto* population therefore includes all non-residents (e.g.,
840 commuters, visitors, tourists) and excludes all permanent residents who are absent. However, the
841 census approach acquires a static snapshot estimate and usually succeeds in only capturing a portion
842 of the population. Population size can also be estimated from hydrochemical parameters that are
843 routinely determined in the WWTPs, including chemical oxygen demand (COD), biological oxygen

844 demand (BOD) and total nitrogen and phosphorus. However, these parameters are highly influenced
845 by wastewater composition (i.e. industrial, domestic or mixed).

846 Addressing the population uncertainty and identifying suitable markers for the population
847 size markers is thus an important aspect of WBE (Been et al., 2014; Brewer et al., 2012; Lai et al.,
848 2011; O'Brien et al., 2014). Many compounds can be considered as biomarkers for population size.
849 Possible candidates are both naturally occurring and synthetic xenobiotics (and their metabolites or
850 formulation impurities), as well as products of endogenous metabolism. A variety of chemicals
851 have been studied as biomarkers of population, including drugs (e.g., carbamazepine (Gasser et al.,
852 2010)), biocides (e.g., triclosan (Singh et al., 2010)), chemicals in household cleaning agents, e.g.,
853 fluorescent whiteners, trialkylamines (Managaki et al., 2006; Valls et al., 1989), and food additives,
854 e.g., sucralose (Oppenheimer et al., 2011). An essential characteristic for a biomarker to be useful
855 for measuring population size is, in addition to the general requirements for a biomarker, to have a
856 low variance in the per capita daily excretion (Daughton, 2012a); the knowledge of quantities
857 excreted daily ensures that diurnal variations (e.g., resulting from circadian biorhythms) are fully
858 accommodated. Another requisite for these groups of biomarkers is that daily per capita excretion
859 should not be affected by variables such as season, weather and geographic location.

860 To date, none of the population size markers proposed have yet met all necessary criteria
861 mentioned above and additional characteristics described before for a WBE biomarker should also
862 be considered. Some specific applications are listed below.

863

864 **5.1 Artificial sweeteners**

865 The most popular artificial sweeteners used in foodstuffs include acesulfame (ACE), alitame
866 (ALI), aspartame (ASP), cyclamate (CYC), neotame (NEO), neohesperidin dihydrochalcone
867 (NHDC), saccharin (SAC) and sucralose (SUC) (**Table S4**) (Kokotou et al., 2012; Lange et al.,
868 2012). All of them, except NEO and ALI, are allowed to be used as additives in food by the

869 European Union (EPCD, 2003), whereas five of them, ACE, ASP, NEO, SAC and SUC are
870 approved to be used in the United States (USFDA, 2006).

871 After ingestion, ACE, CYC and SAC are unaffected by the human metabolism, and thus
872 largely eliminated from human bodies mainly unchanged in urine (Fermin and Vallvey, 2004;
873 Lange et al., 2012; Renwick, 1985; Roberts et al., 2000; Sardesai and Waldshan, 1991). Studies
874 have shown that, due to variations in individual metabolism, CYC could be metabolized to
875 cyclohexylamine and excreted in urine (Renwick et al., 2004). For ALI, 7–22% is excreted
876 unchanged in feces, while the rest, about 78–93% is hydrolyzed to aspartic acid and alanine amide
877 (Fermin and Vallvey, 2004). The glucuronide conjugates of ALI metabolites are the major urinary
878 metabolites in the first 24 hours. ASP is largely broken down in human gut to aspartic acid,
879 phenylalanine and methanol (Fermin and Vallvey, 2004; Lange et al., 2012). NEO and its
880 metabolites are excreted in urine and feces (WHO Food Additive Series No. 52, 2004). Less than
881 2% is excreted unchanged, but it is extensively metabolized in humans via de-esterification to *N*-[*N*-
882 (3,3-dimethylbutyl)-*L*-*alpha*-aspartyl]-*L*-phenylalanine (WHO Food Additive Series No. 52, 2004).
883 Minor metabolites of NEO include *N*-(3,3-dimethylbutyl)-*L*-aspartic acid, 3,3-dimethylbutanoic
884 acid and the carnitine conjugate and glucuronide conjugate of 3,3-dimethylbutanoic acid (WHO
885 Food Additive Series No. 52, 2004). NHDC is hydrolyzed in humans to isoferulic acid, 3-
886 hydroxyphenylpropionic acid, and 3-hydroxycinnamic acid (Fermin and Vallvey, 2004; Lange et
887 al., 2012). SUC is mainly excreted unchanged in human feces, while 8-22% was excreted in urine
888 unchanged together with its glucuronide conjugates (Roberts et al., 2000).

889 ACE, CYC, SAC, and SUC were found highly stable in raw wastewater at 4°C and room
890 temperature over four days (Ordóñez et al., 2012). Under these conditions, only 20-30% of ASP
891 remained after one day and none left after two days. Similarly, the amount of NHDC was found less
892 than 10% in the raw wastewater at 4°C after one day and linearly decreased at room temperature
893 over three days. Similar results were also reported in another study, in which ACE, CYC, SAC and

894 SUC remained stable in raw wastewater at 4°C over three weeks, whereas ASP and NHDC were
895 degraded within a day (Tran et al., 2013).

896 Since they are exclusively non-metabolized in humans and highly stable in wastewater, the
897 parent compounds ACE, CYC, SAC and SUC can be measured for the WBE approach. However,
898 the analysis of the metabolites of ALI, ASP, NEO and NHDC, rather than of the parent compounds,
899 is required, since these artificial sweeteners are largely metabolized in humans. Stability tests for
900 the metabolites in raw wastewater are also necessary for future studies. The use of artificial
901 sweeteners has been shown to be highly related to human activities (Buerge et al., 2009) and,
902 therefore, human consumption is considered as the major source of these substances in raw
903 wastewater; however, other sources, such as animal feedings, agriculture farms and industries, can
904 contribute to their presence in sewage systems (Kokotou et al., 2012).

905 Certain artificial sweeteners also showed a specific weekly pattern: in general higher loads
906 in influents (i.e. consumption) were observed during weekdays than during weekends (Kokotou et
907 al., 2013). This could be associated with more commuters during the weekday than the weekend in
908 the studied catchment. These previous studies together suggested that measuring artificial
909 sweeteners could be useful for the WBE approach to understand the population flow in a given
910 catchment. This concept of using human consumed chemicals, such as the artificial sweetener ACE,
911 to back-estimate the population size from a given wastewater sample was firstly attempted and
912 discussed by (Lai et al., 2011) and further refined using wastewater samples collected on the census
913 day and applying a Bayesian model (O'Brien et al., 2014). Importantly, with chemical-derived
914 population estimates, the robustness of the WBE data was improved, since the total methodological
915 uncertainty of the approach was reduced (Lai et al., 2015a, 2011).

916

917 **5.2. Nicotine**

918 Currently, nicotine and its metabolites have been used as population markers on two
919 occasions (Chen et al., 2014; Senta et al., 2015a). In the first case, the authors focused solely on
920 cotinine, whose loads varied only limitedly over one week and showed good correlation with the
921 size of the investigated populations (i.e., correlation coefficient = 0.981) (Chen et al., 2014).
922 However, geographical/cultural differences in tobacco use or fluctuations in the number of users
923 have been raised as potential flaws to the use of cotinine as population marker (Chen et al., 2014).
924 Moreover, consumption of tobacco could change due to tax and other tobacco-related policies,
925 which could affect the potential of nicotine and its metabolites as population markers. In the second
926 study (Senta et al., 2015a), cotinine and *trans*-3'-hydroxycotinine loads were used to estimate the
927 number of individuals contributing to the collected wastewater samples. Good agreement was found
928 between nicotine metabolite load population estimates and census data, suggesting that the method
929 is a viable approach to estimate the size of a population.

930

931 **5.3. Caffeine**

932 Caffeine and some of its major metabolites were recently tested as a population biomarkers.
933 Caffeine was one of the compounds included in the exploratory study to estimate population size
934 using samples collected on the census day and applying a Bayesian model (O'Brien et al., 2014). A
935 strong correlation between caffeine mass loads and population size was observed. In the second
936 study, generally good agreement between caffeine loads and hydrochemical parameters routinely
937 determined at the WWTPs was found (Rico et al., 2016). In another recent study, three major
938 caffeine metabolites: 1-methylxanthine, 7-methylxanthine and paraxanthine were tested together
939 with caffeine as possible population biomarkers (Senta et al., 2015a). These compounds fulfilled
940 some of the major requirements for an ideal biomarker - they are easily detectable and stable in
941 wastewater samples. However, their mass loads in wastewater did not completely reflect the human
942 excretion profile of caffeine, probably due to biases in caffeine pharmacokinetic data (see section

943 2.4 and **Table S2**) and additional sources of some metabolites and unconsumed caffeine. This
944 makes the possibility of using caffeine and/or its metabolites as biomarkers for population size
945 assessment rather difficult, at least without additional studies.

946

947 **5.4. Pharmaceuticals**

948 Concentrations and mass loads of pharmaceuticals in wastewater were used in the WBE field
949 for the estimation of population size only on three occasions (Lai et al., 2011; O'Brien et al., 2014;
950 Rico et al., 2016). The investigated compounds by Lai et al. (Lai et al., 2011) were atenolol (beta-
951 blocker), gabapentin (anti-convulsant), hydrochlorothiazide (diuretic), and venlafaxine (anti-
952 depressant). Atenolol was concluded to be the best option for this aim for the specific catchment. In
953 addition to the compounds selected by Lai et al., the same group also investigated carbamazepine
954 (antiepileptic), codeine, ibuprofen, paracetamol (analgesics), furosemide (diuretic), iopromide
955 (contrast medium), naproxen (anti-inflammatory) and salicylic acid (metabolite of acetylsalicylic
956 acid) and the measured loads were used in a collective model for the estimation of the population
957 size (O'Brien et al., 2014). By cross validating the data, the authors demonstrated that large
958 populations sizes could be estimated fairly accurately using the information of multiple chemical
959 mass loads. However, it could not be improved for small populations. In the work published by
960 (Rico et al., 2016) twelve human urine biomarkers were tested to estimate population size, six of
961 them being pharmaceuticals (hydrochlorothiazide, carbamazepine, codeine, naproxen, salicylic acid
962 and atenolol). However, by using these compounds, the population was under or overestimated
963 compared to the hydrochemical population, but they have good prospects if the appropriate data
964 sales are available.

965

966 **5.5. Endogenous compounds**

967 An alternative for estimating the population size in the catchment area of a WWTP relies on
968 monitoring influent wastewater for a biomarker linked to human metabolism. Chemicals involved
969 in endogenous metabolism avoid many of the problems encountered with xenobiotics, since their
970 association with human activities has a higher fidelity. Yet, their main problem is excessive intra-
971 and inter-individual variation in excretion. Biomarkers of endogenous origin derive from human
972 biochemical processes and undergo continuous urinary or fecal excretion. Several endogenous
973 biomarkers, which have been considered in the past or which have the potential to estimate the
974 population size more accurately (**Table S4**), are further discussed.

975 An important endogenous biomarker, widely used in clinical chemistry and with detailed
976 knowledge about its excretion, is creatinine (CR). A small portion of creatine (and
977 phosphocreatine), which is stored predominately in skeletal muscle, is continually converted to
978 form the endogenous anhydride, CR (a nitrogenous waste product cleared via the kidney); the rate
979 of conversion, in males for example, is about 1.6–1.7% per day. The major factors involved with
980 variability in CR output have been summarized by (Ryan et al., 2011). However, intra- and inter-
981 day CR excretion is not constant and daily excreted quantities can have high variance, being
982 strongly influenced by diet composition. In addition, CR is being increasingly used as a food and
983 nutritional supplement, adding yet another source of potential variation to CR excretion rates.
984 Although CR has been used in WBE studies as population marker (Brewer et al., 2012; Chiaia et
985 al., 2008), it was shown to be unstable in wastewater (completely decomposed within 24 h) (Chen
986 et al., 2014).

987 Another potential biomarker is coprostanol (CoP) that originates from gut microbial
988 metabolism, making up roughly 60% of the overall sterol content in human feces. CoP is poorly
989 absorbed from the gut (it does not undergo enterohepatic circulation) and is therefore fully excreted
990 in the feces. Since the 2000s, CoP has been used as anthropogenic marker in wastewater and to
991 gauge the degree of dilution of raw or treated wastewater in receiving surface water (Takada and

992 Eganhouse, 1998). However, CoP is excreted by various vertebrates in differing absolute and
993 relative quantities and it is sometimes difficult to distinguish between human and animal
994 contamination (Bull et al., 2002). Furthermore, CoP adsorbs substantially onto particulate matter
995 found in wastewater and was thus discarded as potential population marker (Chen et al., 2014).
996 Similar results were obtained for cholesterol (Chen et al., 2014); cortisol and androstenedione were
997 investigated, but rapidly degraded in wastewater (Chen et al., 2014).

998 Another example of biomarker relatively unique to human metabolism is 1-aminopropan-2-
999 one (1-aminopropanone: APR; 1-aminoketone). Through 1-aminopropan-2-ol, APR serves as a
1000 precursor to vitamin B-12 (Fitzsimons and Belt, 2005). It is very water soluble and it is excreted via
1001 urine, but in much lower daily quantities than CoP. However, it is sometimes found in wastewater
1002 at levels higher than in urine, implicating potential *de novo* microbial formation in sewage
1003 (Fitzsimons and Belt, 2005), whilst it could not be detected on other occasions (Singh and
1004 Gardinali, 2006).

1005 5-hydroxyindoleacetic acid (5-HIAA), a metabolite of serotonin, has also been investigated.
1006 Its excretion might be altered due to diseases (e.g., carcinoid tumors (Zuetenhorst, 2004)) and diet
1007 (i.e., some fruits and nuts (Feldman and Lee, 1985) and salt intake (Sharma et al., 1993)).
1008 Furthermore, intra- and inter-individual variability in excretion has also been highlighted (Curtin et
1009 al., 1996). Results from wastewater analysis showed good correlation with census data and the
1010 authors considered it as a promising marker (Chen et al., 2014).

1011 Ammonium (NH_4^+) represents the major form in which ammonia (NH_3) is found in
1012 wastewater and originates from the breakdown of urea (Udert et al., 2006). It is mainly introduced
1013 via toilets (Butler et al., 1995) and it is routinely measured by WWTP as a water quality parameter.
1014 It is supposedly less affected by non-human sources compared to conventional parameters (e.g.,
1015 chemical or biological oxygen demand, total phosphorous) (van Nuijs et al., 2011b) and can
1016 potentially be measured online using ion-selective electrodes. Fluctuations in ammonium loads have

1017 been shown to link well to population dynamics (Been et al., 2014). Yet, its use to estimate absolute
1018 figures of the size of the *de facto* population might be undermined in rural areas due to the
1019 contribution of agricultural sources.

1020

1021 **5.6. DNA**

1022 Deoxyribonucleic acid (DNA) is a nucleic acid that carries most of the genetic instructions
1023 from all known living organisms and many viruses. DNA can be naturally shed into the
1024 environment through urine, feces, exudates or tissue residues. Compared to most of chemical
1025 compounds as a candidate of population biomarkers, DNA is much more stable and able to persist
1026 in the environment from month to hundred years depending on species (Prüfer et al., 2014;
1027 Thomsen and Willerslev, 2015). DNA biomarkers have been widely used in the field of medical
1028 diagnostics and biomedicine (Altintas and Tothill, 2013; Liu et al., 2011; Ralla et al., 2014; Wang et
1029 al., 2012). For WBE, DNA has a great potential to act as a population biomarker, not only because
1030 of its little affinity to other species in wastewater and constant excretion by humans, but also for its
1031 extreme stability and the possibility of being quantifiable Those robotic characteristics well meet
1032 the proposed criteria of a proper population biomarker candidate (Dejean et al., 2011; Thomsen and
1033 Willerslev, 2015).

1034 Typically, the changes of DNA component and structure such as DNA damage, repair and
1035 mutation could be used as biomarkers. Recently, a H2AX histone phosphorylation assay was
1036 developed as DNA damage biomarker for human population study, as it represents an early event in
1037 the cellular response against DNA double-strand breaks (Sánchez-Flores et al., 2015). However, to
1038 select a population biomarker for WBE uses, one of the crucial criteria is to screen human specific
1039 DNA. Wastewater is a complex matrix, which may contain DNA from various species such as
1040 plants, animals, and viruses. A recent study by Yang *et al* (Yang et al., 2015a, 2015b) has proposed
1041 to use community sewage sensors to identify human-specific mitochondrial DNA as a potential

1042 population biomarkers. In this study, human specific mitochondrial DNA associated with disease
1043 biomarkers (Liu et al., 2011; Tipirisetti et al., 2014) was amplified from wastewater by a
1044 specifically designed primer using quantitative real-time polymerase chain reaction (PCR) (Yang et
1045 al., 2015a). More importantly, the amplicons were detectable by an electrochemical biosensor based
1046 on a custom synthesized ferrocence intercalator as a signal transducer. The developed biosensors
1047 allow for the detection of single nucleotide variation and enable the potential of portable sensors for
1048 rapid identification of specific human biomarkers in wastewater.

1049

1050 6. CONCLUSIONS AND FUTURE PERSPECTIVES

1051 WBE is a rapidly developing scientific discipline with a strong transdisciplinary character. It
1052 has shown great progress, and opens up many possibilities for expanding its application to provide
1053 relevant information about lifestyle and public health.

1054 This review has outlined potential wastewater biomarkers of exposure or effect that could be
1055 used for future applications associated with lifestyle and wellbeing studies. However, it has also
1056 discussed limitations and highlighted that more research is needed, for various proposed
1057 biomarkers, before WBE can appropriately be applied. Moreover, several trends, needs and
1058 recommendations are indicated:

- 1059 - Human pharmacokinetic data (metabolism and urinary profile of excretion) are necessary to
1060 ensure that the candidate biomarker is formed in the body in a high proportion and is excreted
1061 mainly via urine. This information is highly relevant not only to back-calculate the
1062 consumption/exposure of a certain substance by a community, but also to distinguish the
1063 amount of a substance coming from human or other sources.
- 1064 - In-sample and in-sewer stability studies are needed for a better application in WBE. Stability
1065 tests are often performed in the laboratory, trying to reproduce the real conditions of
1066 temperature and sewage composition or in-sewer conditions. An alternative would be the use of
1067 *in-silico* tools to predict the stability of a compound in wastewater treatment processes. These
1068 models do not guarantee the formation of a biotransformation product, so it may be used as an
1069 indicator or a guide about the in-sewer stability of a residue and its potential adsorption (Reid
1070 2014). Sorption onto the solid particulate or the conjugation of the biomarkers must also be
1071 taken into account when assessing stability.
- 1072 - Source identification is needed to ensure that discharges from exogenous sources that might
1073 cause overestimation of the real amounts consumed are considered.

- 1074 - Cross validation of data (e.g. concentrations of pharmaceuticals in wastewater with bench-top
1075 sales) is recommended for all applications.
- 1076 - Multiple biomarkers for estimating the population size need to be set to allow for the
1077 normalization of the data. The development of portable biosensors may allow rapid estimation
1078 of the population contributing to the wastewater samples in the near future.
- 1079 - Regular monitoring of sewage for viruses based on similar DNA biosensors may give an early
1080 warning of a possible upcoming outbreak.
- 1081 - Omics approaches also hold promising and important roles in future developments and
1082 applications of endogenous biomarkers analysis in WBE.
- 1083
- 1084

1085 **ACKNOWLEDGEMENTS**

1086 This work was supported by the COST Action ES1307 “SCORE – Sewage biomarker
1087 analysis for community health assessment”. Emma Gracia-Lor is very grateful to Generalitat
1088 Valenciana, Conselleria d’Educació, Investigació, Cultura i Esport (APOSTD/2015, Programa
1089 VALi+d) for her post-doctoral contract. Lubertus Bijlsma acknowledges NPS-Euronet
1090 (HOME/2014/JDRUG/AG/DRUG/7086), co-funded by the European Union, for his post-doctoral
1091 fellowship. Erika Castrignanò, Richard Bade, Juliet Kinyua, Pedram Ramin, Nikolaos I. Rousis,
1092 Yeonsuk Ryu would like to thank the SEWPROF MC ITN project, ‘A new paradigm in drug use
1093 and human health risk assessment: Sewage profiling at the community level’ [grant agreement
1094 317205] supported by the European Union’s Seventh Framework Programme for research,
1095 technological development and demonstration for the financial support. Iria González-Mariño
1096 extends her gratitude to the Galician Council of Culture, Education and Universities for her
1097 postdoctoral contract (Plan Galego de Investigación, Innovación e Crecemento 2011-2015). Foon
1098 Yin Lai acknowledges her postdoctoral fellowship from the University of Antwerp. Luigi Lopardo,
1099 Axel Rydevik and Barbara Kasprzyk-Hordern would like to acknowledge Leverhulme Trust for
1100 funding ‘TOX-EDC, Wastewater profiling for community-wide human exposure assessment from
1101 environmental endocrine disrupting chemicals in personal care and consumer products’ (Project No:
1102 RPG-2013-297). Frederic Been would like to thank the Swiss National Science Foundation (SNF,
1103 P2LAP2_164892) for his post-doctoral grant. This publication reflects the views only of the
1104 authors, and the European Commission cannot be held responsible for any use which may be made
1105 of the information contained therein.

1106

1107

1108 **TABLES**

1109

1110 **Table 1.** Overview of the most relevant biomarkers used so far and potential biomarkers (for more
1111 details, please read the corresponding text and/or supporting information).

1112

Class	Parent compound	Biomarker/potential biomarker	WBE application	Reference
Illicit drugs	Cocaine	Benzoyllecgonine	YES	(Castiglioni and Gracia-Lor, 2015; Gracia-Lor et al., 2016)
	Amphetamine	Amphetamine	YES	(Castiglioni and Gracia-Lor, 2015; Gracia-Lor et al., 2016)
	Methamphetamine	Methamphetamine	YES	(Castiglioni and Gracia-Lor, 2015; Gracia-Lor et al., 2016)
	MDMA	MDMA	YES	(Castiglioni and Gracia-Lor, 2015; Gracia-Lor et al., 2016)
	THC/Cannabis	THC-COOH	YES	(Castiglioni and Gracia-Lor, 2015; Gracia-Lor et al., 2016)
Alcohol	Ethanol	Ethyl sulfate	YES	(Rodríguez-Álvarez et al., 2015)
Tobacco	Nicotine	Cotinine + trans-3'-hydroxycotinine	YES	(Castiglioni et al., 2015b)
Caffeine	Caffeine	See Table S1	NO	
NPS		See Table S1	NO	
Pesticides	20 pyrethroids	3-PBA	YES	(Rousis et al., 2016b)
	Permetrin, cypermetrin, cyflutrin	cis-DCCA	YES	(Rousis et al., 2016b)
	Permetrin, cypermetrin, cyflutrin	trans-DCCA	YES	(Rousis et al., 2016b)
Mycotoxines		See Table S2	NO	
Parabens		See Table S2	NO	
UV-filters		See Table S2	NO	
Plasticizers		See Table S2	NO	
Flame		See Table S2	NO	

retardants				
Pharmaceuticals	Atenolol	Atenolol	YES	(Baz-Lomba et al., 2016; van Nuijs et al., 2015)
	Citalopram	Citalopram	YES	(Baz-Lomba et al., 2016; van Nuijs et al., 2015)
	Carbamazepine	Carbamazepine	YES	(Baz-Lomba et al., 2016; van Nuijs et al., 2015)
	Diclofenac	Diclofenac	YES	(Baz-Lomba et al., 2016)
	Metformin	Metformin	YES	(van Nuijs et al., 2015)
	Valsartan	Valsartan	YES	(van Nuijs et al., 2015)
Benzodiazepines	Oxazepam	Oxazepam	YES	(Baz-Lomba et al., 2016)
Artificial sweeteners	Acesulfame	Acesulfame	YES	(Lai et al., 2015a)
Endogenous Compounds	Serotonin	5-HIAA	YES	(Rico et al., 2016)
	Ammonia	Ammonium	YES	(Been et al., 2014)

1113

1114

1115

1116 **FIGURE CAPTIONS**

1117

1118 **Figure 1.** Main requirements of a biomarker

1119

1120 **REFERENCES**

- 1121 Altintas, Z., Tothill, I., 2013. Biomarkers and biosensors for the early diagnosis of lung cancer.
1122 *Sensors Actuators B Chem.* 188, 988–998. doi:10.1016/j.snb.2013.07.078
- 1123 Andrés-Costa, M.J., Escrivá, Ú., Andreu, V., Picó, Y., 2016. Estimation of alcohol consumption
1124 during “Fallas” festivity in the wastewater of Valencia city (Spain) using ethyl sulfate as a
1125 biomarker. *Sci. Total Environ.* 541, 616–622. doi:10.1016/j.scitotenv.2015.09.126
- 1126 Anfossi, L., Giovannoli, C., Baggiani, C., 2016. Mycotoxin detection. *Curr. Opin. Biotechnol.* 37,
1127 120–126. doi:10.1016/j.copbio.2015.11.005
- 1128 Asimakopoulos, A., Kannan, K., 2016. Neuropsychiatric pharmaceuticals and illicit drugs in
1129 wastewater treatment plants: A review. *Environ. Chem.*
1130 doi:http://dx.doi.org/10.1071/EN15202
- 1131 Bade, R., Bijlsma, L., Sancho, J. V., Baz-Lomba, J.A., Castiglioni, S., Castrignanò, E., Causanilles,
1132 A., Gracia-Lor, E., Kasprzyk-Hordern, B., Kinyua, J., McCall, A.-K., van Nuijs, A.L.N., Ort,
1133 C., Plósz, B.G., Ramin, P., Rousis, N.I., Ryu, Y., Thomas, K. V., Voogt, P. de, Zuccato, E.,
1134 Hernández, F., 2016. Liquid chromatography-tandem mass spectrometry determination of
1135 synthetic cathinones and phenethylamines in influent wastewater of eight European cities.
1136 *Chemosphere.* doi:http://dx.doi.org/10.1016/j.chemosphere.2016.10.107
- 1137 Baker, D.R., Barron, L., Kasprzyk-Hordern, B., 2014. Illicit and pharmaceutical drug consumption
1138 estimated via wastewater analysis. Part A: Chemical analysis and drug use estimates. *Sci. Total*
1139 *Environ.* 487, 629–641. doi:10.1016/j.scitotenv.2013.11.107
- 1140 Baker, D.R., Kasprzyk-Hordern, B., 2011. Critical evaluation of methodology commonly used in
1141 sample collection, storage and preparation for the analysis of pharmaceuticals and illicit drugs
1142 in surface water and wastewater by solid phase extraction and liquid chromatography-mass
1143 spectrometry. *J. Chromatogr. A* 1218, 8036–8059. doi:10.1016/j.chroma.2011.09.012
- 1144 Barnabé, S., Beauchesne, I., Cooper, D.G., Nicell, J.A., 2008. Plasticizers and their degradation
1145 products in the process streams of a large urban physicochemical sewage treatment plant.
1146 *Water Res.* 42, 153–162. doi:10.1016/j.watres.2007.07.043
- 1147 Barr, D.B., 2008. Biomonitoring of exposure to pesticides. *J. Chem. Heal. Saf.* 15, 20–29.
1148 doi:10.1016/j.jchas.2008.07.001
- 1149 Baselt, R.C., 2004. Disposition of toxic drugs and chemicals in man., 7th ed. Biochemical
1150 Publications (California, USA).
- 1151 Baz-Lomba, J.A., Salvatore, S., Gracia-Lor, E., Bade, R., Castiglioni, S., Castrignanò, E.,
1152 Causanilles, A., Hernandez, F., Kasprzyk-Hordern, B., Kinyua, J., McCall, A.-K., van Nuijs,
1153 A., Ort, C., Plósz, B.G., Ramin, P., Reid, M., Rousis, N.I., Ryu, Y., de Voogt, P., Bramness, J.,
1154 Thomas, K., 2016. Comparison of pharmaceutical, illicit drug, alcohol, nicotine and caffeine
1155 levels in wastewater with sale, seizure and consumption data for 8 European cities. *BMC*
1156 *Public Health* 16, 1035. doi:10.1186/s12889-016-3686-5
- 1157 Been, F., Benaglia, L., Lucia, S., Gervasoni, J.P., Esseiva, P., Delémont, O., 2015. Data
1158 triangulation in the context of opioids monitoring via wastewater analyses. *Drug Alcohol*
1159 *Depend.* 151, 203–210. doi:10.1016/j.drugalcdep.2015.03.022
- 1160 Been, F., Rossi, L., Ort, C., Rudaz, S., Delémont, O., Esseiva, P., 2014. Population normalization
1161 with ammonium in wastewater-based epidemiology: Application to illicit drug monitoring.
1162 *Environ. Sci. Technol.* 48, 8162–8169. doi:10.1021/es5008388
- 1163 Bencharit, S., Morton, C.L., Xue, Y., Potter, P.M., Redinbo, M.R., 2003. Structural basis of heroin
1164 and cocaine metabolism by a promiscuous human drug-processing enzyme. *Nat. Struct. Biol.*
1165 10, 349–356. doi:10.1038/nsb919
- 1166 Benowitz, N.L., Jacob, P., 1994. Metabolism of nicotine to cotinine studied by a dual stable isotope
1167 method. *Clin. Pharmacol. Ther.* 56, 483–493. doi:10.1038/clpt.1994.169
- 1168 Bhat, V.S., Durham, J.L., Ball, G.L., English, J.C., 2014. Derivation of an Oral Reference Dose

1169 (RfD) for the Nonphthalate Alternative Plasticizer 1,2-Cyclohexane Dicarboxylic Acid, Di-
 1170 Isononyl Ester (DINCH). *J. Toxicol. Environ. Health. B. Crit. Rev.* 17, 63–94.
 1171 doi:10.1080/10937404.2013.876288

1172 Bijlsma, L., Beltran, E., Boix, C., Sancho, J. V., Hernández, F., 2014. Improvements in analytical
 1173 methodology for the determination of frequently consumed illicit drugs in urban wastewater.
 1174 *Anal. Bioanal. Chem.* 406, 4261–4272. doi:10.1007/s00216-014-7818-4

1175 Bijlsma, L., Emke, E., Hernandez, F., de Voogt, P., 2012. Investigation of drugs of abuse and
 1176 relevant metabolites in Dutch sewage water by liquid chromatography coupled to high
 1177 resolution mass spectrometry. *Chemosphere* 89, 1399–1406.
 1178 doi:10.1016/j.chemosphere.2012.05.110

1179 Bijlsma, L., Miserez, B., Ibáñez, M., Vicent, C., Guillamón, E., Ramsey, J., Hernández, F., 2016.
 1180 Identification and characterization of a novel cathinone derivative 1-(2,3-dihydro-1H-inden-5-
 1181 yl)-2-phenyl-2-(pyrrolidin-1-yl)-ethanone seized by customs in Jersey. *Forensic Toxicol.* 34,
 1182 144–150. doi:10.1007/s11419-015-0299-0

1183 Bisceglia, K.J., Lippa, K.A., 2014. Stability of cocaine and its metabolites in municipal wastewater
 1184 - the case for using metabolite consolidation to monitor cocaine utilization. *Environ. Sci.*
 1185 *Pollut. Res.* 21, 4453–4460. doi:10.1007/s11356-013-2403-5

1186 Blanchard, J., Sawers, S., Jonkman, J., Tang-Liu, D., 1985. Comparison of the urinary metabolite
 1187 profile of caffeine in young and elderly males. *Br. J. Clin. Pharmacol.* 19, 225–232.
 1188 doi:10.1111/j.1365-2125.1985.tb02635.x

1189 Błędzka, D., Gromadzińska, J., Wąsowicz, W., 2014. Parabens. From environmental studies to
 1190 human health. *Environ. Int.* 67, 27–42. doi:10.1016/j.envint.2014.02.007

1191 Boogaerts, T., Covaci, A., Kinyua, J., Neels, H., van Nuijs, A., 2016. Spatial and temporal trends in
 1192 alcohol consumption in Belgian cities: a wastewater-based approach. *Drug Alcohol Depend.*
 1193 160, 170–176. doi:10.1016/j.drugalcdep.2016.01.002

1194 Borova, V.L., Gago-Ferrero, P., Pistos, C., Thomaidis, N.S., 2015. Multi-residue determination of
 1195 10 selected new psychoactive substances in wastewater samples by liquid chromatography–
 1196 tandem mass spectrometry. *Talanta* 144, 592–603. doi:10.1016/j.talanta.2015.06.080

1197 Borova, V.L., Maragou, N.C., Gago-Ferrero, P., Pistos, C., Thomaidis, N.S., 2014. Highly sensitive
 1198 determination of 68 psychoactive pharmaceuticals, illicit drugs, and related human metabolites
 1199 in wastewater by liquid chromatography-tandem mass spectrometry. *Anal. Bioanal. Chem.*
 1200 406, 4273–4285. doi:10.1007/s00216-014-7819-3

1201 Brausch, J.M., Rand, G.M., 2011. A review of personal care products in the aquatic environment:
 1202 Environmental concentrations and toxicity. *Chemosphere* 82, 1518–1532.
 1203 doi:10.1016/j.chemosphere.2010.11.018

1204 Brewer, A.J., Ort, C., Banta-green, C.J., Berset, J., Field, J. a, 2012. Normalized diurnal and
 1205 between-day trends in illicit and legal drug loads that account for changes in the population.
 1206 *Environ. Sci. Technol.* 46, 8305–8314. doi:dx.doi.org/10.1021/es202447r |

1207 Buerge, I.J., Buser, H.-R., Kahle, M., Müller, M.D., Poiger, T., 2009. Ubiquitous occurrence of the
 1208 artificial sweetener acesulfame in the aquatic environment: An ideal chemical marker of
 1209 domestic wastewater in groundwater. *Environ. Sci. Technol.* 43, 4381–4385.

1210 Buerge, I.J., Kahle, M., Buser, H.-R., Müller, M.D., Poiger, T., 2008. Nicotine Derivatives in
 1211 Wastewater and Surface Waters: Application as Chemical Markers for Domestic Wastewater.
 1212 *Environ. Sci. Technol.* 42, 6354–6360. doi:10.1021/es800455q

1213 Buerge, I.J., Poiger, T., Müller, M., Buser, H., 2003. Caffeine, an anthropogenic marker for
 1214 wastewater contamination of surface waters. *Environ. Sci. Technol.* 37, 291–300.

1215 Bull, I.D., Lockheart, M.J., Elhmmali, M.M., Roberts, D.J., Evershed, R.P., 2002. The origin of
 1216 faeces by means of biomarker detection. *Environ. Int.* 27, 647–654. doi:10.1016/S0160-
 1217 4120(01)00124-6

- 1218 Butler, D., Friedler, E., Gatt, K., 1995. Characterising the quantity and quality of domestic
1219 wastewater inflows. *Water Sci. Technol.* 31, 13–24.
- 1220 Button, J., 2015. New Psychoactive Substances: the benzodiazepine boom. *TIAFT Bull.* June 2015.
- 1221 Bye, C., Munro-Faure, A.D., Peck, A.W., Young, P.A., 1973. A comparison of the effects of 1-
1222 benzylpiperazine and dexamphetamine on human performance tests. *Eur. J. Clin. Pharmacol.*
1223 6, 163–169. doi:10.1007/BF00558280
- 1224 Byrd, G.D., Chang, K.M., Greene, J.M., 1992. Evidence for urinary excretion of glucuronide
1225 conjugates of nicotine, cotinine, and trans-3'-hydroxycotinine in smokers. *Drug Metab.*
1226 *Dispos.* 20, 192–197.
- 1227 Callahan, M.M., Robertson, R.S., Branfman, a. R., McComish, M.F., Yesair, D.W., 1983.
1228 Comparison of caffeine metabolism in three nonsmoking populations after oral administration
1229 of radiolabeled caffeine. *Drug Metab. Dispos.* 11, 211–217.
- 1230 Carrillo, J.A., Benitez, J., 1994. Caffeine metabolism in a healthy Spanish population: N-Acetylator
1231 phenotype and oxidation pathways. *Clin. Pharmacol. Ther.* 293–304.
- 1232 Castiglioni, S., Bagnati, R., Fanelli, R., Pomati, F., Calamari, D., Zuccato, E., 2006. Removal of
1233 pharmaceuticals in sewage treatment plants in Italy. *Environ. Sci. Technol.* 40, 357–363.
1234 doi:10.1021/es050991m
- 1235 Castiglioni, S., Bagnati, R., Melis, M., Panawennage, D., Chiarelli, P., Fanelli, R., Zuccato, E.,
1236 2011a. Identification of cocaine and its metabolites in urban wastewater and comparison with
1237 the human excretion profile in urine. *Water Res.* 45, 5141–5150.
1238 doi:10.1016/j.watres.2011.07.017
- 1239 Castiglioni, S., Bijlsma, L., Covaci, A., Emke, E., Hernández, F., Reid, M., Ort, C., Thomas, K. V.,
1240 Van Nuijs, A.L.N., De Voogt, P., Zuccato, E., 2013. Evaluation of uncertainties associated
1241 with the determination of community drug use through the measurement of sewage drug
1242 biomarkers. *Environ. Sci. Technol.* 47, 1452–1460. doi:10.1021/es302722f
- 1243 Castiglioni, S., Borsotti, A., Senta, I., Zuccato, E., 2015a. Wastewater analysis to monitor spatial
1244 and temporal patterns of use of two synthetic recreational drugs, Ketamine and Mephedrone, in
1245 Italy. *Environ. Sci. Technol.* 49, 5563–5570. doi:10.1021/es5060429
- 1246 Castiglioni, S., Gracia-Lor, E., 2015. Chapter 3. Wastewater drug target residues. European
1247 Monitoring Centre for Drugs and Drug Addiction, EMCDDA, in: European Monitoring Centre
1248 for Drugs and Drug Addiction, EMCDDA. Insight 9 Update.
- 1249 Castiglioni, S., Senta, I., Borsotti, A., Davoli, E., Zuccato, E., 2015b. A novel approach for
1250 monitoring tobacco use in local communities by wastewater analysis. *Tob. Control* 24, 38–42.
1251 doi:10.1136/tobaccocontrol-2014-051553
- 1252 Castiglioni, S., Zuccato, E., Chiabrando, C., Fanelli, R., Bagnati, 2008. Mass spectrometric analysis
1253 of illicit drugs in wastewater and surface water. *Mass Spectrom. Rev.* 27, 378–394.
1254 doi:10.1002/mas
- 1255 Castiglioni, S., Zuccato, E., Fanelli, R., 2011b. *Illicit Drugs in the Environment: Occurrence,*
1256 *Analysis, and Fate Using Mass Spectrometry.* Wiley. doi:10.1007/978-1-60761-527-9
- 1257 Castrignanò, E., Lubben, A., Kasprzyk-Hordern, B., 2016. Enantiomeric profiling of chiral drug
1258 biomarkers in wastewater with the usage of chiral liquid chromatography coupled with tandem
1259 mass spectrometry. *J. Chromatogr. A* 1438, 84–99. doi:10.1016/j.chroma.2016.02.015
- 1260 Chen, C., Kostakis, C., Gerber, J.P., Tschärke, B.J., Irvine, R.J., White, J.M., 2014. Towards
1261 finding a population biomarker for wastewater epidemiology studies. *Sci. Total Environ.* 487,
1262 621–628. doi:10.1016/j.scitotenv.2013.11.075
- 1263 Chen, C., Kostakis, C., Irvine, R.J., White, J.M., 2013. Increases in use of novel synthetic stimulant
1264 are not directly linked to decreased use of 3,4-methylenedioxy-N-methylamphetamine
1265 (MDMA). *Forensic Sci. Int.* 231, 278–283. doi:10.1016/j.forsciint.2013.06.007
- 1266 Chiaia, A.C., Banta-green, C., Field, J., 2008. Eliminating solid phase extraction with large-volume

1267 Injection LC/MS/MS: analysis of illicit and legal drugs and human urine indicators in US
1268 wastewaters 42, 8841–8848. doi:10.1021/es802309v

1269 Collins, M., 2011. Some new psychoactive substances: Precursor chemicals and synthesis-driven
1270 end-products. *Drug Test. Anal.* 3, 404–416. doi:10.1002/dta.315

1271 Colomer-Lluch, M., Calero Caceres, W., Jebri, S., Hmaied, F., Muniesa, M., Jofre, J., 2014.
1272 Antibiotic resistance genes in bacterial and bacteriophage fractions of Tunisian and Spanish
1273 wastewaters as markers to compare the antibiotic resistance patterns in each population.
1274 *Environ. Int.* 73, 167–175. doi:10.1016/j.envint.2014.07.003

1275 Commission Regulation 1881/2006, 2006. , Official Journal of the European Union, L 364/5.

1276 Coscollà, C., Colin, P., Yahyaoui, A., Petrique, O., Yusà, V., Mellouki, A., Pastor, A., 2010.
1277 Occurrence of currently used pesticides in ambient air of Centre Region (France). *Atmos.*
1278 *Environ.* 44, 3915–3925. doi:10.1016/j.atmosenv.2010.07.014

1279 Coutu, S., Wyrsh, V., Wynn, H.K., Rossi, L., Barry, D.A., 2013. Temporal dynamics of antibiotics
1280 in wastewater treatment plant influent. *Sci. Total Environ.* 458–460, 20–26.
1281 doi:10.1016/j.scitotenv.2013.04.017

1282 Crews, H., Olivier, L., Wilson, L., 2001. Urinary biomarkers for assessing dietary exposure to
1283 caffeine. *Food Addit Contam.* 18, 1075–1087. doi:10.1080/02652030110056630

1284 Curtin, F., Walker, J., Schulz, P., 1996. Day-to-day intraindividual reliability and interindividual
1285 differences in monoamines excretion. *J. Affect. Disord.* 38, 173–178.

1286 Dahl, H., Hammarberg, A., Franck, J., Helander, A., 2011. Urinary ethyl glucuronide and ethyl
1287 sulfate testing for recent drinking in alcohol-dependent outpatients treated with acamprosate or
1288 placebo. *Alcohol Alcohol.* 46, 553–557. doi:10.1093/alcalc/agr055

1289 Dahl, H., Stephanson, N., Beck, O., Helander, A., 2002. Comparison of urinary excretion
1290 characteristics of ethanol and ethyl glucuronide. *J. Anal. Toxicol.* 26, 201–204.

1291 Daughton, 2012a. Real-time estimation of small-area populations with human biomarkers in
1292 sewage. *Sci. Total Environ.* 414, 6–21. doi:10.1016/j.scitotenv.2011.11.015

1293 Daughton, 2012b. Using biomarkers in sewage to monitor community-wide human health:
1294 Isoprostanes as conceptual prototype. *Sci. Total Environ.* 424, 16–38.
1295 doi:10.1016/j.scitotenv.2012.02.038

1296 Daughton, C.G., 2001. Illicit Drugs in Municipal Sewage, in: *Pharmaceuticals and Care Products in*
1297 *the Environment*. pp. 348–364. doi:10.1021/bk-2001-0791.ch020

1298 Daughton, C.G., Ruhoy, I.S., 2009. Environmental Footprint of Pharmaceuticals: the Significance
1299 of Factors Beyond Direct Excretion To Sewers. *Environ. Toxicol. Chem.* 28, 2495.
1300 doi:10.1897/08-382.1

1301 De Boer, D., Bosman, I.J., Hidvégi, E., Manzoni, C., Benkö, A.A., Dos Reys, L.J.A.L., Maes,
1302 R.A.A., 2001. Piperazine-like compounds: A new group of designer drugs-of-abuse on the
1303 European market. *Forensic Sci. Int.* 121, 47–56. doi:10.1016/S0379-0738(01)00452-2

1304 Dejean, T., Valentini, A., Duparc, A., Pellier-Cuit, S., Pompanon, F., Taberlet, P., Miaud, C., 2011.
1305 Persistence of environmental DNA in freshwater ecosystems. *PLoS One* 6, 8–11.
1306 doi:10.1371/journal.pone.0023398

1307 Dewalque, L., Pirard, C., Charlier, C., 2014. Measurement of urinary biomarkers of parabens,
1308 benzophenone-3, and phthalates in a Belgian population. *Biomed Res. Int.* 2014.
1309 doi:10.1155/2014/649314

1310 Dresen, S., Weinmann, W., Wurst, F.M., 2004. Forensic confirmatory analysis of ethyl sulfate - A
1311 new marker for alcohol consumption - By liquid-chromatography/electrospray
1312 ionization/tandem mass spectrometry. *J. Am. Soc. Mass Spectrom.* 15, 1644–1648.
1313 doi:10.1016/j.jasms.2004.08.004

1314 Ellefsen, K.N., Wohlfarth, A., Swortwood, M.J., Diao, X., Concheiro, M., Huestis, M.A., 2015. 4-
1315 Methoxy- α -PVP: in silico prediction, metabolic stability, and metabolite identification by

1316 human hepatocyte incubation and high-resolution mass spectrometry. *Forensic Toxicol.* 34,
1317 61–75. doi:10.1007/s11419-015-0287-4

1318 EMCDDA, 2016. Assessing illicit drugs in wastewater. *Advances in wastewater-based drug*
1319 *epidemiology, Assessing illicit drugs in wastewater: advances in wastewater-based drug*
1320 *epidemiology, EMCDDA Insights* 22. doi:10.2810/017397

1321 EMCDDA, 2015a. New psychoactive substances in Europe. An update from the EU Early Warning
1322 System (March 2015).

1323 EMCDDA, 2015b. *European Drug Report: Trends and Developments.*

1324 EMCDDA, 2014. Methoxetamine - Report on the risk assessment of 2-(3-methoxyphenyl)-2-
1325 (ethylamino)cyclohexanone (methoxetamine) in the framework of the Council Decision on
1326 new psychoactive substances. *Risk assessments* 26. doi:10.2810/44197

1327 Emke, E., Evans, S., Kasprzyk-Hordern, B., de Voogt, P., 2014. Enantiomer profiling of high loads
1328 of amphetamine and MDMA in communal sewage: a Dutch perspective. *Sci. Total Environ.*
1329 487, 666–72. doi:10.1016/j.scitotenv.2013.11.043

1330 EPCD, 2003. European parliament and council directive (EPCD) 2003/115/EC of 22 December
1331 2003 amending directive 94/35/EC on sweeteners for use in foodstuffs. EPCD.

1332 Euro-CDC, 2012. Summary of the latest data on antibiotic resistance in the European Union. Euro-
1333 Cdc.

1334 European Food Safety Authority, 2013. Annual Report 2013 Annual Report of the European Food
1335 Safety Authority for 2013 1–36.

1336 European Union, 2005. Council Decision 2005/387/JHA on the information exchange, risk
1337 assessment and control of new psychoactive substances.

1338 Ewald, A.H., Ehlers, D., Maurer, H.H., 2008. Metabolism and toxicological detection of the
1339 designer drug 4-chloro-2,5-dimethoxyamphetamine in rat urine using gas chromatography-
1340 mass spectrometry. *Anal. Bioanal. Chem.* 390, 1837–1842. doi:10.1007/s00216-008-1917-z

1341 Ewald, A.H., Fritschi, G., Bork, W.R., Maurer, H.H., 2006. Designer drugs 2,5-dimethoxy-4-
1342 bromo-amphetamine (DOB) and 2,5-dimethoxy-4-bromo-methamphetamine (MDOB): Studies
1343 on their metabolism and toxicological detection in rat urine using gas chromatographic/mass
1344 spectrometric techniques. *J. Mass Spectrom.* 41, 487–498. doi:10.1002/jms.1007

1345 Fantegrossi, W.E., Moran, J.H., Radominska-Pandya, A., Prather, P.L., 2014. Distinct
1346 pharmacology and metabolism of K2 synthetic cannabinoids compared to Δ^9 -THC:
1347 Mechanism underlying greater toxicity? *Life Sci.* 97, 45–54. doi:10.1016/j.lfs.2013.09.017

1348 Feldman, M., Lee, M., 1985. Serotonin content on foods: effect on urinary excretion of 5-
1349 hydroxyindoleacetic acid. *Am. J. Clin. Nutr.* 42, 639–643.

1350 Fermin, L., Vallvey, C., 2004. Intense sweeteners. *Handbook of Food Analysis: Residues and other*
1351 *food component analysis: Volume 2. Chapter 43 Intense Sweeteners.*

1352 Fernández, P., Regenjo, M., Fernández, A.M., Lorenzo, R.A., Carro, A.M., 2014. Optimization of
1353 ultrasound-assisted dispersive liquid-liquid microextraction for ultra performance liquid
1354 chromatography determination of benzodiazepines in urine and hospital wastewater. *Anal.*
1355 *Methods* 6, 8239–8246. doi:10.1039/c4ay01348d

1356 Fitzsimons, M.F., Belt, S.T., 2005. Dynamic behaviour of 1-aminopropan-2-one in sewage: a
1357 preliminary synthetic and spectroscopic study. *Environ. Chem. Lett.* 3, 70–73.
1358 doi:10.1007/s10311-005-0005-2

1359 Fromme, H., Gareis, M., Völkel, W., Gottschalk, C., 2016. Overall internal exposure to mycotoxins
1360 and their occurrence in occupational and residential settings – An overview. *Int. J. Hyg.*
1361 *Environ. Health* 219, 143–165. doi:http://dx.doi.org/10.1016/j.ijheh.2015.11.004

1362 Fromme, H., Schütze, A., Lahrz, T., Kraft, M., Fembacher, L., Siewering, S., Burkardt, R., Dietrich,
1363 S., Koch, H.M., Völkel, W., 2016. Non-phthalate plasticizers in German daycare centers and
1364 human biomonitoring of DINCH metabolites in children attending the centers (LUPE 3). *Int. J.*

1365 Hyg. Environ. Health 219, 33–39. doi:10.1016/j.ijheh.2015.08.002

1366 Gago-Ferrero, P., Díaz-Cruz, M.S., Barceló, D., 2011. Occurrence of multiclass UV filters in
 1367 treated sewage sludge from wastewater treatment plants. *Chemosphere* 84, 1158–1165.
 1368 doi:10.1016/j.chemosphere.2011.04.003

1369 Gao, D.W., Wen, Z.D., 2016. Phthalate esters in the environment: A critical review of their
 1370 occurrence, biodegradation, and removal during wastewater treatment processes. *Sci. Total*
 1371 *Environ.* 541, 986–1001. doi:10.1016/j.scitotenv.2015.09.148

1372 Garattini, S., 1993. *Caffeine, Coffee and Health*. Raven Press, New York.

1373 Gasser, G., Rona, M., Voloshenko, A., Shelkov, R., Tal, N., Pankratov, I., Elhanany, S., Lev, O.,
 1374 2010. Quantitative evaluation of tracers for quantification of wastewater contamination of
 1375 potable water sources. *Environ. Sci. Technol.* 44, 3919–3925. doi:10.1021/es100604c

1376 Gatidou, G., Kinyua, J., van Nuijs, A.L.N., Gracia-Lor, E., Castiglioni, S., Covaci, A., Stasinakis,
 1377 A.S., 2016. Drugs of abuse and alcohol consumption among different groups of population on
 1378 the Greek Island of Lesbos through sewage-based epidemiology. *Sci. Total Environ.* 563–564,
 1379 633–640. doi:10.1016/j.scitotenv.2016.04.130

1380 González-Mariño, I., Gracia-Lor, E., Bagnati, R., Martins, C.P.B., Zuccato, E., Castiglioni, S.,
 1381 2016a. Screening new psychoactive substances in urban wastewater using High Resolution
 1382 Mass Spectrometry. *Anal. Bioanal. Chem.* 408, 4297–4309.

1383 González-Mariño, I., Gracia-Lor, E., Rousis, N.I., Castrignanò, E., Thomas, K. V., Quintana, J.B.,
 1384 Kasprzyk-Hordern, B., Zuccato, E., Castiglioni, S., 2016b. Wastewater-based epidemiology to
 1385 monitor synthetic cathinones use in different European countries. *Environ. Sci. Technol.*
 1386 doi:10.1021/acs.est.6b02644

1387 González-Mariño, I., Quintana, J.B., Rodríguez, I., Cela, R., 2010. Determination of drugs of abuse
 1388 in water by solid-phase extraction, derivatisation and gas chromatography-ion trap-tandem
 1389 mass spectrometry. *J. Chromatogr. A* 1217, 1748–1760. doi:10.1016/j.chroma.2010.01.046

1390 González-Mariño, I., Quintana, J.B., Rodríguez, I., Cela, R., 2009. Simultaneous determination of
 1391 parabens, triclosan and triclocarban in water by liquid chromatography/electrospray ionisation
 1392 tandem mass spectrometry. *Rapid Commun. Mass Spectrom.* 23, 1756–1766. doi:10.1002/rcm

1393 Gorrod, J.W., Wahren, J., 1993. *Nicotine and Related Alkaloids - Absorption, distribution,*
 1394 *metabolism and excretion*. Springer.

1395 Gracia-Lor, E., Martínez, M., Sancho, J. V., Peñuela, G., Hernández, F., 2012a. Multi-class
 1396 determination of personal care products and pharmaceuticals in environmental and wastewater
 1397 samples by ultra-high performance liquid-chromatography-tandem mass spectrometry. *Talanta*
 1398 99, 1011–1023. doi:10.1016/j.talanta.2012.07.091

1399 Gracia-Lor, E., Sancho, J. V., Serrano, R., Hernández, F., 2012b. Occurrence and removal of
 1400 pharmaceuticals in wastewater treatment plants at the Spanish Mediterranean area of Valencia.
 1401 *Chemosphere* 87, 453–462. doi:10.1016/j.chemosphere.2011.12.025

1402 Gracia-Lor, E., Zuccato, E., Castiglioni, S., 2016. Refining correction factors for back-calculation
 1403 of illicit drug use. *Sci. Total Environ.* 573, 1648–1659. doi:10.1016/j.scitotenv.2016.09.179

1404 Grant, D.M., Tang, B.K., Kalow, W., 1983. Variability in caffeine metabolism. *Clin. Pharmacol.*
 1405 *Ther.* 33, 591–602. doi:10.1038/clpt.1983.80

1406 Group, B.D.W., 2001. Biomarkers and surrogate endpoints: Preferred definitions and conceptual
 1407 framework. *Clin. Pharmacol. Ther.* 69, 89–95. doi:10.1067/mcp.2001.113989

1408 Guo, Y., Alomirah, H., Cho, H.-S., Minh, T.B., Mohd, M.A., Nakata, H., Kannan, K., 2011.
 1409 Occurrence of phthalate metabolites in human urine from several Asian countries. *Environ.*
 1410 *Sci. Technol.* 45, 3138–44. doi:10.1021/es103879m

1411 Hagger, J.A., Jones, M.B., Leonard, D.P., Owen, R., Galloway, T.S., 2006. Biomarkers and
 1412 integrated environmental risk assessment: Are there more questions than answers? *Integr.*
 1413 *Environ. Assess. Manag.* 2, 312–329. doi:10.1002/ieam.5630020403

- 1414 Halter, C.C., Dresen, S., Auwaerter, V., Wurst, F.M., Weinmann, W., 2008. Kinetics in serum and
1415 urinary excretion of ethyl sulfate and ethyl glucuronide after medium dose ethanol intake. *Int.*
1416 *J. Legal Med.* 122, 123–128. doi:10.1007/s00414-007-0180-8
- 1417 Helander, A., Beck, O., 2005. Ethyl sulfate: a metabolite of ethanol in humans and a potential
1418 biomarker of acute alcohol intake. *J. Anal. Toxicol.* 29, 270–274.
- 1419 Helfer, A.G., Turcant, A., Boels, D., Ferec, S., Lelievre, B., Welter, J., Meyer, M.R., Maurer, H.H.,
1420 2007. Elucidation of the metabolites of the novel psychoactive substance 4-methyl-N-ethyl-
1421 cathinone (4-MEC) in human urine and pooled liver microsomes by GC-MS and LC-HR-
1422 MS/MS techniques and of its detectability by GC-MS or LC-MS(n) standard screening
1423 approach. *Drug Test. Anal.* 7, 368–375.
- 1424 Hellmér, M., Paxéus, N., Magnius, L., Enache, L., Arnholm, B., Johansson, A., Bergström, T.,
1425 Norder, H., 2014. Detection of pathogenic viruses in sewage provided early warnings of
1426 hepatitis A virus and norovirus outbreaks. *Appl. Environ. Microbiol.* 80, 6771–6781.
1427 doi:10.1128/AEM.01981-14
- 1428 Hernández, A.F., Parrón, T., Tsatsakis, A.M., Requena, M., Alarcón, R., López-Guarnido, O., 2013.
1429 Toxic effects of pesticide mixtures at a molecular level: Their relevance to human health.
1430 *Toxicology* 307, 136–145. doi:10.1016/j.tox.2012.06.009
- 1431 Hernández, F., Castiglioni, S., Covaci, A., Voogt, P. de, Emke, E., Kasprzyk-Hordern, B., Ort, C.,
1432 Reid, M., Sancho, J. V., van Nuijs, Alexander L.N Zuccato, E., Bijlsma, L., 2016. Mass
1433 spectrometric strategies for the investigation of biomarkers of illicit drug use in wastewater.
1434 *Mass Spectrom. Rev.* doi:DOI 10.1002/mas.21525
- 1435 Herrero, L., Calvarro, S., Fernández, M.A., Quintanilla-López, J.E., González, M.J., Gómara, B.,
1436 2015. Feasibility of ultra-high performance liquid and gas chromatography coupled to mass
1437 spectrometry for accurate determination of primary and secondary phthalate metabolites in
1438 urine samples. *Anal. Chim. Acta* 853, 625–636. doi:10.1016/j.aca.2014.09.043
- 1439 Heyndrickx, E., Sioen, I., Huybrechts, B., Callebaut, A., De Henauw, S., De Saeger, S., 2015.
1440 Human biomonitoring of multiple mycotoxins in the Belgian population: Results of the
1441 BIOMYCO study. *Environ. Int.* 84, 82–89. doi:10.1016/j.envint.2015.06.011
- 1442 Høiseth, G., Bernard, J.P., Stephanson, N., Normann, P.T., Christophersen, A.S., Mørland, J.,
1443 Helander, A., 2008. Comparison between the urinary alcohol markers EtG, EtS, and GTOL/5-
1444 HIAA in a controlled drinking experiment. *Alcohol Alcohol.* 43, 187–191.
1445 doi:10.1093/alcalc/agg175
- 1446 Hu, P., Chen, X., Whitener, R.J., Boder, E.T., Jones, J.O., Porollo, A., Chen, J., Zhao, L., 2013.
1447 Effects of parabens on adipocyte differentiation. *Toxicol. Sci.* 131, 56–70.
1448 doi:10.1093/toxsci/kfs262
- 1449 Huang, C., Renew, J.E., Smeby, K.L., Pinkston, K., Sedlak, D.L., 2011. Assessment of potential
1450 antibiotic contaminants in water and preliminary occurrence analysis. *J. Contemp. Water Res.*
1451 *Educ.* 120, 4.
- 1452 Hukkanen, J., 2005. Metabolism and Disposition Kinetics of Nicotine. *Pharmacol. Rev.* 57, 79–115.
1453 doi:10.1124/pr.57.1.3
- 1454 Hummel, D., Löffler, D., Fink, G., Ternes, T. a, 2006. Simultaneous determination of psychoactive
1455 drugs and their metabolites in aqueous matrices by liquid chromatography mass Spectrometry.
1456 *Environ. Sci. Technol.* 40, 7321–8.
- 1457 Huppertz, L.M., Bisel, P., Westphal, F., Franz, F., Auwarter, V., Moosmann, B., 2015.
1458 Characterization of the four designer benzodiazepines clonazolam, deschloroetizolam,
1459 flubromazolam, and meclonazepam, and identification of their in vitro metabolites. *Forensic*
1460 *Toxicol.* 33, 388–395. doi:10.1007/s11419-015-0277-6
- 1461 Hutter, M., Broecker, S., Kneisel, S., Auwärter, V., 2012. Identification of the major urinary
1462 metabolites in man of seven synthetic cannabinoids of the aminoalkylindole type present as

1463 adulterants in “herbal mixtures” using LC-MS/MS techniques. *J. Mass Spectrom.* 47, 54–65.
1464 doi:10.1002/jms.2026

1465 Hyde, J.F., Adams, R., 1928. Synthetic homologs of d,l-ephedrine. *J. Am. Chem. Soc.* 50, 2287–
1466 2292.

1467 Jelic, A., Gros, M., Petrovic, M., Ginebreda, A., Barceló, D., 2012. Occurrence and elimination of
1468 pharmaceuticals during conventional wastewater treatment. *Emerg. Prior. Pollut. Rivers* 1–23.
1469 doi:10.1007/978-3-642-25722-3

1470 Johnson, R.D., Botch-Jones, S.R., 2013. The Stability of Four Designer Drugs: MDPV,
1471 Mephedrone, BZP and TFMPP in Three Biological Matrices under Various Storage
1472 Conditions. *J. Anal. Toxicol.* 37, 51–55. doi:10.1093/jat/bks138

1473 Joint Fao Oms Expert Committee On Food Additives, 2010. Toxicological and Health Aspects of
1474 Bisphenol A Joint FAO / WHO Expert Meeting. *Fao Who* 60.

1475 Jones, A.W., 1990. Excretion of alcohol in urine and diuresis in healthy men in relation to their age,
1476 the dose administered and the time after drinking. *Forensic Sci. Int.* 45, 217–224.

1477 Kamata, T., Katagi, M., Kamata, H.T., Miki, A., Shima, N., Zaitso, K., Nishikawa, M., Tanaka, H.,
1478 Honda, K., Tsuchihashi, H., 2006. Metabolism of the psychotomimetic tryptamine derivative
1479 5-methoxy-N,N-diisopropyltryptamine in humans: identification and quantification of its
1480 urinary metabolites. *Drug Metab. Dispos.* 34, 281–288. doi:10.1124/dmd.105.005835.

1481 Kasprzyk-Hordern, B., Baker, D.R., 2012. Enantiomeric profiling of chiral drugs in wastewater and
1482 receiving waters. *Environ. Sci. Technol.* 46, 1681–1691. doi:10.1021/es203113y

1483 Kasprzyk-Hordern, B., Dinsdale, R.M., Guwy, A.J., 2008. Multiresidue methods for the analysis of
1484 pharmaceuticals, personal care products and illicit drugs in surface water and wastewater by
1485 solid-phase extraction and ultra performance liquid chromatography-electrospray tandem mass
1486 spectrometry. *Anal. Bioanal. Chem.* 391, 1293–1308. doi:10.1007/s00216-008-1854-x

1487 Khan, U., Nicell, J. a., 2011. Refined sewer epidemiology mass balances and their application to
1488 heroin, cocaine and ecstasy. *Environ. Int.* 37, 1236–1252. doi:10.1016/j.envint.2011.05.009

1489 Kim, M.J., Kwack, S.J., Lim, S.K., Kim, Y.J., Roh, T.H., Choi, S.M., Kim, H.S., Lee, B.M., 2015.
1490 Toxicological evaluation of isopropylparaben and isobutylparaben mixture in Sprague-Dawley
1491 rats following 28 days of dermal exposure. *Regul. Toxicol. Pharmacol.* 73, 544–51.
1492 doi:10.1016/j.yrtph.2015.08.005

1493 Kinyua, J., Covaci, A., Maho, W., McCall, A.-K., Neels, H., van Nuijs, A.L.N., 2015. Sewage-
1494 based epidemiology in monitoring the use of new psychoactive substances: Validation and
1495 application of an analytical method using LC-MS/MS. *Drug Test. Anal.* 7, 812–818.
1496 doi:10.1002/dta.1777

1497 Kokkinos, P., Ziros, P., Meri, D., Filippidou, S., Kolla, S., Galanis, A., Vantarakis, A., 2011.
1498 Environmental surveillance. An additional/alternative approach for virological surveillance in
1499 Greece? *Int. J. Environ. Res. Public Health* 8, 1914–1922. doi:10.3390/ijerph8061914

1500 Kokotou, M.G., Asimakopoulos, A.G., Thomaidis, N.S., 2012. Artificial sweeteners as emerging
1501 pollutants in the environment: Analytical methodologies and environmental impact. *Anal.*
1502 *Methods* 4, 3057–3070.

1503 Kokotou, M.G., Asimakopoulou, A., Thomaidis, N.S., 2013. Determination of eight artificial
1504 sweeteners in wastewater by hydrophilic interaction liquid chromatography-tandem mass
1505 spectrometry. *Anal. Methods* 5, 3825. doi:10.1039/c3ay40599k

1506 Kolpin, D.W., Schenzel, J., Meyer, M.T., Phillips, P.J., Hubbard, L.E., Scott, T.M., Bucheli, T.D.,
1507 2014. Mycotoxins: Diffuse and point source contributions of natural contaminants of emerging
1508 concern to streams. *Sci. Total Environ.* 470–471, 669–676.
1509 doi:10.1016/j.scitotenv.2013.09.062

1510 Kosjek, T., Perko, S., Zupanc, M., Zanoški Hren, M., Landeka Dragičević, T., Žigon, D., Kompare,
1511 B., Heath, E., 2012. Environmental occurrence, fate and transformation of benzodiazepines in

- 1512 water treatment. *Water Res.* 46, 355–368. doi:10.1016/j.watres.2011.10.056
- 1513 Kumaraswamy, R., Amha, Y.M., Anwar, M.Z., Henschel, A., Rodríguez, J., Ahmad, F., 2014.
- 1514 Molecular analysis for screening human bacterial pathogens in municipal wastewater treatment
- 1515 and reuse. *Environ. Sci. Technol.* 48, 11610–9. doi:10.1021/es502546t
- 1516 Kümmerer, K., 2009. Antibiotics in the aquatic environment - A review - Part I. *Chemosphere* 75,
- 1517 417–434. doi:10.1016/j.chemosphere.2008.11.086
- 1518 Laganà, A., Bacaloni, A., De Leva, I., Faberi, A., Fago, G., Marino, A., 2004. Analytical
- 1519 methodologies for determining the occurrence of endocrine disrupting chemicals in sewage
- 1520 treatment plants and natural waters. *Anal. Chim. Acta* 501, 79–88.
- 1521 doi:10.1016/j.aca.2003.09.020
- 1522 Lai, F.Y., Anuj, S., Bruno, R., Carter, S., Gartner, C., Hall, W., Kirkbride, K.P., Mueller, J.F.,
- 1523 O'Brien, J.W., Prichard, J., Thai, P.K., Ort, C., 2015a. Systematic and day-to-day effects of
- 1524 chemical-derived population estimates on wastewater-based drug epidemiology. *Environ. Sci.*
- 1525 *Technol.* 49, 999–1008. doi:10.1021/es503474d
- 1526 Lai, F.Y., Erratico, C., Kinyua, J., Mueller, J.F., Covaci, A., van Nuijs, A.L.N., 2015b. Liquid
- 1527 chromatography-quadrupole time-of-flight mass spectrometry for screening in vitro drug
- 1528 metabolites in humans: Investigation on seven phenethylamine-based designer drugs. *J. Pharm.*
- 1529 *Biomed. Anal.* 114, 355–375. doi:10.1016/j.jpba.2015.06.016
- 1530 Lai, F.Y., Ort, C., Gartner, C., Carter, S., Prichard, J., Kirkbride, P., Bruno, R., Hall, W.,
- 1531 Eaglesham, G., Mueller, J.F., 2011. Refining the estimation of illicit drug consumptions from
- 1532 wastewater analysis: Co-analysis of prescription pharmaceuticals and uncertainty assessment.
- 1533 *Water Res.* 45, 4437–4448. doi:10.1016/j.watres.2011.05.042
- 1534 Lange, F.T., Scheurer, M., Brauch, H.J., 2012. Artificial sweeteners-A recently recognized class of
- 1535 emerging environmental contaminants: A review. *Anal. Bioanal. Chem.* 403, 2503–2518.
- 1536 doi:10.1007/s00216-012-5892-z
- 1537 Le Chatelier, E., Nielsen, T., Qin, J., Prifti, E., Hildebrand, F., Falony, G., Almeida, M.,
- 1538 Arumugam, M., Batto, J.-M., Kennedy, S., Leonard, P., Li, J., Burgdorf, K., Grarup, N.,
- 1539 Jørgensen, T., Brandslund, I., Nielsen, H.B., Juncker, A.S., Bertalan, M., Levenez, F., Pons,
- 1540 N., Rasmussen, S., Sunagawa, S., Tap, J., Tims, S., Zoetendal, E.G., Brunak, S., Clément, K.,
- 1541 Doré, J., Kleerebezem, M., Kristiansen, K., Renault, P., Sicheritz-Ponten, T., de Vos, W.M.,
- 1542 Zucker, J.-D., Raes, J., Hansen, T., Bork, P., Wang, J., Ehrlich, S.D., Pedersen, O., Guedon, E.,
- 1543 Delorme, C., Layec, S., Khaci, G., van de Guchte, M., Vandemeulebrouck, G., Jamet, A.,
- 1544 Dervyn, R., Sanchez, N., Maguin, E., Haimet, F., Winogradski, Y., Cultrone, A., Leclerc, M.,
- 1545 Juste, C., Blottière, H., Pelletier, E., LePaslier, D., Artiguenave, F., Bruls, T., Weissenbach, J.,
- 1546 Turner, K., Parkhill, J., Antolin, M., Manichanh, C., Casellas, F., Boruel, N., Varela, E.,
- 1547 Torrejon, A., Guarner, F., Denariáz, G., Derrien, M., van Hylckama Vlieg, J.E.T., Veiga, P.,
- 1548 Oozeer, R., Knol, J., Rescigno, M., Brechot, C., M'Rini, C., Mérieux, A., Yamada, T., 2013.
- 1549 Richness of human gut microbiome correlates with metabolic markers. *Nature* 500, 541–6.
- 1550 doi:10.1038/nature12506
- 1551 Le Fol, V., Aït-Aïssa, S., Cabaton, N., Dolo, L., Grimaldi, M., Balaguer, P., Perdu, E., Debrauwer,
- 1552 L., Brion, F., Zalko, D., 2015. Cell-specific biotransformation of benzophenone-2 and
- 1553 bisphenol-s in zebrafish and human in vitro models used for toxicity and estrogenicity
- 1554 screening. *Environ. Sci. Technol.* 49, 3860–3868. doi:10.1021/es505302c
- 1555 Liu, S., Wu, P., Li, W., Zhang, H., Cai, C., 2011. An electrochemical approach for detection of
- 1556 DNA methylation and assay of the methyltransferase activity. *Chem. Commun. (Camb)*. 47,
- 1557 2844–6. doi:10.1039/c0cc05153e
- 1558 Löffler, D., Römbke, J., M, M., Ternes TA., 2005. Environmental fate of pharmaceuticals in
- 1559 water/sediment systems. *Environ. Sci. Technol.* 39, 5209–5218.
- 1560 Loftus, N.J., Laird, W.J.D., Steel, G.T., Wilks, M.F., Woollen, B.H., 1993. Metabolism and

- 1561 pharmacokinetics of deuterium-labelled di-2-(ethylhexyl) adipate (DEHA) in humans. *Food*
1562 *Chem. Toxicol.* 31, 609–614. doi:10.1016/0278-6915(93)90042-W
- 1563 Loos, R., Carvalho, R., Comero, S., António, D., Ghiani, M., Lettieri, T., Locoro, G., Paracchini,
1564 B., Tavazzi, S., Gawlik, B., Blaha, L., Jarosova, B., Voorspoels, S., Schwesig, D., Haglund, P.,
1565 Fick, J., Gans, O., 2012. EU Wide Monitoring Survey on Waste Water Treatment Plant
1566 Effluents, JRC scientific and policy report. doi:10.2788/60663
- 1567 Lopes, A., Silva, N., Bronze, M.R., Ferreira, J., Morais, J., 2014. Analysis of cocaine and nicotine
1568 metabolites in wastewater by liquid chromatography-tandem mass spectrometry. Cross abuse
1569 index patterns on a major community. *Sci. Total Environ.* 487, 673–680.
1570 doi:10.1016/j.scitotenv.2013.10.042
- 1571 Lostia, A.M., Vicente, J.L., Cowan, D.A., 2013. Measurement of ethyl glucuronide, ethyl sulphate
1572 and their ratio in the urine and serum of healthy volunteers after two doses of alcohol. *Alcohol*
1573 *Alcohol.* 48, 74–82. doi:10.1093/alcal/ags108
- 1574 Louis, G.M.B., Chen, Z., Kim, S., Sapra, K., Phil, M., Bae, J., Kannan, K., 2015. Urinary
1575 concentrations of benzophenone-type ultraviolet light filters and semen quality. *Fertil. Steril.*
1576 104, 989–996.
- 1577 Louis, G.M.B., Kannan, K., Sapra, K.J., Maisog, J., Sundaram, R., 2014. Urinary concentrations of
1578 benzophenone-type ultraviolet radiation filters and couples' fecundity. *Am. J. Epidemiol.* 180,
1579 1168–1175. doi:10.1093/aje/kwu285
- 1580 Lyssimachou, A., Santos, J.G., André, A., Soares, J., Lima, D., Guimarães, L., Almeida, C.M.R.,
1581 Teixeira, C., Castro, L.F.C., Santos, M.M., 2015. The Mammalian “Obesogen” Tributyltin
1582 Targets Hepatic Triglyceride Accumulation and the Transcriptional Regulation of Lipid
1583 Metabolism in the Liver and Brain of Zebrafish. *PLoS One* 10, e0143911.
1584 doi:10.1371/journal.pone.0143911
- 1585 Mackulák, T., Birošová, L., Grabic, R., Škubák, J., Bodík, I., 2015. National monitoring of nicotine
1586 use in Czech and Slovak Republic based on wastewater analysis. *Environ. Sci. Pollut. Res.*
1587 doi:10.1007/s11356-015-4648-7
- 1588 Managaki, S., Takada, H., Kim, D.M., Horiguchi, T., Shiraishi, H., 2006. Three-dimensional
1589 distributions of sewage markers in Tokyo Bay water - Fluorescent whitening agents (FWAs).
1590 *Mar. Pollut. Bull.* 52, 281–292. doi:10.1016/j.marpolbul.2005.08.025
- 1591 Mardal, M., Meyer, M.R., 2014. Studies on the microbial biotransformation of the novel
1592 psychoactive substance methylenedioxypyrovalerone (MDPV) in wastewater by means of
1593 liquid chromatography-high resolution mass spectrometry/mass spectrometry. *Sci. Total*
1594 *Environ.* 493, 588–595. doi:10.1016/j.scitotenv.2014.06.016
- 1595 Mardal, M., Miserez, B., Bade, R., Portolés, T., Bischoff, M., Hernández, F., Meyer, M.R., 2016. 3-
1596 Fluorophenmetrazine, a fluorinated analogue of phenmetrazine: Studies on in vivo metabolism
1597 in rat and human, in vitro metabolism in human CYP isoenzymes and microbial
1598 biotransformation in *Pseudomonas Putida* and wastewater using GC and LC coupled to (HR). *J.*
1599 *Pharm. Biomed. Anal.* 128, 485–495. doi:10.1016/j.jpba.2016.06.011
- 1600 Marklund, A., Andersson, B., Haglund, P., 2005. Organophosphorus flame retardants and
1601 plasticizers in Swedish sewage treatment plants. *Environ. Sci. Technol.* 39, 7423–7429.
1602 doi:10.1021/es051013l
- 1603 Marroquín-Cardona, A.G., Johnson, N.M., Phillips, T.D., Hayes, A.W., 2014. Mycotoxins in a
1604 changing global environment - A review. *Food Chem. Toxicol.* 69, 220–230.
1605 doi:10.1016/j.fct.2014.04.025
- 1606 Martínez-Bueno, M.J., Uclés, S., Hernando, M.D., Davoli, E., Fernández-Alba, A.R., 2011.
1607 Evaluation of selected ubiquitous contaminants in the aquatic environment and their
1608 transformation products. A pilot study of their removal from a sewage treatment plant. *Water*
1609 *Res.* 45, 2331–2341. doi:10.1016/j.watres.2011.01.011

- 1610 Mastroianni, N., Lopez de Alda, M., Barcelo, D., 2014. Analysis of ethyl sulfate in raw wastewater
1611 for estimation of alcohol consumption and its correlation with drugs of abuse in the city of
1612 Barcelona. *J. Chromatogr. A* 1360, 93–99. doi:10.1016/j.chroma.2014.07.051
- 1613 Maurer, H.H., Kraemer, T., Springer, D., Staack, R.F., 2004. Chemistry, pharmacology, toxicology,
1614 and hepatic metabolism of designer drugs of the amphetamine (ecstasy), piperazine, and
1615 pyrrolidinophenone types: a synopsis. *Ther. Drug Monit.* 26, 127–131.
- 1616 McCall, A.-K., Bade, R., Kinyua, J., Lai, F.Y., Thai, P.K., Covaci, A., Bijlsma, L., van Nuijs,
1617 A.L.N., Ort, C., 2016a. Critical review on the stability of illicit drugs in sewers and wastewater
1618 samples. *Water Res.* 88, 933–947. doi:10.1016/j.watres.2015.10.040
- 1619 McCall, A.-K., Scheidegger, A., Madry, M.M., Steuer, A.E., Weissbrodt, D.G., Vanrolleghem,
1620 P.A., Kraemer, T., Morgenroth, E., Ort, C., 2016b. Influence of Different Sewer Biofilms on
1621 Transformation Rates of Drugs. Submitted.
- 1622 McLellan, S.L., Eren, A.M., 2014. Discovering new indicators of fecal pollution. *Trends Microbiol.*
1623 22, 697–706. doi:10.1016/j.tim.2014.08.002
- 1624 McLellan, S.L., Newton, R.J., Vandewalle, J.L., Shanks, O.C., Huse, S.M., Eren, A.M., Sogin, M.L.,
1625 2013. Sewage reflects the distribution of human faecal lachnospiraceae. *Environ. Microbiol.*
1626 15, 2213–2227. doi:10.1111/1462-2920.12092
- 1627 Meyer, M.R., Bach, M., Welter, J., Bovens, M., Turcant, A., Maurer, H.H., 2013. Ketamine-derived
1628 designer drug methoxetamine: Metabolism including isoenzyme kinetics and toxicological
1629 detectability using GC-MS and LC-(HR-)MSn. *Anal. Bioanal. Chem.* 405, 6307–6321.
1630 doi:10.1007/s00216-013-7051-6
- 1631 Meyer, M.R., Du, P., Schuster, F., Maurer, H.H., 2010a. Studies on the metabolism of the α -
1632 pyrrolidinophenone designer drug methylenedioxy-pyrovalerone (MDPV) in rat and human
1633 urine and human liver microsomes using GC-MS and LC-high-resolution MS and its
1634 detectability in urine by GC-MS. *J. Mass Spectrom.* 45, 1426–1442. doi:10.1002/jms.1859
- 1635 Meyer, M.R., Mauer, S., Meyer, G.M., Dinger, J., Klein, B., Westphal, F., Maurer, H.H., 2014. The
1636 in vivo and in vitro metabolism and the detectability in urine of 3',4'-methylenedioxy-alpha-
1637 pyrrolidinobutyrophenone (MDPBP), a new pyrrolidinophenone-type designer drug, studied
1638 by GC-MS and LC-MS(n). *Drug Test. Anal.* 6, 746–756.
- 1639 Meyer, M.R., Maurer, H.H., 2010. Metabolism of Designer Drugs of Abuse: An Updated Review.
1640 *Curr. Drug Metab.* 11, 468–482. doi:10.2174/138920010791526042
- 1641 Meyer, M.R., Vollmar, C., Schwaninger, A.E., Wolf, E.U., Maurer, H.H., 2012. New cathinone-
1642 derived designer drugs 3-bromomethcathinone and 3-fluoromethcathinone: Studies on their
1643 metabolism in rat urine and human liver microsomes using GC-MS and LC-high-resolution
1644 MS and their detectability in urine. *J. Mass Spectrom.* 47, 253–262. doi:10.1002/jms.2960
- 1645 Meyer, M.R., Wilhelm, J., Peters, F.T., Maurer, H.H., 2010b. Beta-keto amphetamines: Studies on
1646 the metabolism of the designer drug mephedrone and toxicological detection of mephedrone,
1647 butylone, and methylone in urine using gas chromatography - Mass spectrometry. *Anal.*
1648 *Bioanal. Chem.* 397, 1225–1233. doi:10.1007/s00216-010-3636-5
- 1649 Michely, J.A., Helfer, A.G., Brandt, S.D., Meyer, M.R., Maurer, H.H., 2015. Metabolism of the
1650 new psychoactive substances N,N-diallyltryptamine (DALT) and 5-methoxy-DALT and their
1651 detectability in urine by GC-MS, LC-MS n , and LC-HR-MS-MS. *Anal. Bioanal. Chem.*
1652 407, 7831–7842. doi:10.1007/s00216-015-8955-0
- 1653 Moosmann, B., Huppertz, L.M., Hutter, M., Buchwald, A., Ferlino, S., Auwärter, V., 2013.
1654 Detection and identification of the designer benzodiazepine flubromazepam and preliminary
1655 data on its metabolism and pharmacokinetics. *J. Mass Spectrom.* 48, 1150–1159.
1656 doi:10.1002/jms.3279
- 1657 Mwenesongole, E.M., Lata Gautam, S.W.H., Waterhouse, John W. Colea, M.D., 2013.
1658 Simultaneous detection of controlled substances in waste water. *Anal. Methods* 5, 3248–3254.

- 1659 Narimatsu, S., Yonemoto, R., Masuda, K., Katsu, T., Asanuma, M., Kamata, T., Katagi, M.,
1660 Tsuchihashi, H., Kumamoto, T., Ishikawa, T., Naito, S., Yamano, S., Hanioka, N., 2008.
1661 Oxidation of 5-methoxy-N,N-diisopropyltryptamine in rat liver microsomes and recombinant
1662 cytochrome P450 enzymes. *Biochem. Pharmacol.* 75, 752–760. doi:10.1016/j.bcp.2007.09.019
1663 Newton, R.J., McLellan, S.L., Dila, D.K., Vineis, J.H., Morrison, H.G., Murat Eren, A., Sogin,
1664 M.L., 2015. Sewage reflects the microbiomes of human populations. *MBio* 6, 1–9.
1665 doi:10.1128/mBio.02574-14
1666 Ntzani, E.E., Chondrogiorgi, M., Ntritsos, G., Evangelou, E., Tzoulaki, I., 2013. Literature review
1667 on epidemiological studies linking exposure to pesticides. EFSA supporting publication
1668 2013:EN-497.
1669 O'Brien, J.W., Thai, P.K., Brandsma, S.H., Leonards, P.E.G., Ort, C., Mueller, J.F., 2015.
1670 Wastewater analysis of Census day samples to investigate per capita input of
1671 organophosphorus flame retardants and plasticizers into wastewater. *Chemosphere* 138, 328–
1672 334. doi:10.1016/j.chemosphere.2015.06.014
1673 O'Brien, J.W., Thai, P.K., Eaglesham, G., Ort, C., Scheidegger, A., Carter, S., Lai, F.Y., Mueller,
1674 J.F., 2014. A Model to Estimate the Population Contributing to the Wastewater Using Samples
1675 Collected on Census Day. *Environ. Sci. Technol.* 48, 517–525. doi:10.1021/es403251g
1676 Ocaña-González, J.A., Villar-Navarro, M., Ramos-Payán, M., Fernández-Torres, R., Bello-López,
1677 M.A., 2015. New developments in the extraction and determination of parabens in cosmetics
1678 and environmental samples. A review. *Anal. Chim. Acta* 858, 1–15.
1679 doi:10.1016/j.aca.2014.07.002
1680 Olfson, M., King, M., Schoenbaum, M., 2015. Benzodiazepine use in the United States. *JAMA*
1681 *psychiatry* 72, 136–42. doi:10.1001/jamapsychiatry.2014.1763
1682 Olofsson, U., Brorström-Lundén, E., Kylin, H., Haglund, P., 2013. Comprehensive mass flow
1683 analysis of Swedish sludge contaminants. *Chemosphere* 90, 28–35.
1684 doi:10.1016/j.chemosphere.2012.07.002
1685 Oppenheimer, J., Eaton, A., Badruzzaman, M., Haghani, A.W., Jacangelo, J.G., 2011. Occurrence
1686 and suitability of sucralose as an indicator compound of wastewater loading to surface waters
1687 in urbanized regions. *Water Res.* 45, 4019–4027. doi:10.1016/j.watres.2011.05.014
1688 Ordóñez, E.Y., Quintana, J.B., Rodil, R., Cela, R., 2012. Determination of artificial sweeteners in
1689 water samples by solid-phase extraction and liquid chromatography-tandem mass
1690 spectrometry. *J. Chromatogr. A* 1256, 197–205. doi:10.1016/j.chroma.2012.07.073
1691 Ort, C., van Nuijs, A.L.N., Berset, J.D., Bijlsma, L., Castiglioni, S., Covaci, A., de Voogt, P., Emke,
1692 E., Fatta-Kassinos, D., Griffiths, P., Hernández, F., González-Mariño, I., Grabic, R., Kasprzyk-
1693 Hordern, B., Mastroianni, N., Meierjohann, A., Nefau, T., Östman, M., Pico, Y., Racamonde,
1694 I., Reid, M., Slobodnik, J., Terzic, S., Thomaidis, N., Thomas, K. V., 2014. Spatial differences
1695 and temporal changes in illicit drug use in Europe quantified by wastewater analysis.
1696 *Addiction* 109, 1338–1352. doi:10.1111/add.12570
1697 Ozturk, S., Ozturk, Y.E., Yeter, O., Alpertunga, B., 2015. Application of a validated LC-MS/MS
1698 method for JWH-073 and its metabolites in blood and urine in real forensic cases. *Forensic*
1699 *Sci. Int.* 257, 165–171. doi:10.1016/j.forsciint.2015.08.013
1700 Papaseit, E., Farré, M., Schifano, F., Torrens, M., 2014. Emerging drugs in Europe. *Curr. Opin.*
1701 *Psychiatry* 27, 243–50. doi:10.1097/YCO.0000000000000071
1702 Pawlik, E., Plässer, G., Mahler, H., Daldrup, T., 2012. Studies on the phase I metabolism of the new
1703 designer drug 3-fluoromethcathinone using rabbit liver slices. *Int. J. Legal Med.* 126, 231–240.
1704 doi:10.1007/s00414-011-0601-6
1705 Pedrouzo, M., Borrull, F., Pocurull, E., Marcé, R.M., 2011. Drugs of abuse and their metabolites in
1706 waste and surface waters by liquid chromatography-tandem mass spectrometry. *J. Sep. Sci.* 34,
1707 1091–1101. doi:10.1002/jssc.201100043

- 1708 Pertwee, R.G., 2008. Ligands that target cannabinoid receptors in the brain: From THC to
1709 anandamide and beyond. *Addict. Biol.* 13, 147–159. doi:10.1111/j.1369-1600.2008.00108.x
- 1710 Petrie, B., Barden, R., Kasprzyk-Hordern, B., 2015. A review on emerging contaminants in
1711 wastewaters and the environment: Current knowledge, understudied areas and
1712 recommendations for future monitoring. *Water Res.* 72, 3–27.
1713 doi:10.1016/j.watres.2014.08.053
- 1714 Petrie, B., Youdan, J., Barden, R., Kasprzyk-Hordern, B., 2016. New Framework To Diagnose the
1715 Direct Disposal of Prescribed Drugs in Wastewater - A Case Study of the Antidepressant
1716 Fluoxetine. *Environ. Sci. Technol.* 50, 3781–3789. doi:10.1021/acs.est.6b00291
- 1717 Pischon, T., 2009. Use of obesity biomarkers in cardiovascular epidemiology. *Dis. Markers* 26,
1718 247–263. doi:10.3233/DMA-2009-0634
- 1719 Plósz, B.G., Reid, M.J., Borup, M., Langford, K.H., Thomas, K. V., 2013. Biotransformation
1720 kinetics and sorption of cocaine and its metabolites and the factors influencing their estimation
1721 in wastewater. *Water Res.* 47, 2129–2140. doi:10.1016/j.watres.2012.12.034
- 1722 Polesel, F., Andersen, H.R., Trapp, S., Plósz, B.G., 2016. Removal of antibiotics in biological
1723 wastewater treatment systems – A critical assessment using the Activated Sludge Modelling
1724 framework for Xenobiotics (ASM-X). *Submitt. to Environ. Sci. Technol.* acs.est.6b01899.
1725 doi:10.1021/acs.est.6b01899
- 1726 Poste, G., 2011. Bring on the biomarkers. *Nature* 469, 156–157. doi:10.1038/469156a
- 1727 Postigo, C., López de Alda, M.J., Barceló, D., 2010. Drugs of abuse and their metabolites in the
1728 Ebro River basin: Occurrence in sewage and surface water, sewage treatment plants removal
1729 efficiency, and collective drug usage estimation. *Environ. Int.* 36, 75–84.
1730 doi:10.1016/j.envint.2009.10.004
- 1731 Pozo, O.J., Ibañez, M., Sancho, J. V., Lahoz-Beneytez, J., Farre, M., Papaseit, E., de la Torre, R.,
1732 Hernandez, F., 2014. Mass Spectrometric Evaluation of Mephedrone In Vivo Human
1733 Metabolism: Identification of Phase I and Phase II Metabolites, Including a Novel Succinyl
1734 Conjugate. *Drug Metab. Dispos.* 43, 248–257. doi:10.1124/dmd.114.061416
- 1735 Preston, K.L., Epstein, D.H., Davoudzadeh, D., Huestis, M. a, 2003. Methadone and metabolite
1736 urine concentrations in patients maintained on methadone. *J. Anal. Toxicol.* 27, 332–341.
1737 doi:10.1093/jat/27.6.332
- 1738 Prosser, J.M., Nelson, L.S., 2012. The Toxicology of Bath Salts: A Review of Synthetic
1739 Cathinones. *J. Med. Toxicol.* 8, 33–42. doi:10.1007/s13181-011-0193-z
- 1740 Prüfer, K., Racimo, F., Patterson, N., Jay, F., Sankararaman, S., Sawyer, S., Heinze, A., Renaud, G.,
1741 Sudmant, P.H., de Filippo, C., Li, H., Mallick, S., Dannemann, M., Fu, Q., Kircher, M.,
1742 Kuhlwilm, M., Lachmann, M., Meyer, M., Ongyerth, M., Siebauer, M., Theunert, C., Tandon,
1743 A., Moorjani, P., Pickrell, J., Mullikin, J.C., Vohr, S.H., Green, R.E., Hellmann, I., Johnson,
1744 P.L.F., Blanche, H., Cann, H., Kitzman, J.O., Shendure, J., Eichler, E.E., Lein, E.S., Bakken,
1745 T.E., Golovanova, L. V, Doronichev, V.B., Shunkov, M. V, Derevianko, A.P., Viola, B.,
1746 Slatkin, M., Reich, D., Kelso, J., Pääbo, S., 2014. The complete genome sequence of a
1747 Neanderthal from the Altai Mountains. *Nature* 505, 43–9. doi:10.1038/nature12886
- 1748 Racamonde, I., Quintana, J.B., Rodil, R., Cela, R., 2015. Application of polypropylene tubes as
1749 single-use and low-cost sorptive extraction materials for the determination of benzodiazepines
1750 and zolpidem in water samples. *Microchem. J.* 119, 58–65. doi:10.1016/j.microc.2014.10.011
- 1751 Racamonde, I., Rodil, R., Quintana, J.B., Villaverde-de-Sáa, E., Cela, R., 2014. Determination of
1752 benzodiazepines, related pharmaceuticals and metabolites in water by solid-phase extraction
1753 and liquid-chromatography-tandem mass spectrometry. *J. Chromatogr. A* 1352, 69–79.
1754 doi:10.1016/j.chroma.2014.05.064
- 1755 Ralla, B., Stephan, C., Meller, S., Dietrich, D., Kristiansen, G., Jung, K., 2014. Nucleic acid-based
1756 biomarkers in body fluids of patients with urologic malignancies. *Crit. Rev. Clin. Lab. Sci.*

1757 8363, 1–32. doi:10.3109/10408363.2014.914888

1758 Ramin, P., Brock, A.L., Polesel, F., Causanilles, A., Emke, E., Voogt, P. De, Plósz, B.G., 2016.

1759 Transformation and sorption of illicit drug biomarkers in sewer systems: understanding the

1760 role of suspended solids in raw wastewater. *Environ. Sci. Technol.*

1761 doi:10.1021/acs.est.6b03049

1762 Ramos, S., Homem, V., Alves, A., Santos, L., 2016. A review of organic UV-filters in wastewater

1763 treatment plants. *Environ. Int.* 86, 24–44. doi:10.1016/j.envint.2015.10.004

1764 Reid, M.J., Baz-Lomba, J.A., Ryu, Y., Thomas, K. V., 2014a. Using biomarkers in wastewater to

1765 monitor community drug use: A conceptual approach for dealing with new psychoactive

1766 substances. *Sci. Total Environ.* 487, 651–658. doi:10.1016/j.scitotenv.2013.12.057

1767 Reid, M.J., Derry, L., Thomas, K. V., 2014b. Analysis of new classes of recreational drugs in

1768 sewage: Synthetic cannabinoids and amphetamine-like substances. *Drug Test. Anal.* 6, 72–79.

1769 doi:10.1002/dta.1461

1770 Reid, M.J., Langford, K.H., Mørland, J., Thomas, K. V., 2011. Analysis and interpretation of

1771 specific ethanol metabolites, ethyl sulfate, and ethyl glucuronide in sewage effluent for the

1772 quantitative measurement of regional alcohol consumption. *Alcohol. Clin. Exp. Res.* 35, 1593–

1773 1599. doi:10.1111/j.1530-0277.2011.01505.x

1774 Reid, M.J., Thomas, K. V., 2016. Chapter 4: New psychoactive substances: analysis and site-

1775 specific testing, in: *Assessing Illicit Drugs in Wastewater: Advances in Wastewater-Based*

1776 *Drug Epidemiology, EMCDDA Insights 22.* pp. 57–65.

1777 Renwick, A.G., 1985. The fate of intense sweeteners in the body. *Food Chem.* 16, 281–301.

1778 doi:10.1016/0308-8146(85)90122-0

1779 Renwick, A.G., Thompson, J.P., O’Shaughnessy, M., Walter, E.J., 2004. The metabolism of

1780 cyclamate to cyclohexylamine in humans during long-term administration. *Toxicol. Appl.*

1781 *Pharmacol.* 196, 367–380. doi:10.1016/j.taap.2004.01.013

1782 Rice, J., Yang, Z., Kasprzyk-Hordern, B., web-support@bath.ac.uk, 2015. Wastewater proteomics

1783 in community-wide molecular diagnostics of public health, in: *Testing the Waters 2015: 2nd*

1784 *International Conference on Wastewater-Based Drug Epidemiology.* University of Bath,

1785 Ascona.

1786 Rico, M., Andrés-Costa, M.J., Picó, Y., 2016. Estimating population size in wastewater-based

1787 epidemiology. Valencia metropolitan area as a case study. *J. Hazard. Mater.*

1788 doi:10.1016/j.jhazmat.2016.05.079

1789 Rifai, N., Gillette, M.A., Carr, S.A., 2006. Protein biomarker discovery and validation: the long and

1790 uncertain path to clinical utility. *Nat. Biotechnol.* 24, 971–983. doi:10.1038/nbt1235

1791 Rizzati, V., Briand, O., Guillou, H., Gamet-Payrastré, L., 2016. Effects of pesticide mixtures in

1792 human and animal models: An update of the recent literature. *Chem. Biol. Interact.* 254, 231–

1793 246. doi:10.1016/j.cbi.2016.06.003

1794 Roberts, A., Renwick, A.G., Sims, J., Snodin, D.J., 2000. Sucralose metabolism and

1795 pharmacokinetics in man. *Food Chem. Toxicol.* 38, 31–41. doi:10.1016/S0278-

1796 6915(00)00026-0

1797 Rodil, R., Quintana, J.B., Concha-Graña, E., López-Mahía, P., Muniategui-Lorenzo, S., Prada-

1798 Rodríguez, D., 2012. Emerging pollutants in sewage, surface and drinking water in Galicia

1799 (NW Spain). *Chemosphere* 86, 1040–1049. doi:10.1016/j.chemosphere.2011.11.053

1800 Rodríguez-Álvarez, T., Racamonde, I., González-Mariño, I., Borsotti, A., Rodil, R., Rodríguez, I.,

1801 Zuccato, E., Quintana, J.B., Castiglioni, S., 2015. Alcohol and cocaine co-consumption in two

1802 European cities assessed by wastewater analysis. *Sci. Total Environ.* 536, 91–8.

1803 doi:10.1016/j.scitotenv.2015.07.016

1804 Rodríguez-Álvarez, T., Rodil, R., Cela, R., Quintana, J.B., 2014a. Ion-pair reversed-phase liquid

1805 chromatography-quadrupole-time-of-flight and triple-quadrupole-mass spectrometry

- 1806 determination of ethyl sulfate in wastewater for alcohol consumption tracing. *J. Chromatogr. A*
1807 1328, 35–42. doi:10.1016/j.chroma.2013.12.076
- 1808 Rodríguez-Álvarez, T., Rodil, R., Rico, M., Cela, R., Quintana, J.B., 2014b. Assessment of Local
1809 Tobacco Consumption by Liquid Chromatography–Tandem Mass Spectrometry Sewage
1810 Analysis of Nicotine and Its Metabolites, Cotinine and trans-3'-Hydroxycotinine, after
1811 Enzymatic Deconjugation. *Anal. Chem.* 86, 10274–10281. doi:10.1021/ac503330c
- 1812 Rosal, R., Rodruíguez, A., Perdigón-Melón, J.A., Petre, A., García-Calvo, E., Gómez, M.J., Agüera,
1813 A., Fernández-Alba, Fernández-Alba, A.R., 2010. Occurrence of emerging pollutants in urban
1814 wastewater and their removal through biological treatment followed by ozonation. *Water Res.*
1815 44, 578–588. doi:10.1016/j.watres.2009.07.004
- 1816 Roth, B.L., Gibbons, S., Arunotayanun, W., Huang, X.P., Setola, V., Treble, R., Iversen, L., 2013.
1817 The Ketamine Analogue Methoxetamine and 3- and 4-Methoxy Analogues of Phencyclidine
1818 Are High Affinity and Selective Ligands for the Glutamate NMDA Receptor. *PLoS One* 8, 2–
1819 6. doi:10.1371/journal.pone.0059334
- 1820 Rousis, N.I., Zuccato, E., Castiglioni, S., 2016a. Monitoring population exposure to pesticides
1821 based on liquid chromatography-tandem mass spectrometry measurement of their urinary
1822 metabolites in urban wastewater: a novel biomonitoring approach. *Sci. Total Environ.* 571,
1823 1349–1357. doi:10.1016/j.scitotenv.2016.07.036
- 1824 Rousis, N.I., Zuccato, E., Castiglioni, S., 2016b. Wastewater-based epidemiology to assess human
1825 exposure to pyrethroid pesticides. *Environ. Int.* doi:10.1016/j.envint.2016.11.020
- 1826 Ruhoy, I.S., Daughton, C.G., 2008. Beyond the medicine cabinet: An analysis of where and why
1827 medications accumulate. *Environ. Int.* 34, 1157–1169. doi:10.1016/j.envint.2008.05.002
- 1828 Ryan, D., Robards, K., Prenzler, P.D., Kendall, M., 2011. Recent and potential developments in the
1829 analysis of urine: A review. *Anal. Chim. Acta* 684, 8–20. doi:10.1016/j.aca.2010.10.035
- 1830 Rydevik, A., Lopardo, L., Petrie, B., Kasprzyk-Hordern, B., 2015. Wastewater profiling for
1831 community-wide human exposure assessment from environmental endocrine disrupting
1832 chemicals in personal care products, in: *Testing the Waters 2015: 2nd International Conference*
1833 *on Wastewater-Based Drug Epidemiology, 2015-10-11 - 2015-10-15, Ascona.*
- 1834 Ryoo, S., Lee, J., Yeo, J., Na, H.-K., Kim, Y.-K., Jang, H., Lee, J.H., Han, S.W., Lee, Y., Kim,
1835 V.N., Min, D.-H., 2013. Quantitative and Multiplexed MicroRNA Sensing in Living Cells
1836 Based on Peptide Nucleic Acid and Nano Graphene Oxide (PANGO). *ACS Nano* 7, 5882–
1837 5891.
- 1838 Ryu, Y., Barceló, D., Barron, L.P., Bijlsma, L., Castiglioni, S., de Voogt, P., Emke, E., Hernández,
1839 F., Lai, F.Y., Lopes, A., de Alda, M.L., Mastroianni, N., Munro, K., O'Brien, J., Ort, C., Plósz,
1840 B.G., Reid, M.J., Yargeau, V., Thomas, K. V, 2016. Comparative measurement and
1841 quantitative risk assessment of alcohol consumption through wastewater-based epidemiology:
1842 An international study in 20 cities. *Sci. Total Environ.* 565, 977–983.
1843 doi:10.1016/j.scitotenv.2016.04.138
- 1844 Ryu, Y., Reid, M.J., Thomas, K. V., 2015. Liquid chromatography–high resolution mass
1845 spectrometry with immunoaffinity clean-up for the determination of the oxidative stress
1846 biomarker 8-iso-prostaglandin F2alpha in wastewater. *J. Chromatogr. A* 1409, 146–151.
1847 doi:10.1016/j.chroma.2015.07.060
- 1848 Sánchez-Flores, M., Pásaro, E., Bonassi, S., Laffon, B., Valdiglesias, V., 2015. H2AX assay as
1849 DNA damage biomarker for human population studies: Defining experimental conditions.
1850 *Toxicol. Sci.* 144, 406–413. doi:10.1093/toxsci/kfv011
- 1851 Santos, J.L., Aparicio, I., Callejón, M., Alonso, E., 2009. Occurrence of pharmaceutically active
1852 compounds during 1-year period in wastewaters from four wastewater treatment plants in
1853 Seville (Spain). *J. Hazard. Mater.* 164, 1509–1516. doi:10.1016/j.jhazmat.2008.09.073
- 1854 Sardesai, V.M., Waldshan, T.H., 1991. Natural and synthetic intense sweeteners. *J. Nutr. Biochem.*

- 1855 2, 236–244. doi:10.1016/0955-2863(91)90081-F
- 1856 Schenzel, J., Hungerbühler, K., Bucheli, T.D., 2012. Mycotoxins in the environment: II. Occurrence
1857 and origin in Swiss river waters. *Environ. Sci. Technol.* 46, 13076–13084.
1858 doi:10.1021/es301558v
- 1859 Schenzel, J., Schwarzenbach, R.P., Bucheli, T.D., 2010. Multi-residue screening method to quantify
1860 mycotoxins in aqueous environmental samples. *J. Agric. Food Chem.* 58, 11207–11217.
1861 doi:10.1021/jf102737q
- 1862 Schneider, H., Glatt, H., 2004. Sulpho-conjugation of ethanol in humans in vivo and by individual
1863 sulphotransferase forms in vitro. *Biochem. J.* 383, 543–549. doi:10.1042/BJ20040925
- 1864 Schreder, E.D., La Guardia, M.J., 2014. Flame retardant transfers from U.S. households (dust and
1865 laundry wastewater) to the aquatic environment. *Environ. Sci. Technol.* 48, 11575–11583.
1866 doi:10.1021/es502227h
- 1867 Schwartz, R., Milteer, R., LeBeau, M.A., 2000. Drug-facilitated sexual assault (“Date Rape”).
1868 *South. Med. J.* 93, 558–561.
- 1869 Seely, K.A., Lapoint, J., Moran, J.H., Fattore, L., 2012. Spice drugs are more than harmless herbal
1870 blends: A review of the pharmacology and toxicology of synthetic cannabinoids. *Prog. Neuro-
1871 Psychopharmacology Biol. Psychiatry* 39, 234–243. doi:10.1016/j.pnpbp.2012.04.017
- 1872 Senta, I., Gracia-Lor, E., Borsotti, A., Zuccato, E., Castiglioni, S., 2015a. Wastewater analysis to
1873 monitor use of caffeine and nicotine and evaluation of their metabolites as biomarkers for
1874 population size assessment. *Water Res.* 74, 23–33. doi:10.1016/j.watres.2015.02.002
- 1875 Senta, I., Krizman, I., Ahel, M., Terzic, S., 2015b. Multiresidual analysis of emerging
1876 amphetamine-like psychoactive substances in wastewater and river water. *J. Chromatogr. A*
1877 1425, 204–212. doi:10.1016/j.chroma.2015.11.043
- 1878 Senta, I., Krizman, I., Ahel, M., Terzic, S., 2014. Assessment of stability of drug biomarkers in
1879 municipal wastewater as a factor influencing the estimation of drug consumption using sewage
1880 epidemiology. *Sci. Total Environ.* 487, 659–665. doi:10.1016/j.scitotenv.2013.12.054
- 1881 Sharma, A., Schorr, U., Thiede, H., Distler, A., 1993. Effect of dietary salt restriction on urinary
1882 serotonin and 5-hydroxyindoleacetic acid excretion in man. *J Hypertens* 1381–1386.
- 1883 Shima, N., Katagi, M., Kamata, H., Matsuta, S., Sasaki, K., Kamata, T., Nishioka, H., Miki, A.,
1884 Tatsuno, M., Zaitzu, K., Ishii, A., Sato, T., Tsuchihashi, H., Suzuki, K., 2014. Metabolism of
1885 the newly encountered designer drug α -pyrrolidinovalerophenone in humans: Identification
1886 and quantitation of urinary metabolites. *Forensic Toxicol.* 32, 59–67. doi:10.1007/s11419-013-
1887 0202-9
- 1888 Silva, M.J., Samandar, E., Reidy, J.A., Hauser, R., Needham, L.L., Calafat, A.M., 2007. Metabolite
1889 profiles of Di-n-butyl phthalate in humans and rats. *Environ. Sci. Technol.* 41, 7576–7580.
1890 doi:10.1021/es071142x
- 1891 Singh, S.P., Azua, A., Chaudhary, A., Khan, S., Willett, K.L., Gardinali, P.R., 2010. Occurrence
1892 and distribution of steroids, hormones and selected pharmaceuticals in South Florida coastal
1893 environments. *Ecotoxicology* 19, 338–350. doi:10.1007/s10646-009-0416-0
- 1894 Singh, S.P., Gardinali, P.R., 2006. Trace determination of 1-aminopropanone, a potential marker for
1895 wastewater contamination by liquid chromatography and atmospheric pressure chemical
1896 ionization-mass spectrometry. *Water Res.* 40, 588–594. doi:10.1016/j.watres.2005.11.036
- 1897 Smith, H.S., 2009. Opioid metabolism. *Mayo Clin. Proc.* 84, 613–624. doi:10.4065/84.7.613
- 1898 Snip, L.J.P., Flores-Alsina, X., Aymerich, I., Rodríguez-Mozaz, S., Barceló, D., Plósz, B.G.,
1899 Corominas, L., Rodríguez-Roda, I., Jeppsson, U., Gernaey, K. V., 2016. Generation of
1900 synthetic influent data to perform (micro)pollutant wastewater treatment modelling studies.
1901 *Sci. Total Environ.* 569–570, 278–290. doi:10.1016/j.scitotenv.2016.05.012
- 1902 Staack, R.F., Fehn, J., Maurer, H.H., 2003. New designer drug p-methoxymethamphetamine:
1903 Studies on its metabolism and toxicological detection in urine using gas chromatography-mass

1904 spectrometry. *J. Chromatogr. B Anal. Technol. Biomed. Life Sci.* 789, 27–41.
1905 doi:10.1016/S1570-0232(02)01018-8

1906 Staack, R.F., Fritschi, G., Maurer, H.H., 2001. GC-MS studies on the metabolism and on the
1907 toxicological analysis of the new piperazine-like designer drugs BZP, MDBP, TFMPP, mCPP,
1908 MeOPP, Proceedings of the 39th International TIAFT Meeting in Prague, Czech Republic.

1909 Staack, R.F., Maurer, H.H., 2005. Metabolism of designer drugs of abuse. *Curr Drug Metab* 6, 259–
1910 274. doi:10.2174/1389200054021825

1911 Staub, C., Marset, M., Mino, a, Mangin, P., 2001. Detection of acetylcodeine in urine as an
1912 indicator of illicit heroin use: method validation and results of a pilot study. *Clin. Chem.* 47,
1913 301–7.

1914 Takada, H., Eganhouse, R., 1998. Molecular markers of anthropogenic waste, in: Meyers, R. A. Ed.
1915 *Encyclopedia of Environmental Analysis and Remediation*. John Wiley & Sons, Inc., New
1916 York, NY, USA. John Wiley & Sons, Inc. pp. 2883–2940.

1917 Thai, P.K., Jiang, G., Gernjak, W., Yuan, Z., Lai, F.Y., Mueller, J.F., 2014. Effects of sewer
1918 conditions on the degradation of selected illicit drug residues in wastewater. *Water Res.* 48,
1919 538–547. doi:10.1016/j.watres.2013.10.019

1920 Thai, P.K., Lai, F.Y., Edirisinghe, M., Hall, W., Bruno, R., O’Brien, J.W., Prichard, J., Kirkbride,
1921 K.P., Mueller, J.F., 2016. Monitoring temporal changes in use of two cathinones in a large
1922 urban catchment in Queensland, Australia. *Sci. Total Environ.* 545–546, 250–255.
1923 doi:10.1016/j.scitotenv.2015.12.038

1924 Thomaidis, N.S., Gago-Ferrero, P., Ort, C., Maragou, N.C., Alygizakis, N.A., Borova, V.L.,
1925 Dasenaki, M.E., 2016. Reflection of Socioeconomic Changes in Wastewater: Licit and Illicit
1926 Drug Use Patterns. *Environ. Sci. Technol.* acs.est.6b02417. doi:10.1021/acs.est.6b02417

1927 Thomas, K. V., Bijlsma, L., Castiglioni, S., Covaci, A., Emke, E., Grabic, R., Hernández, F.,
1928 Karolak, S., Kasprzyk-Hordern, B., Lindberg, R.H., Lopez de Alda, M., Meierjohann, A., Ort,
1929 C., Pico, Y., Quintana, J.B., Reid, M., Rieckermann, J., Terzic, S., van Nuijs, A.L.N., de
1930 Voogt, P., 2012. Comparing illicit drug use in 19 European cities through sewage analysis. *Sci.*
1931 *Total Environ.* 432, 432–439. doi:10.1016/j.scitotenv.2012.06.069

1932 Thomas, K. V., Reid, M.J., 2011. What Else Can the Analysis of Sewage for Urinary Biomarkers
1933 Reveal About Communities? *Environ. Sci. Technol.* 45, 7611–7612. doi:10.1021/es202522d

1934 Thomsen, P.F., Willerslev, E., 2015. Environmental DNA - An emerging tool in conservation for
1935 monitoring past and present biodiversity. *Biol Conserv* 183, 4–18.

1936 Tipirissetti, N.R., Govatati, S., Pullari, P., Malempati, S., Thupurani, M.K., Perugu, S., Guruvaiah,
1937 P., Rao K, L., Digumarti, R.R., Nallanchakravarthula, V., Bhanoori, M., Satti, V., 2014.
1938 Mitochondrial control region alterations and breast cancer risk: A study in south Indian
1939 population. *PLoS One* 9, 1–8. doi:10.1371/journal.pone.0085363

1940 Toxicological profile for di(2-ethylhexyl)phthalate (DEHP), 2002. . U.S Dep. Heal. Hum. Serv.
1941 Public Heal. Serv. Agency Toxic Subst. Dis. Regist.

1942 Toxicological profile for Di-n-Phthalate, 2001. . U.S Dep. Heal. Hum. Serv. Public Heal. Serv.
1943 Agency Toxic Subst. Dis. Regist.

1944 Tran, N.H., Hu, J., Ong, S.L., 2013. Simultaneous determination of PPCPs, EDCs, and artificial
1945 sweeteners in environmental water samples using a single-step SPE coupled with HPLC-
1946 MS/MS and isotope dilution. *Talanta* 113, 82–92. doi:10.1016/j.talanta.2013.03.072

1947 Trunnelle, K.J., Bennett, D.H., Tancredi, D.J., Gee, S.J., Stoecklin-Marois, M.T., Hennessy-Burt,
1948 T.E., Hammock, B.D., Schenker, M.B., 2013. Pyrethroids in house dust from the homes of
1949 farm worker families in the MICASA study. *Environ. Int.* 61, 57–63.
1950 doi:10.1016/j.envint.2013.09.007

1951 Tscharke, B.J., Chen, C., Gerber, J.P., White, J.M., 2016. Temporal trends in drug use in Adelaide,
1952 South Australia by wastewater analysis. *Sci. Total Environ.* 565, 384–391.

- 1953 doi:10.1016/j.scitotenv.2016.04.183
- 1954 Tsui, M.M.P., Leung, H.W., Lam, P.K.S., Murphy, M.B., 2014. Seasonal occurrence, removal
1955 efficiencies and preliminary risk assessment of multiple classes of organic UV filters in
1956 wastewater treatment plants. *Water Res.* 53, 58–67. doi:10.1016/j.watres.2014.01.014
- 1957 Tsujikawa, K., Mikuma, T., Kuwayama, K., Miyaguchi, H., Kanamori, T., Iwata, Y.T., Inoue, H.,
1958 2012. Degradation pathways of 4-methylmethcathinone in alkaline solution and stability of
1959 methcathinone analogs in various pH solutions. *Forensic Sci. Int.* 220, 103–110.
1960 doi:10.1016/j.forsciint.2012.02.005
- 1961 Tsutsumi, H., Katagi, M., Miki, A., Shima, N., Kamata, T., Nishikawa, M., Nakajima, K.,
1962 Tsuchihashi, H., 2005. Development of simultaneous gas chromatography-mass spectrometric
1963 and liquid chromatography-electrospray ionization mass spectrometric determination method
1964 for the new designer drugs, N-benzylpiperazine (BZP), 1-(3-trifluoromethylphenyl)piperazine
1965 (TFMPP). *J. Chromatogr. B Anal. Technol. Biomed. Life Sci.* 819, 315–322.
1966 doi:10.1016/j.jchromb.2005.02.016
- 1967 Turner, N.W., Bramhmbhatt, H., Szabo-Vezse, M., Poma, A., Coker, R., Piletsky, S.A., 2015.
1968 Analytical methods for determination of mycotoxins: An update (2009-2014). *Anal. Chim.*
1969 *Acta* 901, 12–33. doi:10.1016/j.aca.2015.10.013
- 1970 Udert, K.M., Larsen, T.A., Gujer, W., 2006. Fate of major compounds in source-separated urine.
1971 *Water Sci. Technol.* 54, 413–420. doi:10.2166/wst.2006.921
- 1972 UNODC, U.N.O. on D. and C., 2015. World Drug Report. doi:10.1017/CBO9781107415324.004
- 1973 Uralets, V., Rana, S., Morgan, S., Ross, W., 2014. Testing for designer stimulants: Metabolic
1974 profiles of 16 synthetic cathinones excreted free in human urine. *J. Anal. Toxicol.* 38, 233–
1975 241. doi:10.1093/jat/bku021
- 1976 USFDA, 2006. Artificial sweeteners: No calories ... sweet! FDA Consum, The US Food and Drug
1977 Administration (USFDA). USFDA 40, 27–28.
- 1978 Valle-Sistac, J., Molins-Delgado, D., Díaz, M., Ibáñez, L., Barceló, D., Silvia Díaz-Cruz, M., 2016.
1979 Determination of parabens and benzophenone-type UV filters in human placenta: First
1980 description of the existence of benzyl paraben and benzophenone-4. *Environ. Int.* 88, 243–249.
1981 doi:10.1016/j.envint.2015.12.034
- 1982 Valls, M., Bayona, J.M., Albaigés, J., 1989. Use of trialkylamines as an indicator of urban sewage
1983 in sludges, coastal waters and sediments. *Nature* 337, 722–724.
- 1984 Van den Eede, N., Heffernan, A.L., Aylward, L.L., Hobson, P., Neels, H., Mueller, J.F., Covaci, A.,
1985 2015. Age as a determinant of phosphate flame retardant exposure of the Australian population
1986 and identification of novel urinary PFR metabolites. *Environ. Int.* 74, 1–8.
1987 doi:10.1016/j.envint.2014.09.005
- 1988 Van den Eede, N., Maho, W., Erratico, C., Neels, H., Covaci, A., 2013. First insights in the
1989 metabolism of phosphate flame retardants and plasticizers using human liver fractions.
1990 *Toxicol. Lett.* 223, 9–15. doi:10.1016/j.toxlet.2013.08.012
- 1991 van Nuijs, A.L., Gheorghe, A., Jorens, P.G., Maudens, K., Neels, H., Covaci, A., 2014.
1992 Optimization, validation, and the application of liquid chromatography-tandem mass
1993 spectrometry for the analysis of new drugs of abuse in wastewater. *Drug Test. Anal.* 6, 861–
1994 867.
- 1995 van Nuijs, A.L.N., Castiglioni, S., Tarcomnicu, I., Postigo, C., de Alda, M.L., Neels, H., Zuccato,
1996 E., Barcelo, D., Covaci, A., 2011a. Illicit drug consumption estimations derived from
1997 wastewater analysis: A critical review. *Sci. Total Environ.* 409, 3564–3577.
1998 doi:10.1016/j.scitotenv.2010.05.030
- 1999 van Nuijs, A.L.N., Covaci, A., Beyers, H., Bervoets, L., Blust, R., Verpooten, G., Neels, H., Jorens,
2000 P.G., 2015. Do concentrations of pharmaceuticals in sewage reflect prescription figures?
2001 *Environ. Sci. Pollut. Res.* 22, 9110–9118. doi:10.1007/s11356-014-4066-2

2002 van Nuijs, A.L.N., Mougel, J.F., Tarcomnicu, I., Bervoets, L., Blust, R., Jorens, P.G., Neels, H.,
2003 Covaci, A., 2011b. Sewage epidemiology - A real-time approach to estimate the consumption
2004 of illicit drugs in Brussels, Belgium. *Environ. Int.* 37, 612–621.
2005 doi:10.1016/j.envint.2010.12.006

2006 Van Nuijs, A.L.N., Pecceu, B., Theunis, L., Dubois, N., Charlier, C., Jorens, P.G., Bervoets, L.,
2007 Blust, R., Meulemans, H., Neels, H., Covaci, A., 2009. Can cocaine use be evaluated through
2008 analysis of wastewater? A nation-wide approach conducted in Belgium. *Addiction* 104, 734–
2009 741. doi:10.1111/j.1360-0443.2009.02523.x

2010 Vazquez-Roig, P., Kasprzyk-Hordern, B., Blasco, C., Pico, Y., 2014. Stereoisomeric profiling of
2011 drugs of abuse and pharmaceuticals in wastewaters of Valencia (Spain). *Sci. Total Environ.*
2012 494–495, 49–57. doi:10.1016/j.scitotenv.2014.06.098

2013 Verlicchi, P., Al Aukidy, M., Zambello, E., 2012. Occurrence of pharmaceutical compounds in
2014 urban wastewater: Removal, mass load and environmental risk after a secondary treatment-A
2015 review. *Sci. Total Environ.* 429, 123–155. doi:10.1016/j.scitotenv.2012.04.028

2016 Wang, C., Yang, L., Wang, S., Zhang, Z., Yu, Y., Wang, M., Cromie, M., Gao, W., Wang, S.-L.,
2017 2016. The classic EDCs, phthalate esters and organochlorines, in relation to abnormal sperm
2018 quality: a systematic review with meta-analysis. *Sci. Rep.* 6, 19982. doi:10.1038/srep19982

2019 Wang, P., Wu, H., Dai, Z., Zou, X., 2012. Picomolar level profiling of the methylation status of the
2020 p53 tumor suppressor gene by a label-free electrochemical biosensor. *Chem. Commun.* 48,
2021 10754. doi:10.1039/c2cc35615e

2022 Warth, B., Sulyok, M., Krska, R., 2013. LC-MS/MS-based multibiomarker approaches for the
2023 assessment of human exposure to mycotoxins. *Anal. Bioanal. Chem.* 405, 5687–5695.
2024 doi:10.1007/s00216-013-7011-1

2025 Webb, A.L., Kruczkiewicz, P., Selinger, L.B., Inglis, G.D., Taboada, E.N., 2015. Development of a
2026 comparative genomic fingerprinting assay for rapid and high resolution genotyping of
2027 *Arcobacter butzleri*. *BMC Microbiol* 15, 94. doi:10.1186/s12866-015-0426-4

2028 Welter-Luedeke, J., Maurer, H.H., 2015. New Psychoactive Substances: Chemistry, Pharmacology,
2029 Metabolism, and Detectability of Amphetamine Derivatives with Modified Ring Systems.
2030 *Ther. Drug Monit.* 38, 4–11. doi:10.1097/FTD.0000000000000240

2031 Wettstein, F.E., Bucheli, T.D., 2010. Poor elimination rates in waste water treatment plants lead to
2032 continuous emission of deoxynivalenol into the aquatic environment. *Water Res.* 44, 4137–
2033 4142. doi:10.1016/j.watres.2010.05.038

2034 WHO Food Additive Series No. 52, 2004. Safety evaluation of certain food additives and
2035 contaminants.

2036 Wohlfarth, A., Scheidweiler, K.B., Chen, X., Liu, H., Huestis, M.A., 2013. Qualitative
2037 Confirmation of 9 Synthetic Cannabinoids and 20 Metabolites in Human Urine Using
2038 LC–MS/MS and Library Search. *Anal. Chem.* 85, 3730–3738.

2039 World Health Organization, 2015. WHO global report on trends in prevalence of tobacco smoking.

2040 World Health Organization, 2014. WHO global status report on alcohol and health.

2041 Wurst, F.M., Dresen, S., Allen, J.P., Wiesbeck, G., Graf, M., Weinmann, W., 2006. Ethyl sulphate:
2042 A direct ethanol metabolite reflecting recent alcohol consumption. *Addiction* 101, 204–211.
2043 doi:10.1111/j.1360-0443.2005.01245.x

2044 Yang, Z., D’Uriac, M.A., Goggins, S., Kasprzyk-Hordern, B., Thomas, K. V., Frost, C.G., Estrela,
2045 P., 2015a. A novel DNA biosensor using a ferrocenyl intercalator applied to the potential
2046 detection of human population biomarkers in wastewater. *Environ. Sci. Technol.* 49, 5609–
2047 5617. doi:10.1021/acs.est.5b00637

2048 Yang, Z., Kasprzyk-Hordern, B., Frost, C.G., Estrela, P., Thomas, K. V., 2015b. Community
2049 sewage sensors for monitoring public health. *Environ. Sci. Technol.* 49, 5845–5846.
2050 doi:10.1021/acs.est.5b01434

2051 Yang, Z., Kasprzyk-Hordern, B., Goggins, S., Frost, C.G., Estrela, P., 2015c. A novel
2052 immobilization strategy for electrochemical detection of cancer biomarkers: DNA-directed
2053 immobilization of aptamer sensors for sensitive detection of prostate specific antigens. *Analyst*
2054 140, 2628–2633. doi:10.1039/C4AN02277G

2055 Yuan, S.-F., Liu, Z.-H., Huang, R.-P., Yin, H., Dang, Z., 2015. Levels of six antibiotics used in
2056 China estimated by means of wastewater-based epidemiology. *Water Sci. Technol.*
2057 doi:10.2166/wst.2015.526

2058 Yusa, V., Millet, M., Coscolla, C., Roca, M., 2015. Analytical methods for human biomonitoring of
2059 pesticides. A review. *Anal. Chim. Acta* 891, 15–31. doi:10.1016/j.aca.2015.05.032

2060 Zaitsev, K., Katagi, M., Tatsuno, M., Sato, T., Tsuchihashi, H., Suzuki, K., 2011. Recently
2061 abused α -keto derivatives of 3,4- methylenedioxyphenylalkylamines: A review of their
2062 metabolisms and toxicological analysis. *Forensic Toxicol.* 29, 73–84. doi:10.1007/s11419-011-
2063 0111-8

2064 Zawilska, J.B., Andrzejczak, D., 2015. Next generation of novel psychoactive substances on the
2065 horizon—a complex problem to face. *Drug Alcohol Depend.* 157, 1–17.
2066 doi:10.1016/j.drugalcdep.2015.09.030

2067 Zhang, Z., Sun, L., Hu, Y., Jiao, J., Hu, J., 2013. Inverse antagonist activities of parabens on human
2068 oestrogen-related receptor γ (ERR γ): In vitro and in silico studies. *Toxicol. Appl. Pharmacol.*
2069 270, 16–22. doi:10.1016/j.taap.2013.03.030

2070 Zhong, R., Wang, H., Wu, X., Cao, Y., He, Z., He, Y., Liu, J., 2013. In vitro investigation of the
2071 effect of plasticizers on the blood compatibility of medical grade plasticized poly (vinyl
2072 chloride). *J. Mater. Sci. Mater. Med.* 24, 1985–1992. doi:10.1007/s10856-013-4950-1

2073 Zhou, N., Lin, X., Wang, S., Wang, H., Li, W., Tao, Z., Xu, A., 2014. Environmental Surveillance
2074 for Human Astrovirus in Shandong Province, China in 2013. *Sci. Rep.* 4, 7539.
2075 doi:10.1038/srep07539

2076 Zolfaghari, M., Drogui, P., Seyhi, B., Brar, S.K., Buelna, G., Dubé, R., 2014. Occurrence, fate and
2077 effects of Di (2-ethylhexyl) Phthalate in wastewater treatment plants: a review. *Environ.*
2078 *Pollut.* 194, 281–93. doi:10.1016/j.envpol.2014.07.014

2079 Zuccato, E., Castiglioni, S., Senta, I., Borsotti, A., Genetti, B., Andreotti, A., Pieretti, G.,
2080 Serpelloni, G., 2016. Population surveys compared with wastewater analysis for monitoring
2081 illicit drug consumption in Italy in 2010-2014. *Drug Alcohol Depend.* 161, 178–188.
2082 doi:10.1016/j.drugalcdep.2016.02.003

2083 Zuccato, E., Chiabrando, C., Castiglioni, S., Bagnati, R., Fanelli, R., 2008. Estimating community
2084 drug abuse by wastewater analysis. *Environ. Health Perspect.* 116, 1027–1032.
2085 doi:10.1289/ehp.11022

2086 Zuccato, E., Chiabrando, C., Castiglioni, S., Calamari, D., Bagnati, R., Schiarea, S., Fanelli, R.,
2087 2005. Cocaine in surface waters: a new evidence-based tool to monitor community drug abuse.
2088 *Environ. Heal. A Glob. Access Sci. Source* 4, 14. doi:10.1186/Received

2089 Zuetenhorst, J.M., 2004. Daily Cyclic Changes in the Urinary Excretion of 5-Hydroxyindoleacetic
2090 Acid in Patients with Carcinoid Tumors. *Clin. Chem.* 50, 1634–1639.
2091 doi:10.1373/clinchem.2004.032151
2092